

Copyright Undertaking

This thesis is protected by copyright, with all rights reserved.

By reading and using the thesis, the reader understands and agrees to the following terms:

- 1. The reader will abide by the rules and legal ordinances governing copyright regarding the use of the thesis.
- 2. The reader will use the thesis for the purpose of research or private study only and not for distribution or further reproduction or any other purpose.
- 3. The reader agrees to indemnify and hold the University harmless from and against any loss, damage, cost, liability or expenses arising from copyright infringement or unauthorized usage.

IMPORTANT

If you have reasons to believe that any materials in this thesis are deemed not suitable to be distributed in this form, or a copyright owner having difficulty with the material being included in our database, please contact lbsys@polyu.edu.hk providing details. The Library will look into your claim and consider taking remedial action upon receipt of the written requests.

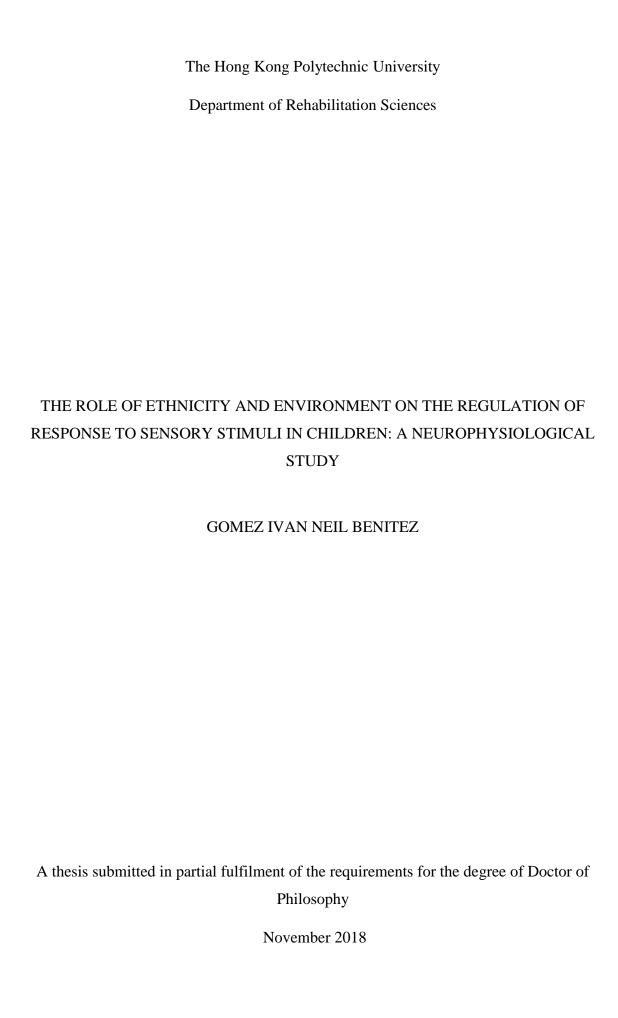
THE ROLE OF ETHNICITY AND ENVIRONMENT ON THE REGULATION OF RESPONSE TO SENSORY STIMULI IN CHILDREN: A NEUROPHYSIOLOGICAL STUDY

GOMEZ IVAN NEIL BENITEZ

PhD

The Hong Kong Polytechnic University

2019



CERTIFICATE OF ORIGINALITY

I hereby declare that this thesis is my own work and that, to the best of my knowledge and
belief, it reproduces no material previously published or written, nor material that has been
accepted for the award of any other degree or diploma, except where due acknowledgement
has been made in the text.

	(Signed)
Ivan Neil B. Gomez	(Name of Student)

DEDICATION

This thesis is dedicated to:

God Almighty my creator, my strong pillar, my source of inspiration, wisdom, knowledge and understanding;

My beloved family, Mommy and *Ate*, whose encouragement has made sure that I give it all it takes to finish that which I have started;

My father and grandparents, I hope you could have witnessed how far I have gone in life. Your souls flow through me and will always be remembered;

My friends who believed in me, even when I sometimes stopped believing in myself;

All the people in my life, past and present, who touched my heart and changed me for good;

My eternal gratitude.

i

Abstract

Background. The world is full of sensory stimuli and adaptation to this is necessary for childhood development. Adaptation involves regulation, a response to the demands of the environment through modifications at the neurophysiological level. The activity between parasympathetic and sympathetic branches of the autonomic nervous systems has been suggested to be appropriate measures of neurophysiological regulation. Various factors have been suggested to influence the neurophysiological regulation of response towards sensory stimuli. However, the role of ethnicity and environment has rarely been examined. Ethnicity refers to distinct populations that share biological origins and genetic similarities. On the other hand, the environment is defined as the geographic niche that serves as dwelling place among groups of individuals. Understanding the influence of ethnicity and the environment on the regulations of neurophysiological mechanism in response to external challenges can provide a better understanding of the development of adaptive abilities in children.

Aim. This thesis aims to examine the role of ethnicity and environment of the regulation of response towards sensory stimuli among children using neurophysiological methods. It is hypothesised that children from different ethnicities and environments have significantly different regulation of response to sensory stimuli.

Methods. There are four main hypotheses tested in this thesis that recruited 156 typically developing children ages 7-12 years old from different ethnic groups or environment: geographic (country of habitat) or physical (urban or rural setting) as follows: [1] 31 Chinese children living in Hong Kong (CHK); [2] 28 Filipino children living in Hong Kong (FHK); [3] 52 Filipino children who are living in urban areas (FU); and [4] 43 Filipino children who are living in rural areas (FR) in the Philippines. Heart rate variability (HRV) and electrodermal activity (EDA) were respectively recorded and measured using a Polar H2 heart rate monitor and the eSense GSR skin response sensor. Children were subjected to a sensory laboratory paradigm with a resting, auditory stimulation and recovery conditions. Neurophysiological measures of regulation of response to sensory stimuli between groupwise (CHK, FHK, FU and FR) and different pairwise (CHK and FHK, FHK and FU, CHK and FU, FU and FR) combinations at different conditions were tested using mixed factorial analysis of variance, multivariate analysis of variance and correlation set a p=0.05.

Results. The pattern of neurophysiological regulation of response to sensory stimuli was generally similar across children from different ethnicities and environments. However, pairwise combinations between groups described the influence of ethnicity and environments on differences in the regulation of response to sensory stimuli using neurophysiological measure. The key findings in this thesis are:

- [1] Between children (CHK and FHK groups) with different ethnicities but lives in the same geographic environments (Hong Kong) and environment landscapes (urban setting), there is no significant difference in the LF n.u., high-frequency bands of HRV (HF n.u.) and skin conductance level/response (SCL/SCR) at resting (Λ = 0.01, F(3,11)= 0.20, p= 0.89, d= 0.21), stimulation (Λ = 0.03, F(3,11)= 0.52, p= 0.67, d= 0.34) and recovery (Λ = 0.00, F(3,11)= 0.05, p= 0.99, d= 0.10) conditions.
- [2] Between children (FHK and FU groups) with the same ethnicity (Filipino) but lives in different geographic environments (Hong Kong and Philippines) and similar environment landscapes (urban settings), there is significant difference in the LF n.u., HF n.u. and SCL/SCR at resting (Λ = 0.23, F(3,32)= 7.42, p<0.00, d= 1.08), stimulation (Λ = 0.14, F(3,32)= 4.26, p= 0.01, d= 0.82) and recovery (Λ = 0.10, F(3,32)= 2.712, p= 0.05, d= 0.65) conditions.
- [3] Between children (CHK and FU groups) with different ethnicities living in geographic environments (Hong Kong and Philippines) but similar environment landscapes (urban setting), there is significant difference in the LF n.u., HF n.u. and SCL/SCR) at resting (Λ = 0.17, F(3,38) = 5.60, p < 0.00, d = 0.91), stimulation ($\Lambda = 0.15$, F(3,38) = 4.66, p = 0.01, d = 0.83) and recovery ($\Lambda = .14$, F(3,38) = 4.44, p = 0.001, d = 0.81) conditions.
- [4] Between children (FU and FR groups) from similar ethnicities (Filipinos) and geographic environments (Philippines) living in different physical environments (urban and rural setting) there is significant difference in the LF n.u., HF n.u. and SCL/SCR at resting (Λ = 0.19, F(3,11)= 7.29, p<0.00, d= 0.97), stimulation (Λ = 0.30, F(3,11)= 13.09, p<0.00, d= 1.31) and recovery (Λ = 0.14, F(3,11)= 5.00, p<0.00, d= 0.81) conditions.

Conclusion. In this study, the neurophysiological autonomic activity among four groups of children (CHK, FHK, FU and FR) was examined. This thesis found that in response to sensory stimuli: 1) there are differences in the patterns of change in the SCL among CHK and FHK groups; and there are differences in the levels of autonomic activity between: a) FHK and FU groups; b) CHK and FU groups; and FU and FR group.

To conclude, this thesis provides evidence on the influence of ethnicity and environments on the regulation of response to sensory stimuli using a neurophysiological perspective. The notion that autonomic activity as an underlying neurophysiologic mechanism that enables adaptation is further reinforced. The results of this study have implication on clinical, research and policy studies on the influence of the environment on children's response to sensory stimuli, as well as the implicit influence of migration on children's health, behaviour and well-being.

PUBLICATIONS ARISING DIRECTLY FROM THE THESIS

Gomez, I. N. B., Lai, C. Y. Y., Morato-Espino, P. G., Chan, C. C. H., & Tsang, H. W. H. (2017). Behavioural and Autonomic Regulation of Response to Sensory Stimuli among Children: A Systematic Review of Relationship and Methodology. *BioMed Research International*, 2017. doi:10.1155/2017/2629310.

Gomez, I. N. B., Lai, C. Y. Y., Chan, C. C. H., & Tsang, H. W. H. (2018). The Role of Ethnicity and Environment in the Regulation of Response to Sensory Stimulus in Children: Protocol and Pilot Findings of a Neurophysiological Study. *JMIR Research Protocols*, 7(1). doi:10.2196/resprot.8157.

Gomez, I. N. B., Lai, C. Y. Y., Yung, T. W. K., Chan, C. C. H., & Tsang, H. W. H. (2018). Migration Influences on the Allostatic Load of Children: Systematic Review Protocol. *JMIR Research Protocols*, 7(1). doi:10.2196/resprot.8332.

PRESENTATIONS AT INTERNATIONAL CONFERENCES ARISING FROM THE THESIS

Gomez, I. N. B., Tsang, H. W. H. & Lai, C. Y. Y. (2016, November). *The role of ethnicity and environment on the regulation of response to sensory stimuli in children: a neurophysiological study.* Paper presented at the 10th Pan-Pacific Conference on Rehabilitation, Shanghai, China.

Gomez, I. N. B., Tsang, H. W. H. & Lai, C. Y. Y. (2016, November). A systematic review of literature on the role of genetics and environment on the autonomic activity of children that influences child behaviour. Poster presented at the 10th Pan-Pacific Conference on Rehabilitation, Shanghai, China.

Gomez, I. N. B., Lai, C. Y. Y., Chan, C. C. H. & Tsang, H. W. H. (2017, August). Migration influences autonomic regulation of response to sensory stimuli in children. Poster presented at the International Society for Autonomic Neuroscience 2017 Conference, Nagoya, Japan.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2017, October). *Behavioural and Autonomic Regulation of Response to Sensory Stimuli among Children: A Systematic Review of Relationship and Methodology*. Paper presented at the 1st Asia-Pacific Occupational Therapy Symposium, Chang Gung University, Taoyuan City, Taiwan.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2017, October). *The relationship between behavioural and autonomic regulation of response to sensory stimuli in children*. Paper presented at the 1st Asia-Pacific Occupational Therapy Symposium, Chang Gung University, Taoyuan City, Taiwan.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2017, October). *Environmental influence on the autonomic regulation of response towards sensory stimuli in children*. Paper presented at the 1st Asia-Pacific Occupational Therapy Symposium, Chang Gung University, Taoyuan City, Taiwan.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2018, November). *Behavioural and physiological adaptation in children of migrant origins*. Poster presented at the 11th Pan-Pacific Conference on Rehabilitation, The Hong Kong Polytechnic University, Hong Kong SAR.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2018, November). *Parasympathetic resting state differences between children with and without ADHD*. Paper presented at the 11th Pan-Pacific Conference on Rehabilitation, The Hong Kong Polytechnic University, Hong Kong SAR.

Gomez, I. N. B., Lai, C. Y. Y., & Tsang, H. W. H. (2018, November). *Utilising neuroscience evidence to inform research: a reflective inquiry*. Poster presented at the 11th Pan-Pacific Conference on Rehabilitation, The Hong Kong Polytechnic University, Hong Kong SAR.

Acknowledgements

Firstly, I would like to express my sincere gratitude to my chief supervisor Dr. Cynthia Lai and co-supervisor Prof. Hector Tsang for their continuous support of my PhD study and related research, for their patience, motivation, and immense knowledge. Their guidance helped me in all the time of research and writing of this thesis. I could not have imagined having better mentors for my PhD study.

Besides my supervisors, my sincere thanks also go to Prof. Chetwyn Chan and to the Applied Cognitive Neuroscience Laboratory team of the Hong Kong Polytechnic University- Department of Rehabilitation Sciences. I thank my fellow lab mates from 2014-2019 for the stimulating discussions, for the exchanges of advice and words of encouragements, and for all the fun we have had in the last five years.

I am likewise grateful for the support given to me by the organizations, schools, school administrators, parents, and the children who participated in my research. Your kind understanding, and support has enabled me to conduct and finish my thesis.

Furthermore, I would like to mention the support of my friends through the process of this thesis. To my HK Pinoy Scholars friends, you have made the burden of living in foreign land bearable. To my friends in the University of Santo Tomas, your words of encouragement enabled me to push and persevere through. To my friends back in my hometown, thank you for listening to me and keeping me grounded always.

Last but not the least, I would like to thank my family: my mom and sister for supporting me spiritually throughout writing this thesis and my life in general.

This research was funded by Hong Kong PhD Fellowship Scheme and the Department of Rehabilitation Sciences of the Hong Kong Polytechnic University.

TABLE OF CONTENTS

			Page			
	Abstr	ract	i			
	Publi	cations Arising from the Thesis	iv			
	Prese	entations at International Conferences Arising from the Thesis	V			
	Ackn	owledgement	vii			
	List of Tables, Figures and Abbreviations					
Chapter 1	Intro	oduction	1			
	I.	Background	1			
	II.	Statement of Purpose	5			
	III.	Organisation of Chapters	6			
Chapter 2	Liter	rature Review	7			
	I.	Key Concepts Related to Adaptation	7			
		A. Definition of Adaptation	7			
		B. Selected Theories of Adaptation	8			
		i. Theory of Biological Adaptation	8			
		ii. Information-Processing Theory of Adaptation	9			
		iii. Life History Theory	9			
		iv. Gene x Environment Theory	9			

	v. Theory of Allostasis	10
	C. Mechanism of Adaptation	10
	D. Adaptation Enables Survival and Development	12
	E. Factors Influencing Adaptation	13
	F. Issues on the Role of Ethnicity and Environment on	13
	Adaptation	
II.	The Role and Ethnicity and Environment on Adaptation	14
	A. Ethnicity and Adaptation	14
	i. Definition of Ethnicity	14
	ii. Influence of Ethnicity on Adaptation	15
	B. Environment and Adaptation	15
	i. Definition of Environment	15
	ii. Influence of the Environment on Adaptation	17
	C. Implications of the Influence of Ethnicity and	17
	Environments on Adaptation	
III.	The Theory of Allostasis: An overview	17
	A. Theoretical Model of Allostasis	18
	B. Individual Differences Influencing Adaptation to	19
	External Challenges: The Role of Environments and	
	Ethnicity	
	i. Environment-dependent Information Processing	20
	ii. Biological Embedding in Ethnicity	20
	C. Mediators of Allostasis	21

	D. Implications of Allostasis on Adaptation	22		
IV.	Adaptation to External Sensory Stimuli	23		
	A. Features of Auditory Stimuli	23		
	B. Subcortical System and Auditory Stimuli	24		
	C. Behavioural Response to Sensory Stimuli	25		
	D. Physiological Response to Sensory Stimuli	26		
	i. Parasympathetic Response to Sensory Stimuli	26		
	ii. Sympathetic response to sensory stimuli	28		
	E. Implications of the Regulation to Response to Sensory	30		
	Stimuli on Adaptation			
V.	Ethnicity and Environment Influences the Regulation of			
	Response to Sensory Stimuli			
	A. Ethnicity and the Regulation of Response to Sensory	30		
	Stimuli			
	B. Environment and the Regulation of Response to Sensory	32		
	Stimuli			
	C. Ethnicity and Environment and the Regulation of	32		
	Response to Sensory Stimuli			
	D. Summary on Ethnicity and Environment's Influence on	34		
	the Regulation of Response to Sensory Stimuli			
VI.	Synopsis of the Autonomic Nervous System	34		

	A. Anatomy and Physiology of the Autonomic Nervous	35	
	System		
	i. Sympathetic Nervous System	36	
	ii. Parasympathetic Nervous System	38	
	B. Measures of Autonomic Functions	40	
	i. Heart Rate Variability		
	a. Overview of Heart Rate Variability	41	
	b. Physiological Mechanisms of HRV	42	
	c. HRV Analysis: Time Domain Methods	44	
	d. HRV Analysis: Frequency Domain	46	
	Methods		
	ii. Electrodermal Activity	48	
	a. Overview of Electrodermal Activity	48	
	b. The Electrodermal System	48	
	c. EDA Components	50	
	d. Standards of Skin Measurement	51	
	C. Implications of ANS on the Regulation of Response to	52	
	Sensory Stimuli		
VII.	Summary of Literature and Knowledge Gaps	52	

Chapter 3	Conceptual Framework	54		
	I. Allostasis as an Adaptive Mechanism	55		
	II. Adaptation is Influenced by Individual Differences	56		
	A. Environment-dependent Information Processing	56		
	B. Biological-embedding in Ethnicity	56		
	III. Adaptation Involves Regulation	56		
	IV. Adaptation in Facing Challenges	57		
	A. Response to Sensory Stimuli	57		
	B. Auditory Stimuli as a representative Sensory Stimuli	58		
	C. Neurophysiological Regulation of Response to Sensory	59		
	Stimuli			
	V. Ethnicity Influences Adaptation	59		
	A. Biological-embedding of Ethnicity Influences Adaptation	59		
	B. Ethnicity and Adaptive Responses to Sensory Stimuli	60		
	VI. Environment Influences Adaptation	60		
	A. Two Levels of Environment	60		
	B. Environment and Adaptive Responses to Sensory Stimuli	61		
	VII. Ethnicity x Environment Interaction and its Influence on	62		
	Adaptation			
	VIII. Knowledge Gap	63		
	IX Research Question/Motivation	6/		

	X. Research Aims	64
	XI. Research Hypotheses	65
	XII. Significance of this Study	66
Chapter 4	Methods	67
	I. Research Design	67
	II. Participants	68
	III. Experimental Protocol	70
	IV. Procedures	73
	V. Instrumentation Measures	75
	A. Heart Rate Variability	76
	B. Electrodermal Activity	77
	VI. Data Analysis	78
	A. Neurophysiological Signal Processing	78
	i. Heart Rate Variability Data Processing	78
	ii. Electrodermal Activity Data Processing	80
	B. Statistical Analysis	84
Chapter 5	Results	88
	I. Research Hypothesis 1: Regulation of response (HRV and	88

	conditions between CHK and FHK groups	
	A. Overview	88
	B. Summary of Participant Characteristics	88
	C. Differences in the Patterns of Neurophysiological	89
	Regulation Between CHK and FHK Groups	
	i. LF n.u.	90
	ii. HF n.u.	91
	iii. SCR	92
	iv. SCL	94
	D. Differences in the Neurophysiological Response to	99
	Sensory Stimuli Across Conditions Between CHK and	
	FHK Groups	
	i. Resting Condition	99
	ii. Stimulation Condition	99
	iii. Recovery Condition	100
	E. Summary of Results for Hypothesis Testing 1	102
II.	Research Hypothesis 2: Regulation of response (HRV and	104
	EDA) to sensory stimuli at resting, stimulation and recovery	
	conditions between FHK and FU groups.	
	A. Overview	104
	B. Summary of Participant Characteristics	104
	C. Differences in the Patterns of Neurophysiological	105
	Regulation Between FHK and FU Groups	
	i. LF n.u.	106

EDA) to sensory stimuli at resting, stimulation and recovery

	ii. HF n.u.	107
	iii. SCR	108
	iv. SCL	110
	D. Differences in the Neurophysiological Response to	115
	Sensory Stimuli Across Conditions Between FHK and	
	FU Groups	
	i. Resting Condition	115
	ii. Stimulation Condition	115
	iii. Recovery Condition	116
	E. Summary of Results for Hypothesis Testing 2	118
III.	Research Hypothesis 3: Regulation of response (HRV and	119
	EDA) to sensory stimuli at resting, stimulation and recovery	
	conditions between CHK and FU groups	
	A. Overview	119
	B. Summary of Participant Characteristics	119
	C. Differences in the Patterns of Neurophysiological	120
	Regulation Between CHK and FU Groups	
	i. LF n.u.	121
	ii. HF n.u.	122
	iii. SCR	123
	iv. SCL	124
	D. Differences in the Neurophysiological Response to	130
	Sensory Stimuli Across Conditions Between CHK and	
	FU Groups	
	i. Resting Condition	130

		ii. Stimulation Condition	130
		iii. Recovery Condition	131
		E. Summary of Results for Hypothesis Testing 3	133
	IV.	Research Hypothesis 4: Regulation of response (HRV and	135
		EDA) to sensory stimuli at resting, stimulation and recovery	
		conditions between FU and FR groups	
		A. Overview	135
		B. Summary of Participant Characteristics	135
		C. Differences in the Patterns of Neurophysiological	136
		Regulation Between FU and FR Groups	
		i. LF n.u.	137
		ii. HF n.u.	138
		iii. SCR	139
		iv. SCL	141
		D. Differences in the Neurophysiological Response to	146
		Sensory Stimuli Across Conditions Between FU and FR	
		Groups	
		i. Resting Condition	146
		ii. Stimulation Condition	146
		iii. Recovery Condition	147
		E. Summary of Results for Hypothesis Testing 4	149
Chapter 6	Discu	ussion	151
	I. Sı	ummary of Findings	151

	xvii
II. Synthesis of Results	152
III. Discussion Proper	152
A. Discussion of Salient Findings for Hypotheses 1-4	152
i. Hypothesis 1: Ethnicity Influence on the	152
Regulation of Physiological Response to	
Sensory Stimuli	
ii. Hypothesis 2: Geographic Environment	155
Influence on the Regulation of Physiological	
Response to Sensory Stimuli	
iii. Hypothesis 3: Ethnicity and Geographic	157
Environment Influence on the Regulation of	
Physiological Response to Sensory Stimuli	
iv. Hypothesis 4: Physical Environment	160
Influence on the Regulation of Physiological	
Response to Sensory Stimuli	
B. Novel Finding in this Thesis: Migration Influences	162
the Regulation of Physiological Response to Sensory	
Stimuli	
IV. Limitations of the Study	166
A. Sample Size	166
B. Representativeness of Samples	167
C. Measure of Change in the Environment	167

	D. Statistical/Data Limitations	167
	V. Future Research and Recommendations	168
	VI. Implications of this Study	169
Chapter 7	Conclusion	172
	I. Summary of Conclusions	172
	II. Future Work Directions	173
	References	175

LIST OF TABLES, FIGURES AND ABBREVIATIONS

	Tables	Page
Table 2.1	Measures of Allostatic Load	22
Table 2.2	Set of recommended octave-band sound pressure levels	24
Table 2.3	HRV Time Domain Methods	46
Table 2.4	HRV Frequency Domain Methods	47
Table 2.5	EDA Components	50
Table 2.6	Standards of EDA Skin Measurement	52
Table 4.1	Inclusion and Exclusion Criteria	69
Table 4.2	Specific Subgroups in this Thesis and the Gathered Sample	69
	Size	
Table 4.3	Income Classification	86
Table 4.4	Socio-economic Classification	86
Table 5.1.1	Summary of Participant Characteristics Between K and FHK	89
	Groups $(n=59)$.	
Table 5.1.2	Summary of Differences in the Patterns of Neurophysiological	96
	Regulation Between CHK and FHK groups using Main Effects	
	of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.1.3	Summary of Differences in the Patterns of Neurophysiological	98
	Regulation Between CHK and FHK Group Using Condition	
	Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.1.4	Summary Differences in the Neurophysiological Response to	101

	Sensory Stimuli Across Conditions Between CHK and FHK	
	Groups Using MANOVA Tests ($n=59$)	
Table 5.2.1	Summary of Participant Characteristics Between FHK and FU	105
	Groups (<i>n</i> = 82)	
Table 5.2.2	Summary of Differences in the Patterns of Neurophysiological	112
	Regulation Between FHK and FU Groups Using Main Effects	
	of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.2.3	Summary of Differences in the Patterns of Neurophysiological	114
	Regulation Between FHK and FU Groups Using Condition	
	Pairwise (FHK and FU) baseline Corrected 2x3 Mixed	
	Factorial ANCOVA by Neurophysiological Parameter	
Table 5.2.4	Summary of Differences in the Neurophysiological Response	117
	to Sensory Stimuli Across Conditions Between FHK and FU	
	groups Using MANOVA Tests (n= 82)	
Table 5.3.1	Summary of Participant Demographics Between CHK and FU	120
	Groups (<i>n</i> = 85)	
Table 5.3.2	Summary of Differences in the Patterns of Neurophysiological	127
	Regulation Between CHK and FU Groups Using Main Effects	
	of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.3.3	Summary of Differences in the Patterns of Neurophysiological	129
	Regulation Between CHK and FU Groups Using Condition	
	Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	

Table 5.3.4	Summary of Differences in the Neurophysiological Response	132
	to Sensory Stimuli Across Conditions Between CHK and FU	
	Groups Using MANOVA Tests (n= 85).136	
Table 5.4.1	Summary of Participant Characteristics Between FU and FR	136
	Groups $(n=97)$	
Table 5.4.2	Summary of Differences in the Patterns of Neurophysiological	143
	Regulation Between FU and FR Groups Using Main Effects of	
	Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.4.3	Summary of Differences in the Patterns of Neurophysiological	145
	Regulation Between FU and FR Groups Using Condition	
	Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA	
	by Neurophysiological Parameter	
Table 5.4.4	Summary of Differences in the Neurophysiological Response	148
	to Sensory Stimuli Across Conditions Between FU and FR	
	Groups MANOVA Tests (n= 97)	
	Figures	
Figure 2.1	The concept of allostasis and allostatic load	19
Figure 2.2	Distribution of the SNS	38
Figure 2.3	Distribution of the PNS	40
Figure 2.4	R-R interval	41
Figure 2.5	R-R interval tachogram	42
Figure 2.6	Physiologic mechanisms of HRV	43
Figure 2.7	The electrodermal system	49

Figure 3.1	Conceptual Framework: Adaptation as an approach to examine	55
	the influence of ethnicity and the environment on the	
	neurophysiologic regulation of response to sensory stimuli	
	among children	
Figure 4.1	Summary of the groupings of the participants based on the	67
	variables to be tested	
Figure 4.2	Experimental paradigm	71
Figure 4.3	Experimental paradigm stimulus	71
Figure 4.4	Screen-shot of the instructions on the computer monitor for the	73
	laboratory paradigm	
Figure 4.5	Experimental set-up	75
Figure 4.6	Sample set-up of the experiment with a participant	75
Figure 4.7	Polar H2 chest strap	77
Figure 4.8	eSense GSR sensor	77
Figure 4.9	Representation of time event epochs processed and analysed in	83
	the experimental laboratory paradigm	
Figure 5.1.1	Covariate (age and migration status) adjusted estimated	91
	marginal means for LF n.u. using baseline corrected 2x3 mixed	
	factorial ANCOVA (CHK and FHK)	
Figure 5.1.2	Covariate (age and migration status) adjusted estimated	92
	marginal means for HF n.u. using baseline corrected 2x3 mixed	
	factorial ANCOVA (CHK and FHK)	
Figure 5.1.3	Covariate (age and migration status) adjusted estimated	98
	marginal means for SCR using baseline corrected 2x3 mixed	
	factorial ANCOVA (CHK and FHK)	

Figure 5.1.4	Covariate (age and migration status) adjusted estimated	95
	marginal means for SCL using baseline corrected 2x3 mixed	
	factorial ANCOVA (CHK and FHK)	
Figure 5.2.1	Covariate (migration status and gender) adjusted estimated	107
	marginal means for LF n.u. using baseline corrected 2x3 mixed	
	factorial ANCOVA (FHK and FU)	
Figure 5.2.2	Covariate (migration status and gender) adjusted estimated	108
	marginal means for HF n.u. using baseline corrected 2x3 mixed	
	factorial ANCOVA (FHK and FU)	
Figure 5.2.3	Covariate (migration status and gender) adjusted estimated	109
	marginal means for SCR using baseline corrected 2x3 mixed	
	factorial ANCOVA (FHK and FU)	
Figure 5.2.4	Covariate (migration status and gender) adjusted estimated	111
	marginal means for SCL using baseline corrected 2x3 mixed	
	factorial ANCOVA (FHK and FU)	
Figure 5.3.1	Estimated marginal means for LF n.u. using baseline corrected	122
	2x3 mixed factorial ANCOVA (FHK and FU)	
Figure 5.3.2	Estimated marginal means for HF n.u. using baseline corrected	123
	2x3 mixed factorial ANCOVA (FHK and FU)	
Figure 5.3.3	Estimated marginal means for SCR using baseline corrected	124
	2x3 mixed factorial ANCOVA (FHK and FU)	
Figure 5.3.4	Estimated marginal means for SCL using baseline corrected	126
	2x3 mixed factorial ANCOVA (FHK and FU)	
Figure 5.4.1	Covariate (BMI) adjusted estimated marginal means for LF	128
	n.u. using baseline corrected 2x3 mixed factorial ANCOVA	

(FU and FR)

Figure 5.4.2 Covariate (BMI) adjusted estimated marginal means for HF

n.u. using baseline corrected 2x3 mixed factorial ANCOVA

(FU and FR)

Figure 5.4.3 Covariate (BMI) adjusted estimated marginal means for SCR

using baseline corrected 2x3 mixed factorial ANCOVA (FU and FR)

Figure 5.4.4 Covariate (BMI) adjusted estimated marginal means for SCL

using baseline corrected 2x3 mixed factorial ANCOVA (FU and FR)

Abbreviations

Abbreviation	Meaning
Ach	Acetylcholine
Ag/AgCl	Silver-Silver Chloride
ANCOVA	Analysis of Covariance
ANOVA	Analysis of Variance
ANS	Autonomic Nervous System
AR	Autoregression
BCa	Bias-corrected and accelerated
BMI	Body mass index
CDA	Continuous decomposition analysis
CHK	Chinese children living in Hong Kong
CI	Confidence Interval
CNS	Central Nervous System
CSP	Chinese Sensory Profile
d	Cohen's d
D4	D4 dopamine receptor gene
dB	Decibel
df	Degrees of Freedom
DV	Dependent variable
ECG	Electrocardiography
EDA	Electrodermal Activity
EDR	Electrodermal Response

ER-SCR Event-related SCR

ES Effect Size F-test

FFT Fast Fourier Transform

FHK Filipino children living in Hong Kong
FR Filipino children living in rural-Philippines
FU Filipino children living in urban-Philippines

GLM General linear model GSR Galvanic Skin Response

HF High Frequency

HF n.u. High Frequency normalized unit

HPA Hypothalamic-pituitary-adrenocortical

HRV Heart Rate Variability

HRV Heart Rate
Hz Hertz

ISI Inter-stimulus Interval IV Independent variable

KHz Kilohertz

LB Lower Bound Limit LF Low Frequency

LF n.u. Low Frequency normalized unit

M Mean

MANCOVA Multivariate Analysis of Covariance MANOVA Multivariate Analysis of Variance

MD Mean difference ms Millisecond MS Mean Squared

NN50 Number of pairs of adjacent intervals differing by more than 50 ms

NS-SCR Non-Specific SCR p Significance level

Pillai's V MANOVA Test statistics

pNN50 NN50 count divided by the total number of all NN intervals

PNS Parasympathetic Nervous System

PSD Power spectral density
R Canonical correlation
r Pearson's correlation

R² Coefficient of Regression

RMSSD Square toot of the mean of the sum of squares of differences between

adjacent NN intervals

RSA Respiratory Sinus Arrhythmia

SA Sino-atrial

SC Skin conductance

SCL Skin Conductance Level SCP Sensory Challenge Protocol

SCR Skin Conductance Response

SD Standard Deviation

SDANN Standard deviation of the average NN intervals

SDNN Standard deviation of all NN intervals

SE Sensory Experiment
SE Standard Error
Sig Significance

SNS Sympathetic Nervous System
SPS Sensory Processing Sensitivity

SS Sum of Squares

SSP Short Sensory Profile

TIE Touch Inventory for Elementary School Children

UB Upper Bound Limit
VLF Very Low Frequency

 $\begin{array}{ccc} VT & & Vagal\ Tone \\ \Lambda & & Pillai's\ V \\ \mu S & & Microsiemens \end{array}$

Chapter 1: Introduction

This section of the thesis introduces the background of the study that is concerned on the role of ethnicity and environments on the regulation of response to sensory stimuli in children. It is then followed by the statement of purpose. This section ends with an overview of how the thesis is organised.

I. Background

Adaptation is defined to be a biological response to challenges posed by the external environments (Bock, 1980; Karatsoreos & McEwen, 2013; Losos, 2011; McEwen, 2007; Pery et al., 1995). Adaptation to the situation is needed to overcome this change of environment or the challenges faced in the environment. In fact, adaptation has been implicated to be important in the development of stress regulation (Bortoluzzi et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012; Roberts et al., 2015), emotion regulation (Dieleman et al., 2015), academic (Pagliaccio et al., 2015) and social behaviours (Pagliaccio et al., 2015) to name a few. Thus, adaptation represents a growing field of inquiry in research, specifically the factors that might influence its development. An individual's ability to respond to external challenges is influenced by individual differences (McEwen, 1999, 2008). These individual differences may be related to one's genetic traits, age, gender, experiences, and the living environment. There are two main categories of individual differences that can influence adaptation: expressive-dependent information appraisal and biological embedding (Danese & McEwen, 2008). Biological embedding refers to how genetic traits are expressed as a representation of a child's ethnic origins. Experience-dependent information appraisal is related to the environment where a child develops in that shapes how they respond to challenges and changes emanating from their extrinsic contexts. Understanding how individual differences such as ethnicity and environment influence a child's ability to adapt to external challenges may help understand the underlying mechanisms that enable survival and development.

Adaptation involves responding to the demands of the environment through modifications of forms and features of an individual (Bock, 1980; Nemenman, 2010). These responses can be characterized as physiological or behavioural (Shimada et al., 2010). However, behavioural and physiological responses may not always be related (Gomez et al., 2017). Physiological responses represent an internal mechanism that allows adaptation (McEwen,

1998, 2000; McEwen & Wingfield, 2003; Sterling & Eyer, 1988). These physiological mechanisms have been suggested to support behavioural responses (Danese & McEwen, 2012). The use of neurophysiological measures has been suggested to offer a more valid method to index adaptive responses (Gomez, Lai, Chan & Tsang, 2018).

The external environment is full of sensory stimuli. The sensory stimuli represent the most basic form of stimuli within a child's environment. Adaptation to these information and stimuli is imperative to support development across lifespan. The response to sensory stimuli is defined to be an individual's reactivity to the sensory information input from the environment (Gomez, Lai, Morato-Espino, Chan & Tsang, 2017). The literature that examines the regulation of response to sensory stimuli seems to largely utilize behavioural and physiological measures (Schoen, Miller, Brett-Green & Nielsen, 2009). Behavioural outcomes, whether through clinical observations or parent reports, are vulnerable to subjectivity (Schoen et al., 2009). Furthermore, behavioural evidence may not be as objective, and the information may be confounded with parental factors (Hoyle, Harris & Judd, 2002; Ooi et al., 2016; Orekhova & Stroganova, 2014). A more objective and sensitive approach to measure the underlying mechanisms of regulation in internal states as a response to external challenges is the measurement of physiological responses (Appelhans & Luecken, 2006).

Child development does not occur in a vacuum, and several factors such as ethnicity and environment may influence children's adaptation to various sensory stimuli in their external environment. Ethnicity refers to distinct populations that share biological origins and genetic similarities (Ali-Khan, Krakowski, Tahir & Daar, 2011; Jorde & Wooding, 2004). However, the literature concerned on the influence of ethnicity on the regulation of response to sensory stimuli yields conflicting results. Royeen and Mu (2000) found no significant difference in the sensory responses of European and American school-aged children, suggesting that the response to sensory stimuli is a universal phenomenon. On the other hand, Tirosh, Bendrian, Golan, Amir and Dar (2003) suggested that for children with comparable ages, the regulation of response to sensory stimuli is different between Arabic and Jewish children, suggesting ethnic differences. The foregoing critic of the literature concerned with examining the influence of ethnicity on the regulation of response to sensory stimuli appears to be inconclusive and inconsistent. Furthermore, the available evidence has come from behavioural perspectives (i.e. clinical observation, behavioural checklists).

The environment is defined to be the geographic range where a group of individuals exists (Lee & Minard, 2013). The environment has been suggested to be the sum of extrinsic factors that individuals may potentially encounter. These factors include sociocultural constructs and the biological ideation of environments as ecological niches among research in the biological sciences. Biological features of the environment (i.e. climate, habitat, geography) as well as "objective characteristics of the physical context related to habituation and gradients of man-made or natural structures and components" (i.e. nature, urbanisation; Gomez et al., 2018, p. 3) are different levels of the environment (Bock, 1990; Lee & Minard, 2013; McDonnell & Pickett, 1990; Perloff, 2015). The geographic environment (i.e. country of habitat) where a child develops in has the ability override ethnic influences on adaptive sensory behaviours (Gunn et al., 2009). Nevertheless, the contexts within experiences are constructed in one environment may be different another. For example, Caron, Schaaf, Benevides and Gal (2012) compared the sensory behaviours of children living in Israel and the United States of America, and found that the physical setting, culture and parental factors contributed to differences seen among the studied population. Furthermore, the physical features within an environment where children dwell can greatly impact how individuals regulate responses to the external world (Laumann, Garling, & Stormark, 2003; Rutter, Pickles, Murray & Eaves, 2001; Ulrich et al., 1991). These physical environments have been suggested to comprise the physical characteristics of the landscapes such as in the case of urban and rural contexts (McDonnell & Pickett, 1990). An urban setting is characterized by a density of man-made human structures (i.e. houses, buildings, bridges, railways, etc.). On the other hand, a rural environment may be viewed as an open strip of natural environment (i.e. mountainous, coastal, agricultural) where fewer humans are living with lesser man-made establishments. However, whether the physical environment influences the variations in response toward sensory stimuli among individuals sharing similar ethnicities across environments is still uncertain. It has been suggested that sociocultural constructs and the biological ideation of environments should be individually examined. Moreover, the available evidence that supports the influence of the environment on the regulation of response to sensory stimuli in children has likewise come from behavioural perspectives.

Indexing the activity of the autonomic nervous system has been suggested to offer an objective measure of the regulation of response to sensory stimuli. Previous research has found supporting evidence that associates sympathetic (McIntosh et al., 1999a, 1999b) and

parasympathetic autonomic functions (Schaaf, Miller, Seawell & O'Keefe, 2003; Schaaf et al., 2010) to the regulation of response to sensory stimuli. Autonomic measures could provide objective measures of the capacity to regulate and the status of internal states. However, the influence of ethnicity and environment on the neurophysiological regulation of response to sensory stimuli in children is yet to be fully understood (Gomez, Lai, Chan & Tsang, 2018).

Findings in previous studies have a limitation on its ability to explain the influence of ethnicity and environment. It is unclear whether differences in the regulation of response to sensory stimulation are due to a child's ethnicity or because of the environment of children's habitat. The context upon experiencing the sensory stimulation may be varied between levels of the environment and may influence how children adapt to sensory stimuli. The previous studies mainly used behavioural outcomes to measure adaptation to sensory stimuli, which may reflect a lesser objective measure for precise information about the regulation of response to sensory stimuli (Schoen et al., 2009). In this research, children are recruited from similar ethnic origins (i.e. Filipino) from different geographic environments (i.e. Hong Kong and Philippines) and physical environments (i.e. urban and rural Philippines). Neurophysiological measures that represent the interrelated operations of the parasympathetic and sympathetic branches of the ANS are primarily measured on how children adaptively regulate responses to sensory stimuli.

Adaptation to external challenges such as sensory stimuli is important in the development of children. Examining the factors that contribute to a child's ability to adapt provides a deeper understanding of the mechanisms related to adaptation. The influence of ethnicity and environments on the regulation of response has been sparse or at best yielding inconsistent results. However, there is a conceptual reason to believe that a child's ethnicity and their living environments can influence how children regulate responses to sensory stimuli. Methodological variations and the limited inquiry on using more objective measures of the underlying mechanism that enables adaptation through regulation were suggested.

II. Statement of Purpose

This thesis aims to examine the role of ethnicity and environment on the regulation of response to sensory stimuli among children using neurophysiological methods. To answer such an aim, several hypotheses were tested. Hypothesis testing 1 aimed to examine whether ethnicity influences the regulation of response to sensory stimuli between children with different ethnicities (Chinese and Filipino) but lives in the same geographic environments (Hong Kong) and environment landscapes (urban setting). Hypothesis testing 2 aimed to examine whether geographic environments influence the regulation of response to sensory stimuli, between children with similar ethnicity (Filipino) but live in different geographic environments (Hong Kong and Philippines) but of similar environment landscapes (urban settings). Hypothesis testing 3 aimed to examine whether ethnicity and geographic environments influence the regulation of response to sensory stimuli, between children with different ethnicities (Chinese and Filipino) living in different geographic environments (Hong Kong and Philippines) but similar environment landscapes (urban setting). Lastly, Hypothesis testing 4 aimed to examine whether physical environments influence the regulation of response to sensory stimuli, between children with similar ethnicity (Filipino) living in similar geographic environments (Philippines) but different physical environment landscapes (urban and rural setting)..

The findings of this research have several implications that address research, practice and policy development relevant to childhood populations. Firstly, this research will be able to demonstrate a reconceptualization of the neurophysiological mechanisms behind the regulation of response to sensory stimuli. Understanding such mechanisms helps in better understanding of factors that may influence the development of children. Secondly, clinical practice may be informed in the development of relevant assessment tools that address physiological components; and in creating innovative and novel interventions that address the underlying mechanistic problems that support behaviours related to the dysregulation of response to sensory stimuli. Lastly, the findings of this research may be able to support the idea of developing policies that address the health and well-being needs of children who are migrating from one environment to another.

III. Organisation of Chapters

The chapters within this thesis are organised according the different parts that enables it to answer the research question. Chapter 1 provides a brief introduction to the thesis topic.

Chapter 2 provides a review of the relevant literature describing the concept of allostasis, adaptation to sensory stimuli, the role of ethnicity and environment on adaptation and a synopsis of the autonomic nervous system. Chapter 3 is a critical and reflexive synopsis of the literature explaining the core concepts applied in this thesis. Chapter 4 describes the general methods applied in conducting the experiments that support in answering the research questions within this thesis. Chapter 5 reports the results of the four individual hypotheses testing in this thesis. Chapter 6 provides a discussion of the results of this thesis. This chapter offers a summary of the findings, synthesis of results, a summary statement of findings per study and novel findings, limitations of the study, future research and recommendations, and the implications of the results of the thesis. Finally, Chapter 7 describes the summary of the conclusions of this thesis and briefly expounds future research plans and directions. This document ends with the references to the works cited in this thesis.

Chapter 2: Literature Review

Chapter 2 of this thesis provides a review of the relevant literature describing the concept of allostasis, adaptation to sensory stimuli, the role of ethnicity and environment on adaptation and a synopsis of the autonomic nervous system. The end of this chapter summarises the extant literature in light of the gaps in knowledge that informs the research questions of this thesis.

I. Key Concepts Related to Adaptation

Adaptation is necessary for survival. Individuals must be able to respond to the demands of their external environments in order to thrive. This section of the literature commences with a discussion of the key concepts of adaptation. Furthermore, this section also describes relevant theories of adaptation, the mechanics, and processes related to adaptation, its importance and factors that might affect such.

A. Definition of Adaptation

The term "adaptation" has long been used in the biological sciences and conceptually defined and redefined within the contexts of varying research inquiries (Bock, 1980). However, a key feature to the various definitions suggest that adaptation is generally an individual's response to challenges posed by their external environments (Bock, 1980; Karatsoreos & McEwen, 2013; Losos, 2011; McEwen, 2007; Pery et al., 1995) that enables them to survive and thrive (Shimada, Ishii & Shibao, 2010). Adaptation involves a biological role in modifying one's characteristics as a response to environmental demands (Bock, 1980; Nemenman, 2010).

In the biological field, two types of adaptation have been suggested by Shimada et al. (2010): physiological adaptation and biological adaptation. Physiological adaptation refers to short-term and context-based features of an individual related to separate physiological and behavioural adjustments (Withagen & van Wermerskerken, 2010) to their current environments. Physiologic adaptation describes the plasticity of the individual's phenotype, without permanent change. For example, studies on the effects of previous experiences (i.e. trauma, stress) have found that while the genetic expression related to the stress response of children changes in the hypothalamic-pituitary-adrenal axis response, no morphological

changes occur (Bortoluzzi et al., 2015; DeRijk, 2009; Pagliaccio et al., 2015a,b; Roberts et al., 2015). Biological adaptation, on the other hand, describes a more permanent phenotypic feature change of the individual that enables them to survive in the environment. For example, Schlichting and Wund (2014) suggested that constant exposure to high altitude environments resulted in genetic accommodations with Tibetan ethnic groups that altered their physiological oxygen consumptions across generations distinctly different from other populations.

Adaptation as a concept has been used to describe either as an outcome or a process (Bock, 1980; Futuyma, 2009, Perry et al., 1995). As an outcome, adaptation can refer to physiological and behavioural modifications within the context of a specific environmental condition (Shimada et al., 2010; Withagen and van Wermerskerken, 2010). On the other hand, adaptation can also refer to the process by which an individual becomes better suited to some feature within their environments by modifying physiology or behaviour (Futuyma, 2009).

In this research, adaptation is defined as the physiological changes in the internal environment in response to a challenge from the external environment.

B. Selected Theories of Adaptation

There are several theories that try to encapsulate the concept of adaptation ranging from the classical (i.e. nature selection, evolution) to the more contemporary. In order to review the theories of adaptation, this thesis focuses on salient theories relevant to its research questions. Five selected theories are summarised in the preceding subsections.

i. Theory of Biological Adaptation

Bock (1980) described the theory of biological adaptation underscoring that adaptation refers to sets of phenotypic features relative to the demands of the individual's environment. These phenotypic features range in form resulting from physiological alterations of its biological role in meeting external environmental demands. Responding to the environment produces sets of physiological responses that are well suited to the organism's precise environment, enabling survival. It is the interaction between the demands of the external environments as well as the individual's physiology and the

morphology that occurs within structural networks that characterizes biological adaptation.

ii. Information-Processing Theory of Adaptation

A more contemporary theory of adaptation was offered by Nemenman (2010), where adaptation was described as a property of how an individual's own characteristics are changed in response to stimuli coming from the external environment. Adaptation is achieved through a manner of processing the incoming external information that shapes how one responds. There is an interaction between how internal states respond to the external world. However, no clear distinction of whether the adaptation is achieved through short-term responses or long-term exposures.

iii. Life History Theory

The life history theory was suggested by Fabian and Flatt (2012) to be a branch of evolutionary ecology. While similar to other evolutionary theories that try to explain diversity in life species, the life history theory is unique in its focus on understanding adaptation. It tries to explain the development of traits or features of an individual that enables survival and posterity. The life history theory attempts to describe traits (i.e. age, behaviour, physiology) as an explanatory factor to adaptation as an outcome. Genetic variations and probable differentiation of phenotypic traits occur because of interactions with the environment. Adaptation is a plastic phenomenon that is occurring within a generation, without permanent phenotypic changes, but rather on its expression as a function of behaviour or physiology. While useful in explaining the development of behavioural and physiological features of organisms, the life history theory has largely been used among non-human samples.

iv. Gene x Environment Theory

The gene x environment theory describes how organisation of biological and psychological systems and their interactions can result in molecular changes as a function of adaptation to external challenges (Cicchetti, 2010; Cloninger et al., 1982; Lopizzo et al., 2015; Powledge, 2011). In this theory,

exposure to environmental experiences moderates the genetic effects on the developmental outcomes. For example, experiences under specific environments may only relate to outcomes among individuals with particular genetic characteristics expressed through behavioural and physiological features. It is through this gene x environment interaction that supports individuals, specifically among children, that enables them to effectively adapt.

v. Theory of Allostasis

The earlier works of McEwen (1998, 2000), Sterling and Eyer (1988), and McEwen and Wingfield (2003). Allostasis explains how individuals adapt to challenges imposed by external environments through an internal regulatory mechanism mediated by multiple physiological systems. Thus, allostasis enables adaptation through perturbations among physiological parameters that support behavioural responses. Furthermore, allostasis recognizes that various factors may affect the underlying mechanisms of adaptation. The ability of adaptation to external challenges is influenced by two principal factors: environment-dependent processing and biological embedding (Danese & McEwen, 2012). A more detailed discussion of allostasis can be found in section III of this chapter.

C. Mechanism of Adaptation

Adaptation entails response generation in light of the information that comes from the external environment (Nemenman, 2010). This information can come in the form of changes, stimuli, challenges or threats. Adaptation can occur in two ways: as an outcome and as a process (Bock, 1980; Futuyma, 2009, Perry et al., 1995). However, the key to understanding whichever type of adaptation is the mechanism that underlies such response generation. In this section, the mechanism underlying adaptation is summarised below and was chosen based on relevant theories (i.e. gene x environment and allostasis) that help support the conceptual framework of this thesis.

The theory on gene x environment interactions proposes that the environmental exposures can influence the genetic traits (Lopizzo et al., 2015). The exposure of individuals to certain environmental conditions can alter how genes express

themselves and eventually behaviours of individuals (Cicchetti, 2010). A popular way of examining this is by looking at adoption studies. For example, the seminal work by Cloninger, Sigvardsson, Bohrnan, and von Knorring's (1982) was noted to be the first study to demonstrate the interaction between genetic heritability and the influence of the environment in the development of disruptive behaviours in children. The authors found that under low genetic risk conditions (i.e. having a parent with no known criminal history), the support offered by the living environment (i.e. adoptive parents with no known criminal history) results to lower risk of criminality among the children when they grow up. Nevertheless, a high genetic risk coupled with a supportive environment (i.e. adoptive parents with no known criminal history) results to a lower probability of criminality compared to those with an unfavourable (i.e. adoptive parents with a known criminal history). This exemplar study illustrates how genetic susceptibility can be influenced by a supportive environment. Exposure to such a supportive environment can override genetic susceptibility. It is likely that molecular adaptations in response to the environment supported the lower probability of the expressing the genetic predispositions (Bortoluzzi et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012; Roberts et al., 2015). However, the major limitation of the gene x environment is that it fails to succinctly describe how morphisms occur in response to the environment. While adaptation is seen as an outcome, the process by which it is achieved remains highly theoretical. This may be due to the fact that genetic responses are not readily available for non-invasive and in-situ examination. Other physiological indices that can represent the mechanism of adaptation are needed for better illustration, modelling, and experimentation (Bock, 1980).

Adaptation through perturbations across multiple yet interrelated physiological systems was previously suggested by proponents of the concept of allostasis (McEwen, 1998, 2003). In allostasis, the central mechanism that enables adaptation is related to the nervous system, which is deemed responsible for the regulation of response to the challenges, information, and stimuli from the external environment. Adaptation as a process of internal state homeostasis is achieved through the responses of various mediators of allostasis (i.e. glucocorticoids, cytokines, neutrophils, acetylcholine, serotonin). When the individual is faced with a challenge from the external environment, these mediators respond by increasing or decreasing

their parameters. For example, when a child hears a loud siren, several physiological systems are activated, with the objective of protecting the individual from harm through adaptation. Blood pressure may go up, along with an increase in heart and respiratory rate, or even hand sweating. Alterations in the activity of these mediators underlie physiological homeostasis that in turns support behaviours. A more succinct description of the mechanism of allostasis is described in section III of this chapter.

In this research, adaptation as a feature of the internal physiological mechanism is utilised to index the changes in response to the challenges from the external environment.

D. Adaptation Enables Survival and Development

Adaptation as a property of being able to change one's characteristics in response to the external environment has been suggested greatly influence survival and thriving (Futuyma, 2009; Karatsoreos & McEwen, 2013; McEwen, 2007; Nemenman, 2010; Perry et al., 1995; Shimada et al., 2010). It suggests a sense of mastery over the environment where an individual situates himself; the environment pertaining to the sum of all extrinsic factors encountered by individuals such as habitat, climate, temperature, etc. (Losos, 2011). Various evolutionary authors have illustrated how the inability of an organism to adapt to their environment can cause catastrophic consequences. However, the importance of adaptation is not limited to a dichotomous condition of living or not.

Previous authors have implicated on the implications of adaptation to the development of children. To illustrate, Fazel Reed, Panter-Brick and Stein (2012) found that adaptation to displacement in children refugees may upset mental health functioning. However, immediate subsequent positive experiences (i.e. resettlement, social support) can amend future success educational functioning for these children. Margolin and Vickerman (2007) supported this finding in their review and found that childhood exposure to family violence may increase the risk for post-traumatic stress disorder (PTSD) that can affect multi-dimensional development (i.e. social, physiological, cognitive, behavioural, and affective). Adaptation to the situation is needed to overcome this change of environment or the challenges faced in the new environment. Certainly, adaptation is not limited to

these examples alone. Moreover, adaptation has been implicated to be important in the development of stress regulation (Bortoluzzi et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012; Roberts et al., 2015), emotion regulation (Dieleman et al., 2015), academic (Pagliaccio et al., 2015) and social behaviours (Pagliaccio et al., 2015) to name a few. Given the importance of adaptation and its implications for childhood populations, this research is being conducted.

E. Factors Influencing Adaptation

Individual differences shape how individuals adapt through responding to external challenges (McEwen, 1999, 2008). Individual differences may be categorized into two: 1) expressive-dependent information appraisal; and 2) biological embedding (Danese & McEwen, 2008). Biological embedding refers to how genetic traits are physiologically expressed by the body's condition (McEwen, 2008). These genetic may include but are not limited to one's gender, age, ethnicity, diseases, etc. Experience-dependent information appraisal suggests that one's life history, which may include experiences situated in the environment of context (i.e. geographic, physical, social, cultural, etc.) has shaped our abilities to make sense of situations and instilled in us a set of codes that determines how we adapt to a situation (McEwen, 2008; Mims & Olden, 2012; Templer, 2008; Wolf, van Doorn, Leimar & Weissing, 2007). Specific to this research, the factors related to a child's ethnicity and environment will be explored on its influence on their adaptation in response to sensory stimuli.

F. Issues on the Role of Ethnicity and Environment on Adaptation

Adaptation is necessary for survival. Adaptation is an individual's response to challenges posed by their external environments (Bock, 1980; Karatsoreoes & McEwen, 2013; Losos, 2011; McEwen, 2007; Pery, et al., 1995) that enables them to survive and thrive (Shimada, Ishii & Shibao, 2010). There are several theories that try to encapsulate the concept of adaptation. One of these is theory allostasis, which suggests that adaptation is a response to challenges or changes in one's external environment through perturbations among physiological parameters (McEwen, 1998, 2000; McEwen & Wingfield, 2003; Sterling & Eyer, 2003). Adaptation as a process of internal state homeostasis is achieved through the responses of various mediators of allostasis, such as indices of the parasympathetic

and sympathetic branches of the ANS. The ability to adapt to external challenges can be influenced by several factors such as the ethnicity and the living environment of an individual (Danese & McEwen, 2012). An understanding of how one's ethnicity and environment will further enhance the knowledge of the mechanisms that underlie the adaptation of individuals throughout the lifespan and its effects on their development. In this research, adaptation is approached from a biological perspective. It is intended to examine the biological influence of ethnicity and environment on the physiological responses to challenges from one's external environments.

II. The Role and Ethnicity and Environment on Adaptation

Individual differences are variations is different factors from one person to another that distinguishes or separates them, from others and makes then unique individuals. When applied to the ability to adapt to external challenges, individual differences can be broadly differentiated into two broad categories: inherited or acquired (Danese & McEwen, 2012). In this research, inherited differences are conceptualized to be factors relating to one's ethnicity. Acquired differences, on the other hand, is conceptualized to be the influence of the child's environments. The aim of this research is to explore how differences in a child's ethnicity and environment influence their ability to regulate the response to sensory stimuli.

A. Ethnicity and Adaptation

i. Definition of Ethnicity

Several definitions exist explaining the concept of ethnicity. In biology, ethnicity generally refers to distinct populations that share biological origins and genetic similarities (Ali-Khan et al., 2011; Jorde & Wooding, 2004). In this group of people, there is a sense of shared descent and morphological variations and expressions of their DNAs that can distinctly characterize one ethnic group from another in terms of appearance, and their physiology. The genetic makeup of an individual strongly influences behaviours, which in turn can moderate transmission of behaviour and genetic makeup of a child (Klahr & Burt, 2014). On the other hand, it has been argued that ethnicity can be a social construct that may include social, cultural, heritage and language (People & Bailey, 2010), biological construction of the concept should independently differentiate itself and

focus more on parameters of genetic similarities or variances (Jackson, 1992; Overfield, 2017). This research adopts the definition of ethnicity from a biological perspective, which describes the genetic characteristic of a group of people that influences their internal physiology.

ii. Influence of Ethnicity on Adaptation

Adaptation has been defined to be the internal modifications of an individual in response to changes in their external environment (Bock, 1980; Karatsoreoes & McEwen, 2013; Losos, 2011; McEwen, 2007; Pery, et al., 1995). Specifically, physiological changes have been pointed out to be a key indicator of adaptation (Shimada et al., 2010; Withagen & van Wermerskerken, 2010). Since ethnicity refers to the physiological makeup of a group of similar people, there is a reason to believe that they likewise share the same mechanisms that enable them to adapt to challenges or changes from the environment. These responses have been contextualised specifically to their environment and the information and experiences within it (Bock, 1980; Nemenman, 2010). Constant exposures to these environments have likely shaped an ethnic group's physiology to specifically match their environments and attain adaptation (Cicchetti, 2010; Fabian & Flat, 2012). Physiological responses to the environment have been suggested to support behavioural responses (McEwen 1998, 2000, 2007). Thus, different ethnic groups have different underlying morphological variations of genetic traits that ultimately influence their adaptation to their environments (Jackson, 1992; Overfield, 2017).

B. Environment and Adaptation

i. Definition of Environment

The environment has been suggested to be the sum of extrinsic factors that individuals may potentially encounter. The environment is defined to be the geographic range where a group of individuals exists (Lee & Minard, 2013). Several researchers argued that the environment involves the similarities socio-cultural traits and experiences embedded within similar geographic environments results into behavioural responses that are shared by a category of people that set apart one group of people from another

(Robinson-Wood, 2016; Zimmerman & Woolf, 2014). However, there needs to be a clear distinction between these sociocultural and biological constructs of environments as ecological niches among research in the biological sciences. Biological features of the environment (i.e. climate, habitat, geography) as well as objective characteristics of the physical context related to habituation and gradients of man-made or natural structures and components (i.e. nature, urbanisation) are different levels of how previous researchers describes and conceptualised the environment (Bock, 1990; Lee & Minard, 2013; McDonnell & Pickett, 1990; Perloff, 2015).

In this research, two distinct levels of the environment are examined: geographic and physical environments. The geographic environment refers to the ecological niche represented the country of abode, where sociocultural experiences are likewise embedded.

The physical environment, on the other hand, refers to specific biological features within the environment related to nature or lack thereof (i.e. urbanisation) represented by the dichotomous characteristics of urban or rural contexts. Several definitions are available to describe urban and rural physical environments. However, their focus on economics and country-specific definitions result in confusion and non-consensus. Thus, to operationalise the definition of the physical environment, this research operationalises its definition and adopts the contemporary description by Perloff (2015). The urban context is described to be set in a metropolitan densely populated within a number of man-made human structures (i.e. houses, buildings, bridges, railways, etc.). On the other hand, a rural environment may be viewed as an open strip of natural environment (i.e. mountainous, coastal, agricultural) populated with fewer human inhabitants and man-made establishments.

ii. Influence of Environment on Adaptation

Adaptation is a feature that has been shaped in response to the specific characteristics of the environment (Losos, 2010; Shimada et al., 2010). Development has been suggested to have an interplay of how children

interact with features of the environment which includes the experiences within (Hertzman, 2012). The information from the environment and the adaptation that occurs along with it influences how the nervous systems respond to such (Bock, 1980; Losos, 2010). At the molecular levels, neurons are excited from the influx of information from the external environment, signaling information processing (Nemenman, 2010). This reactivity may involve several systems including the brain, the hypothalamic-pituitary-adrenocortical (HPA) axis or the autonomic nervous system (ANS), which triggers physiological events aimed at regulating the internal state in response to the external challenge (McEwen 1998, 2000, 2007). Following repetitive and constant exposures to sums of these extrinsic stimuli from the environment, physiological systems develop specific reactions. Thus, individuals living in similar environments can be hypothesized to have more likely similar internal physiological regulatory responses to external stimuli.

C. Implications of the Influence of Ethnicity and Environments on Adaptation

Adaptation is an important aspect that supports the development of individuals,
especially among children. Therefore, understanding how factors such as a child's
ethnicity and environments influence their ability to respond to external challenges
is of great importance. This includes an investigation of how ethnicity and
environments ultimately influence the neurophysiological mechanisms that support
adaptation. This research adopts the definition of ethnicity and environments from a
biological perspective, examining their influences on the internal physiological
responses that enable adaptation to external challenges from the environment.

III. The Theory of Allostasis: An Overview

Allostasis is a theory that suggests adaptation to external challenges through changes in the behavioural or physiological responses. The theory of allostasis may provide insight into understanding how a child's ethnicity and environment can influence the regulation of response to sensory stimuli.

A. Theoretical Model of Allostasis

Allostasis explains how individuals adapt to challenges imposed by external environments through an internal regulatory mechanism mediated by multiple physiological systems (McEwen, 1998, 2000; McEwen & Wingfield, 2003; Sterling & Eyer, 1998).

In allostasis, alteration of the regulatory parameters (e.g. an increase or decrease in the conventional homeostatic physiological mechanism) allows a person to adapt to environmental challenges. This was described by Ganzel and Morris (2011) as an adaptive response to external challenges that result in a dynamic process of forming a new physiological homeostasis. This was differently explained by Juster et al. (2011) who suggested allostasis as a continuous re-evaluation and readjustments that are able to develop new baselines that can optimize resources (e.g., increased cardiac output when running). Allostasis allows homeostasis, survival and shortterm adaptation. However, it can likewise be involved in permanent changes after a prolonged and protracted stressor subsequently referred to as allostatic load (McEwen & Wingfield, 2003). When frequent allostatic response happens, it takes on a cost (i.e., an allostatic load). The initiation and sustenance of allostatic responses are done by central (i.e. hippocampus, prefrontal cortex, amygdala) and peripheral (i.e. ANS) structures (Ganzel et al., 2010). The first physiological axis that is activated as a response to environmental challenges is the ANS. Indicators of responding to environmental challenges include blood pressures, cardiac output, electrodermal activity. heart rate, and respiratory rate. (Matsubara et al., 2011; McKay, Buen, Bohan, & Maye, 2010).

Allostasis as a theoretical concept is represented by Figure 2.1 based on the works of McEwen (1999). In this model, there are several important components that influence the allostatic process (i.e. environmental stressors, previous life experiences, individual differences). Previously, it has been defined that allostasis refers to the active regulatory process of continuously responding to external challenges and evaluating the individual's physiological needs and consequently dynamically adapting to such (Karlamangla, Singer, McEwen, Rowe & Seeman, 2002). Flexibility of responses is needed to successfully adapt to the incessant stimulation or challenges in the environment. However, several factors related to

individual differences may likely mediate how people respond to said external information. One factor can be related to expressive-dependent information appraisal (Danese &McEwen, 2011), which refers to the perception and interpretation of external information (McEwen, 2008). Thus, an individual's life history (i.e. social or economic backgrounds) has instilled a set of codes determining the reaction or appraisal of a situation. The second factor is related to biological-embedded traits (Danese & McEwen, 2011), which is more closely related to the physiological expression of genetic traits exemplified by our body's condition (McEwen, 2008). People who are genetically predisposed to certain physiological symptoms may react to stress a bit more than those who are not. A more recent suggestion posits that it may likely be the combination of both factors, *gene X environment* that has a stronger ability to influence the internal response of an individual upon external challenges (Danese & McEwen, 2012).

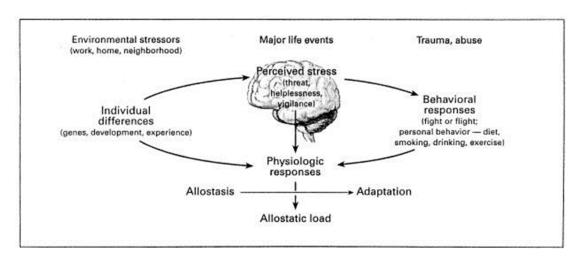


Figure 2.1. The concept of allostasis and allostatic load (Reprinted with permission from McEwen, B. S. (2000). Allostasis and allostatic load: implications for neuropsychopharmacology. *Neuropsychopharmacology*, 22(2), 108-124.).

B. Individual Differences Influencing Adaptation to External Challenges: The Role of Environments and Ethnicity

The ability of adaptation to external challenges is influenced by two principal factors: environment-dependent information processing and biological embedding in ethnicity (Danese & McEwen, 2012).

i. Environment-dependent information processing

Environment-dependent information processing refers to the perception and interpretation of events or challenges by the individual as influenced by their previous experiences (Danese & McEwen, 2012). For example, Grassi-Oliveria, Ashy, and Stein (2008) synthesized the extant literature on the deleterious consequences to previous harsh experiences related to child abuse and neglect and suggested that reprogramming activity of the HPA axis ANS occurs. Early stressful events, childhood trauma, and parenting experiences have been shown not only to increase salivary cortisol levels that modulate the autonomic state of children and these children had a higher risk of developing anxiety disorders. It was suggested that ANS and HPA measures are associated with stressful conditions and can influence social and academic behaviours for both children with and without anxiety disorder (Dieleman et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012). Kuhlman et al. (2015) found that childhood HPA-axis functioning is affected by early trauma exposure that may further influence how children regulate stress. A supportive environment is associated with a better functioning HPA axis and can influence the regulation of stress which can influence subsequent childhood behaviours in other areas of functioning (Marsman et al., 2012). However, Dieleman et al. (2015) noted that the HPA axis might be different from the autonomic activity and recommended the use of specific autonomic measures of the sympathetic (SNS) and parasympathetic (PNS) branches of the ANS (i.e. heart rate variability, respiratory rate, electrodermal activity). Thus, how individuals, specifically among children, process stimuli from the external environment is dependent on the environment where they grow up in.

ii. Biological embedding in ethnicity

Development of childhood behaviours can be influenced by genetic predispositions. Several genetic markers have been suggested to influence ethnic differences among humans. One of them is the D4 dopamine receptor gene, which has been implicated in its ability to moderate cultural differences (i.e. European-American and Asian-Americans) in social behaviours (Kitayama, King, Yoon, Tompson, Huff & Liberzon, 2014).

For example, Knafo, Israel and Ebstein (2011) examined genotyping (molecular genetic strategy) of the D4 dopamine receptor gene (D4) and its role in social behaviours. They found out that D4 is responsible for controlling compliant prosocial behaviours elicited in response to social requests and self-initiated prosocial behaviour enacted voluntarily. Bakermans-Kranenburg and van Ijzendoorn (2011) suggested the role of dopamine-related genes as a factor that leads to susceptibility in influencing the modulation of a child's attention and executive control; prosocial behaviour (i.e. donating, sharing); irritability; and stress dysregulation among others. Genetic markers related to atypical and/or dysregulation of autonomic activity underlying pathology in children with neurobehavioural disorders are identified, such as, CRHR1 (rs4792887, rs110402, rs242941, rs242939, rs1876828) and FKPB5 (rs1360780, rs3800373, rs9296158, rs9470080, rs4713916 (Bortoluzzi et al., 2015; DeRijk, 2009; Pagliaccio et al., 2015a,b; Roberts et al., 2015). Thus, biologically-embedded genetic information and its consequent expression may thus influence how children respond and adapt to external challenges.

C. Mediators of Allostasis

The common way of measuring allostatic load is through a various range of biomarkers that sensitive to dynamic and subtle changes within internal physiological states, measured after environmental exposures. (Read & Grundy, 2012). The state of allostasis refers to a sustained alteration of the primary allostatic mediators' activity (e.g., cardiovascular, hormonal, or cytokines regulation), that combines physiological and behavioural responses associated with challenges imposed by the changing external and internal environments (McEwen & Wingfield, 2003, 2010). Efferent activity combination and permutations (i.e. increase/decrease in heart rate, piloerection, respiration, sweating of the palms, temperature) mediates physiological mechanism and processes aimed at target organs (Everly & Lating, 2012) that are directed towards many diverse organs. The neurological axes consisting of the HPA and ANS are the first systems that activate and respond in the face of external challenges. Specifically, mobilization of the sympathetic, parasympathetic, and neuromuscular nervous systems consists of

immediate response to external challenges to maintain internal homeostatic balance (Everly & Lating, 2012).

Table 2.1. Measures of Allostatic Load (source: McEwen, 1999).

Measures of Allostatic Load

Systolic and diastolic blood pressure

Indices of cardiovascular activity

Waist-hip ratio

An index of more chronic levels of metabolism and adipose tissue (influenced by increased glucocorticoid activity)

Serum HDL and total cholesterol

Related to the development of atherosclerosis (increased risks being seen with higher levels in the case of total cholesterol and lower levels in the case of HDL)

Blood plasma levels of glycosylated haemoglobin

An integrated measure of glucose metabolism over several days)

Serum dehydroepiandrosterone sulphate (DHEA-S)

A functional HPA axis antagonist

Overnight urinary cortisol excretion

An integrated measure of 12-hr HPA axis activity

Overnight urinary noradrenalin and adrenalin excretion

Integrated indices of 12-hr sympathetic nervous system activity

D. Implications of Allostasis in Adaptation

Allostasis is a theory that suggests adaptation to external challenges through changes in the behavioural or physiological responses as influenced by individual differences. A child's ethnicity and environment can influence behavioural and physiological responses. The interrelated activity between multiphysiological systems (i.e. PNS and SNS) when responding to external challenges allows individuals to adapt. While behavioural responses may serve externally valid outcomes, its vulnerability to subjectivisms has been questioned. Measures of physiological responses using neurophysiological autonomic measures are suggested due to their validity and efficacy. Thus, in examining how individual differences related to ethnicity and environment, the use of neurophysiological measures is highly suggested.

IV. Adaptation to External Sensory Stimuli

A stimulus is a measurable change in the external or internal environment (Strickland, 2001). Sensory information is processed by the central nervous system (CNS). But it can also elicit system-wide responses throughout the body, such as the fight-or-flight response of the SNS.

There are five basic sensory stimuli emanating from the environment: tactile, visual, olfactory, gustatory and auditory. The auditory stimulus is characterised by sounds from the external environment and is transmitted to ossicles. The sound is then transmitted to the inner ear through by way of vibrations.

A. Features of Auditory Stimuli

Auditory stimuli cause pressure changes that reach the external ear and continues to the tympanic membrane that articulates with the auditory ossicles of the middle ear. Such pressure fluctuations are multiplied as it continues down the cochlea of the inner ear. The hairs of the organ of Corti moves along with the fluid and membrane as the sound travels to the cochlear chambers. The process is monitored by bipolar sensory neurons on the cochlear centre, which consequently passes information to the brainstem by way of the cranial nerve VIII. Auditory information is ultimately processed in the primary auditory cortex (Martini, Nath & Bartholomew 2010). The minimum level of sound (pure tone) that a human can hear within the normal hearing capacity and devoid of any other sound present, is referred to as the absolute threshold of hearing (Durrant & Lovrinic, 1984; Gelfand & Levitt, 2004). For this to happen, auditory stimuli must meet a certain threshold, however, this is not a discrete point. Thus, the auditory threshold can be considered margin at which an auditory stimulus can elicit a response for a period of time (Durrant & Lovrinic, 1984). The range by which humans can audibly hear an auditory stimulus has been suggested to be from 20 Hz to 20,000 Hz (Self, 2012). The range is dependent on the individual's previous exposure to noise, age, health, etc. Furthermore, it was suggested that ear sensitivity occurs at the 1000-5000 Hz range, specifically for sounds at 4000 Hz (Porter, 2011). The threshold for the safety of loudness of auditory stimuli is set at around 85 dB (NIOSH, 1996), at a constant exposure of no more than eight hours. Table 2.2 presents a set of recommended octave-band sound pressure levels.

The auditory cortex has extra-stimulus functions. Previous studies demonstrated how auditory stimuli are able to reflect autonomic responses among individuals (Santana, Martiniano, Monteiro, Valenti, Garner, Sorpreso & de Abreu, 2017). For example, atypical autonomic responses (i.e. diminished habituation, decreased resting-state levels) to auditory stimuli is related to social functioning (Järvinen et al., 2016; Perdue et al., 2017). Thus, the auditory cortex has extra-auditory and extra-sensory involvement processes associated with behavioural implications (Rutkowski & Weinberger 2005). The response to auditory stimuli has been suggested to reflect underlying autonomic regulation of the PNS (Berntson, Cacioppo & Quigley, 1991; Berntson et al., 1997; Gomez et al., 2017; Matsushima et al., 2016; Schauder & Bennetto, 2016; Sokhadze et al., 2016) and SNS (Dawson, Schell & Fillion, 2000; Oji-Mmuoa, Gardnera & Dohenya, 2018; Sokhadze, et al., 2016).

Table 2.2. Set of Recommended Octave-band Sound Pressure Levels (source: http://personal.cityu.edu.hk/~bsapplec/single.htm).

Recommended octave-band sound pressure levels	
Band Center Frequency (Hz)	Sound Pressure Level (dB)
31.5	74
63	66
125	71
250	61
500	60
1000	75
2000	82
4000	80
8000	87
16000	90

B. Subcortical System and Auditory Stimuli

Traditionally, the auditory cortex has been implicated for its role as acoustic analyser; however, more recently, the auditory cortex has been suggested to play a role in learning-induced plasticity through facilitative sensory analysis (Weinberger, 2012). The auditory environment can likewise have an effect on brain regions (i.e. limbic system) traditionally having non-auditory functions. Specifically, the amygdala and hippocampal regions of the limbic system can receive input from the central auditory system in a direct or indirect manner (LeDoux, 2007; Winer & Lee, 2007; Munoz-Lopez, Mohedano-Moriano & Insausti, 2010). Therefore, auditory

stimuli may functionally stimulate the limbic system. This connection provides evidence on the role of auditory stimuli in influencing neuronal activity or modulate plasticity (Kraus & Canlon, 2012; Marsh et al., 2002).

Evaluating whether sensory information is important or not is a central role of the amygdala (Garrido, Barnes, Sahani & Dolan, 2012). Traditionally, the amygdala has been linked to emotion processing (Phelps & LeDoux, 2005; Rasia-Filho et al., 2000), such as happiness or fear and anger (Baxter & Murray, 2002; Davis & Whalen, 2001). However, the amygdala is likewise capable in regulating the auditory cortex and plasticity through cholinergic projections of the nucleus basalis located in the basal forebrain (Keuroghlian & Knudsen, 2007; Nelson & Mooney, 2016). The amygdala receives direct communication with primitive sensory systems through subcortical pathways. This direct input has been suggested to support the adaptive and rapid appraisal of salient sensory information in a cortical manner (Garrido et al., 2012; Mendez-Bertolo et al., 2016).

The direct and indirect dual route model to the amygdala (Garrido et al., 2012) forms parallel cortical and subcortical pathways. Specifically, the subcortical route has been suggested to better explain physiological responses to salient sensory (i.e. auditory) information. Furthermore, the subcortical pathway has been implicated in its importance in rapid stimulus appraisal and processing of sensory stimuli (Garrido et al., 2012). In certain pathologies (i.e. anxiety disorders) the cortical pathway is bypassed, and sensory stimuli are processed via the subcortical pathway prior to reaching the amygdala. This subcortical pathway processes auditory stimuli in a different manner, recruiting the amygdala's emotional role in the context of fear, producing aversive reactions.

C. Behavioural Response to Sensory Stimuli

Behavioural responses are observable reactions to external sensory stimuli, which has previously been suggested to be impaired among childhood clinical populations resulting in symptoms or issues related to sensations (Dunn, 1999; Lane & Schaaf, 2010; Schauder & Benetto, 2016). Parent/caregiver reported measures has long been used in examining sensory behavioural differences. Individual differences (i.e. developmental stages, gender, ethnicity and environment, etc.) influencing behavioural regulation of responses to sensory stimuli was previously suggested.

(Caron et al, 2012; Leekam et al., 2007; Tirosh et al., 2003). More apparent is the body of literature examining the difference between children with and without clinical conditions (Bundy et al., 2007; Ghanizadeh, 2011; Tomchek & Dunn, 2008). However, behavioural evidence may not be as objective, and the information may be confounded with parental factors (Hoyle, Harris & Judd, 2002; Ooi et al., 2016; Orekhova & Stroganova, 2014). Utilizing direct observations (i.e. use of physiological outcomes) may be a sounder choice (Baranek et al., 2007; Schoen et al., 2008).

D. Physiological Response to Sensory Stimuli

In responding to an external stimulus, internal physiological parameters deviate from the normal set standards as a form of adaptation; this is referred to as a physiological response (Matthews, 1986). The use of physiological measures has traditionally been used to measure functions of the CNS and ANS (McEwen, 2008). Specifically, parasympathetic and sympathetic measures were suggested as indices of physiologic regulation of response to sensory stimuli (Schoen et al., 2009).

i. Parasympathetic Response to Sensory Stimuli

Measures of parasympathetic nervous system activity levels include cardiac vagal tone (VT), respiratory sinus arrhythmia (RSA), and HRV. While different in terminologies, they all refer to the variation in heart rate and time in between heartbeats influenced by cranial nerve X, the vagus. Such time in between heartbeats is also referred to as inter-beat intervals or R-R intervals (Kandel et al., 2000; Porges & Byrne, 1992), commonly used by researchers interested in the varying components contributing to heart and respiratory rates.

Previous studies have indicated that cardiac measures are sensitive to responding to auditory stimuli (Gomez et al., 2017; Lydon et al., 2016). For example, Sokhadze, Casanova, Tasman & Brockett (2016) used cardiac measures including heart rate and heart rate variability in determining autonomic responses to auditory stimuli among children ages 8-14 yrs. The researchers found a consistent parasympathetic response to the auditory stimuli among the children over repeated trials. Similarly, Matsushima et al. (2016) found significant lower parasympathetic cardiac

activity indexed by high-frequency component of HRV among children with autism as compared to a typically developing group. The foregoing studies imply that the PNS, specifically cardiac-related autonomic measures, are sensitive to respond to auditory stimuli.

Schaaf et al. (2003, 2010, 2015) carried out several studies that used vagal tone measures to determine if the PNS activity is a significant biomarker in sensory responding among children. Using RSA as an index of cardiac VT tone representing the activity levels of the PNS, children are subjected to the Sensory Challenge Protocol¹ (SCP; Miller et al., 1999). SCP is a non-invasive experimental paradigm that evaluates a child's physiologic reactivity to a series of sensory stimuli. The protocol has been used successfully in several studies and is described much further in the literature (McIntosh et al., 1999a, 1999b; Miller et al., 1999, 2012; Schaaf et al., 2003, 2010, 2015).

RSA was collected using ECG by using three Silver-Silver Chloride (Ag/AgCl) electrodes positioned on the chest of a child, with the ground lead located at the midline of the stomach while the other two lead at the axilla level with the heart. Data analysis was performed using PSYLAB software (Contact Precision Instruments, 2002) and MXEdit (Delta Biometrics, 1993) allowing artefact detection and reduction, and frequency domain analysis within the respiratory range using a moving polynomial filter. The parasympathetic response was indexed by averaging the cardiac VT across 30 s epochs across the SCP conditions (baseline, sensory domain stimulation, recovery, and prolonged auditory stimulation).

1

¹ There are four phases in the SCP: (1) a baseline period of three min where the child is resting and seated quietly in a chair; (2) the administration of a set of sensory challenges (consists of different sensory stimuli in seven domains, consecutively repeated for eight times each), (3) a recovery phase where the child returns to quiet sitting for a period of three minutes, and (4) an exposure to a two minute prolonged auditory tone (post-recovery condition). The different stimuli include: auditory tones (84 dB), visual (strobe lights of 20 watts at 10 Hz), auditory siren sound (78dB), olfactory (oil of wintergreen scent passed under the nose), tactile (moving touch along the jawline from one right to left using a feather), and vestibular (chair is suddenly tilted back to a 30° angle). After recovery, a two-minute prolonged auditory tone (75 dB) was administered. All stimuli, except for the prolonged auditory stimuli, occurred for three seconds with a pseudo-random interval of 12–17 second of interstimuli interval. By using the SCP, researchers can describe the PNS activity levels at resting, during stimulation and recovery phases while determining associations between different sensory stimuli.

Schaaf et al., (2003, 2010, 2015) et al suggests associations between cardiac VT and behavioural measures. For instance, when behavioural and cardiac VT are compared between typically developing children and children autism, they found lower cardiac VT levels among the latter (Schaaf et al., 2003). This suggests that typically developing children have more effective parasympathetic functioning compared to those with sensory behaviour problems. Their hypothesis was further tested with another study (Schaaf et al., 2010) where similar results were reported. Children who exhibited problems in sensory regulation (i.e. autism spectrum disorders (ASD), ADHD, etc.) had lower baseline VT that approached significant levels. These findings are consistent with the body of literature that suggests lower VT is associated with poor behaviour regulation, whereas high resting (i.e., baseline) VT is related with positive temperament and improved regulatory behaviours (Hardaway, Wilson, Shaw & Dishion, 2012; Schaaf, 2010). These data seem to suggest that the neurophysiological index of the PNS as measured by the cardiac VT of RSA is associated with a child's capacity to regulate responses to sensory stimuli. However, respiration greatly influences the RSA thus, establishing the vagal inputs in RSA might be difficult to establish (Grossman & Taylor, 2007). Moreover, the RSA has been criticised on its ability to consistently index vagal inputs (Grossman & Taylor, 2007). Therefore, a different neurophysiological index of the PNS (i.e. high-frequency band of heart rate variability) can be suggested as another alternative.

ii. Sympathetic response to sensory stimuli

There is some evidence that looks at the role of the sympathetic nervous system (SNS) in its ability to index regulation of response to sensory stimuli. Of interest is using electrodermal activity (EDA) as a neurophysiological index to demonstrate children's responses to various sensory stimuli. Emotional arousal (i.e. from sensory stimulation) activates the ANS from the SNS branch causing sweating with consequent salt and water concentrations, thereby increasing electrodermal conductivity of the skin to electrical current. Central neural circuits originating from the prefrontal cortex and limbic structures have been

postulated to mediate the electrodermal response via the SNS (Critchley, 2002).

Previous studies found that using auditory stimuli can elicit a physiological response from the SNS among children (Widmann, Schröger & Wetzel, 2018). For example, Oji-Mmuoa, Gardnera and Dohenya (2018) found that using auditory stimuli can reflect sympathetic regulation by indexing the skin conductance response among neonates. The researchers found that higher sympathetic arousal (as measured by skin conductance response) in response to an auditory stimulus persists at discharge among children exposed to opiates. Another study likewise used electrodermal activity measures (skin conductance level) to index autonomic responses to auditory stimuli among children ages 8-14 yrs. (Sokhadze, Casanova, Tasman & Brockett, 2016). The researchers found a consistent sympathetic response to the auditory stimuli among the children even when given repeated trials. Measures of EDA (i.e. skin conductance response, skin conductance level) are sensitive in responding to auditory stimuli among children.

Research using EDA that is of interest in responding to sensory stimulus has primarily targeted the study of children's sensory regulation. It was hypothesized by Miller et al. (1999, 2003, 2007, 2012) that the SNS is responsible for the regulation of reactivity and recovery from an external challenge, specifically, sensory stimuli. To better understand the association between ANS functions and behavioural responsiveness to sensation in children, Miller and colleagues studied SNS physiologic functioning in children using the SCP (McIntosh et al., 1999a, 1999b; Miller et al., 1999, 2012; Schaaf et al., 2003, 2010, 2015). Behavioural measures were taken using the Sensory Profile (Dunn, 1999) among all their experiments.

Results in their studies (Miller et al., 1999, 2003, 2007, 2012) are uniform in their findings. Between typically developing children and children with probable sensory regulation problems (i.e. Fragile X syndrome, autism, ADHD) results suggest significant differences in the SNS levels between

two populations. EDA responses in one sensory modality are strongly predicted by responses in the other sensory modalities. Furthermore, underlying physiological enhancement of SNS levels is exhibited as overreactivity to sensory stimulus in daily patterns of activities as measured by the caregiver version of the Sensory Profile (Dunn, 1999). Results from these researchers, therefore, suggest that regulation of sensory behaviours is dependent on the integrity of the SNS activity levels among children.

E. Implications of the Regulation to Response to Sensory Stimuli in this Research

Adaptation through the regulation of neurophysiological activity in response to sensory stimuli allows children to master their environments. The auditory stimuli have been implicated in its ability to stimulate autonomic responses related to PNS and SNS functions. Autonomic activity in response to sensory stimulation such as an auditory stimulus measured at resting, stimulation and recovery conditions may reflect the underlying neurophysiological regulation mechanism. In this research, an auditory stimulus is used to index the ANS responses.

Neurophysiological measures related to the functions of the ANS have been suggested by various authors to index the regulation of response to sensory stimuli. Several neurophysiological measures have been used in previous researches. To represent the PNS functions, the more common neurophysiological measure has been heart rate variability. To represent SNS functions, EDA was suggested. However, the evidence rarely examined both measures at the same time. The mechanism that enables adaptation to external challenges such as sensory stimuli entails examining both measures to represent the dynamic and interrelated relationship between the PNS and SNS is proposed.

- V. Ethnicity and Environment Influences the Regulation of Response to Sensory Stimuli
 - A. Ethnicity and the Regulation of Response to Sensory Stimuli

Ethnicity refers to the biomedical origins and genetic similarities among individuals (Ali-Khan et al., 2011; Jorde & Wooding, 2004). The genetic makeup of humans strongly influences their behaviours, and such behaviours could likewise moderate transmission of behaviour and genetic makeup of a child (Klahr & Burt, 2014). The

response to sensory stimuli seems to be stable and a universal phenomenon. For example, Royeen and Mu (2003) conducted a study to examine the phenomenon of tactile defensiveness across cultural groups. They investigated the stability of the concept across different cultures, using samples of European and American children 6-11 years old living in different countries. In their study, the Touch Inventory for Elementary School Children (TIE) was used. TIE is a parent report behavioural checklist that investigates the tactile hyperresponsiveness of children. The results of Royeen and Mu (2003) indicate that there is no significant difference between the two samples.

A similar study was done by Tirosh, Bendrian, Golan, Tamir and Dar (2003), to identify sensory regulatory problems among children of different ethnicities, particularly Arabic and Jewish children, that lives in the same country. Tirosh et al. (2003) developed a questionnaire that they used for their study. It was revealed by ethnic-cultural effects that the sensory regulation is almost identical in the two ethnic groups of infants, as well as among Hebrew and Russian speaking Jewish participants. Furthermore, they also found that in the older group, there is a significant ethnic difference. It was suggested that such a difference can be due to underlying physiologic regulatory mechanisms across ethnic origins. Among the older age group of children, attitude development as suggested by their temperament behaviours is likely affected by socio-cultural influences. It can then be assumed that there is an underlying physiologic mechanism in sensory regulatory disorders that affects older children across ethnic origins. However, it is unclear whether such differences in the regulation of response to sensory stimuli is due to ethnic differences or parental contexts (i.e. parental education and behaviours) since a parent-answered behavioural measure was used.

Behavioural studies on the influence of ethnicity on the regulation of response to sensory stimuli yield inconsistent and inconclusive results. It is unknown whether differences in the response to sensory stimuli can be explained by a child's ethnicity or the environment in context. The knowledge gap, therefore, lies in the inconsistency of using behavioural measures and lack of control for environmental variables. This research examines a group of children from different ethnicities (with the environment controlled) and tries to understand their regulation of response to sensory stimuli from a neurophysiological perspective.

B. Environment and the Regulation of Response to Sensory Stimuli

The environment is the geographic range where a group of individuals exists (Lee & Minard, 2013). Similar socio-cultural traits and experiences embedded within similar geographic environments result into behavioural responses that are shared by a category of people that set apart one group of people from another (Robinson-Wood, 2016; Zimmerman & Woolf, 2014). The socio-cultural contexts and experiences within the geographic environments have been implicated to account for differences or associations between individuals from different or similar geographic environments (Caprio et al., 2008). The geographic environment where a child develops has the ability to override ethnic influences on responding to external challenges (Gunn et al., 2009). Response to external challenges may have been likely shaped by the experiences within a child's environment. In this thesis, the environment is classified into two levels: geographic environments and physical environment.

Children's responses to sensory stimuli can be influenced by the physical environment where they live in (Brown & Dunn, 2010). The sensory environment and the opportunities in the environment generally affect the brain and its functions (Kempermann, van Praag & Gage, 1999). However, the role of the physical contexts of environments, specifically between different contexts such as urban and rural settings, on the response behaviours of children to environmental challenges such as different sensory stimuli is still unknown.

C. Ethnicity and Environment and the Regulation of Response to Sensory Stimuli

Looking at both the ethnicity and environmental variables could give significant information on how children respond to sensory stimuli. Caron, et al. (2012) studied the influence of culture and the environment on the sensory behaviours of children by comparing the regularity of caregiver-reported responses to sensory information in daily activities between children from Israel and the United States. There are two groups in this study, 54 normative group of children (28 from Israel, 26 from the United States) and 57 clinical group of children with ASD (37 from Israel, 20 from the United States). The Hebrew version of the Short Sensory Profile (SSP; Caron et al., 2012; McIntosh et al., 1999) and a 38-item version of the Sensory Profile (Dunn, 1999) were used in the study. The z-scores of children who are typically

developing in both countries were within normal ranges; however, scores of the normative group from Israel were significantly greater on four of the seven sections when compared their American counterparts. Unusual responses to sensory experiences were reported by fewer caregivers of Israeli children. It is possible that the differences from the findings suggest either that participants that are typically developing from Israel may have a less intense behavioural response to sensory experiences than those from the U.S., or Israeli caregivers may likely rate a behaviour as not significantly different from the usual. Super and Harkness's (2002) proposed three operational subsystems comprise a child's developmental niche. These are: (1) the social and physical settings, (2) the ingrained practices and customs of childcare, and (3) caretaker's psychology (Super & Harkness, 2002). A specific cultural niche is created from the interaction of these subsystems that organizes the daily environment of a child, thus influences development.

In this study, the physical environment is referring to empirical aspects of the physical setting, which includes tangible natural or man-made structures and/or components (McDonell & Pickett, 1990). The physical environment is represented herein by the contrasting characteristics of urban and rural settings. Perloff (2015) suggested specific operationalisations between such urban and rural settings based on tangible dichotomous characteristics between the two. The urban setting is described to have metropolitan characteristics of a developed setting where manmade structures abound (i.e. houses, buildings, bridges, railways, etc.). On the other hand, the rural setting is an unspoiled region of natural environment (i.e. mountainous, coastal, agricultural) populated with fewer human inhabitants and man-made establishments.

The ability to regulate one's responses to challenges from the external world is influenced by their physical environments (Rutter et al., 2001). In comparing urban and rural environments, it was found that the latter produces less physiological arousal and attentional demands (Laumann, Garling, & Stormark, 2003; Ulrich et al., 1991). Among Taiwanese children, Lin (2004) suggested that children living in urban and rural areas may have differences in the regulation of responses to sensory stimuli. Tirosh et al. (2003) similarly found urban-rural differences and specifically suggested socio-cultural influences (i.e. cultural differences, maternal education) as factors that can moderate the regulation of response to sensory stimuli in children

living in rural settings. Conversely, it was suggested that the physical features of children's dwelling may influence improved regulation of response to sensory information such as seen among children living in urban environments who exhibits significantly higher abilities in cognitive performance, perceptual-motor, and perception. However, it remains unknown whether variations in physical environments can influence the response towards sensory stimuli.

D. Summary on Ethnicity and Environment's Influence on the Regulation of Response to Sensory Stimuli

The finding of previous researches on the influence of ethnicity and environment appears inconclusive (Gomez et al., 2018). Some researchers suggest that the ability to regulate responses to sensory stimuli is universal and constant despite individual differences due to a child's ethnicity and environments (Chang et al., 2012; Royeen & Mu, 2000). However, others suggested that both ethnicity and environment have influences on the ability to regulate responses to sensory stimuli among children (Caron et al., 2012; Tirosh et al., 2003). One limitation of previous studies is the uncontrolled environment where data on the response to sensory stimuli was measured (Gomez et al., 2017; 2018). This thesis, therefore, aimed to employ an experimental design to examine the regulation of response to sensory stimuli from a neurophysiological perspective in regard to ethnicity and environments in children. Furthermore, methodological limitations pertaining to the choice of behavioural checklists/questionnaires as a measurement instrument is observed. Behavioural outcomes (i.e. parent reports, clinical observations) have a limitation in providing precise and objective data supporting the regulation of response towards sensory stimuli (Gomez et al., 2017; Schoen et al., 2009). Previous research has found that sympathetic (SNS; McIntosh et al., 1999) and parasympathetic (PNS) autonomic functions (Schaaf et al., 2003) are associated with the ability to regulate responses to sensory stimuli. Measuring the underlying neurophysiological mechanism in response to sensory challenges offers a more sensitive and objective method (Appelhans & Luecken, 2006).

VI. Synopsis of the Autonomic Nervous System

Arising from the peripheral nervous system is the autonomic nervous system (ANS) implicated in its role in physiologic homeostasis. It is comprised of sympathetic and

parasympathetic branches. Their dynamic relationship has been described to support regulatory mechanisms responsible for adaptation to external challenges. Regulation of the ANS is maintained by way of top-down control from higher brain centres. However, the ANS has likewise been implicated in bottom-up regulation that influences higher-order functions such as behaviours.

A. Anatomy and Physiology of the Autonomic Nervous System

The human nervous system consists of two major divisions: the central and peripheral nervous system divisions. The central nervous system (CNS) has various centres, subdivided into lower centres (including the spinal cord and brain stem) and higher centres (communicating with the brain via effectors), that carry out the sensory, motor and integration of data. The peripheral nervous system, on the other hand, is a complex network of cranial and spinal nerves, linked to the brain and the spinal cord. It carries sensory receptors, which supports processing event changes within one's external and internal environments. Afferent sensory nerves then carry this information to the CNS. The peripheral nervous system is then subdivided into the autonomic nervous system, responsible for involuntary control of cardiac and smooth muscles, internal organs and blood vessels; while the somatic nervous system control s voluntary control of skeletal muscles, joints, bones, and skin. These two systems function together. Anatomically, the peripheral nervous system may be thought of as an extension of the CNS as the functional control centres for the peripheral nervous system lie in the CNS (Everly & Lating, 2012). Central control of the ANS mostly comes from the hypothalamus, with some input from the reticular activating system and the limbic system.

The ANS can be further subdivided into the sympathetic and parasympathetic branches, which are different functionally, anatomically and physiologically. The sympathetic branch is responsible for organising the body for action, with its generalized arousal effect on the organs it innervates. The parasympathetic branch is concerned with restorative functions and the relaxation of the body, thus slowing and maintaining the body's basic bodily requirements. These branches of the ANS are often mentioned as a pair to function together but in an opposite manner.

i. Sympathetic Nervous System

The adrenal gland is supplied by the sympathetic nervous system. The adrenal medulla is regarded as a specialized synapse, which releases transmitters (adrenaline and noradrenaline) directly into the bloodstream. Direct neural connections and noradrenergic effects of eth endocrine systems mediate the effects of the SNS.

The neurotransmitter acting between the SNS and its target organs (except for acetylcholine-mediated sweat glands) is noradrenaline, further categorized as α or β . Noradrenaline α receptors found in blood vessels promote vasoconstriction. On the other hand, noradrenaline β receptors can be found in the heart (β 1) where it is responsible for stimulating rate and force of contraction; and the smooth muscle of the uterus (β 2) and the bronchial tree where they cause relaxation

As spinal sympathetic motor neurons SNS nerves exit spinal cord, preganglionic efferent emerge in the ventral roots running along the periphery of the cord where it enters an interconnected set of sympathetic ganglionic chain. Upon entry, these preganglionic fibres may ascend or descend prior to termination at the sympathetic ganglion cells giving rise to postganglionic axons where it projects to various visceral organs.

Acetylcholine is the primary neurotransmitter involved at the ganglionic synapse, which becomes the catecholamine neurotransmitter noradrenaline (norepinephrine) at the ganglionic synapse. However, this is not the case for all, such as in the case of direct preganglionic fibre innervation at the adrenal medulla, where secretory cells release the catecholamines act humorally on many organ systems as it is generally released in the general circulation including to visceral organs that do not receive direct innervation.

Due to the various peripheral actions of the SNS that are characterised to be activational and promoting the metabolism of energy, the SNS has been implicated in its role to mobilize the internal systems in adapting to external challenges. For example, previous studies have demonstrated the

neurophysiological sensitivity of the SNS in responding to auditory stimuli (Oji-Mmuoa Gardnera & Dohenya, 2018; Sokhadze, et al., 2016) by looking at the electrodermal activity. The sweat glands found in the subdermal layer of the skin is innervated by the SNS. At resting conditions, relatively constant electrodermal tonic activity can be measured in the skin surface. When an auditory stimulus is presented, several physiological events happen similar to the description in the previous paragraphs. The presentation of such auditory stimuli causes sympathetic activity, which in turn activates the sweat glands producing sweat on the surface of the skin as a psychophysiological response (Dawson, Schell & Fillion, 2000; Oji-Mmuoa Gardnera & Dohenya, 2018; Sokhadze, et al., 2016). Hand sweating is a common regulatory response of the SNS to an external challenge (Boucsein, 2012; Dawson, Schell & Fillion, 2000).

Sympathetic functions are responsible for preparing the body in dealing with danger or threat: the "fight or flight" response. Blood is directed to the brain, heart, and muscles, and diverted away from the non-vital areas, while increased heart rate and respiration occur, to meet extra demands for blood and oxygen. Figure 2.2 illustrates the distribution of the SNS.

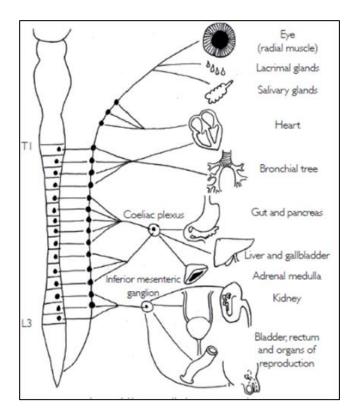


Figure 2.2. Distribution of the SNS (Reprinted with permission from O'Donnell, A., & Glasgow, B. (2011). The autonomic nervous system. *New Zealand Medical Student Journal*, (13)).

ii. Parasympathetic Nervous System

The parasympathetic nervous system (PNS) consists mostly of fibres which travel in cranial nerves (CN) III, VII, IX, and X. However, a minute partition is subserved from sacral nerve roots S2, 3 and 4. Taken together, the major player in PNS functions is the vagus nerve (CN X), supplying vagal signals to almost all the organs in thorax and abdomen. Throughout its process, the neurotransmitter is acetylcholine. There are many and complex functions of the vagus nerve; only parasympathetic functions are described here.

Arising from the medulla oblongata, the vagus nerve exits the skull through the jugular foramen and travels caudally as a bundle of neurovascularities (i.e. common carotid artery, internal jugular vein). The vagus nerve crosses the diaphragm while traversing parallel to the esophagus on both sides in the area of the mediastinum. As it moves downward, the vagus nerve subserves the coeliac arterial trunk below the diaphragm along with sympathetic

fibres. These form a plexus which passes throughout the abdomen where fibres extend through most of the viscera.

The PNS includes a peripheral ganglion. Generally found near their target visceral organs. This unique configuration allows the PNS direct localized action compared to the regional effect of the SNS. However, similar to the SNS, ACh is the neurotransmitter found in preganglionic axons acting primarily via nicotinic cholinergic synapses. Their difference lies in the neurotransmitter at the postganglionic axons. AcH in the PNS acts through muscarinic cholinergic receptors. Such distinguishing configuration in the neurochemical coding between the PNS and SNS reflects the complexity of neurotransmitter, neurohormonal, and neuromodulatory interactions within the ANS.

Previous studies have implicated the responsiveness of the PNS to external challenges such as in the case of auditory stimuli. Specifically, cardiac autonomic measures have been demonstrated to be good measures of physiological responses among children (Matsushima et al., 2016; Sokhadze et al., 2016). The presentation of an external challenge such as auditory stimuli to an individual is regulated by the PNS. Several internal processes occur, which causes the PNS to lower or withdraw its activity in order to allow for the SNS act and respond (Berntson, Cacioppo & Quigley, 1991; Berntson et al., 1997). Among cardiac autonomic activity, a decrease in the heart rate and the high frequency component of HRV (i.e. CVT, HF) has been seen suggested as parasympathetic regulation in response to external challenge to promote adaptation (Berntson, Cacioppo & Quigley, 1991; Berntson et al., 1997; Gomez et al., 2017; Schauder & Bennetto, 2016).

Physiologically, the functions of the PNS can be thought of as "rest and digest," which includes defecation, peristalsis, production of digestive juices, and salivation. The PNS regulates at the level of the organs and glands that it innervates during rest and has been described to be slow in activating, dampening the internal physiological systems to conserve and restore the energy of the body. Figure 2.3 illustrates the distribution of the PNS.

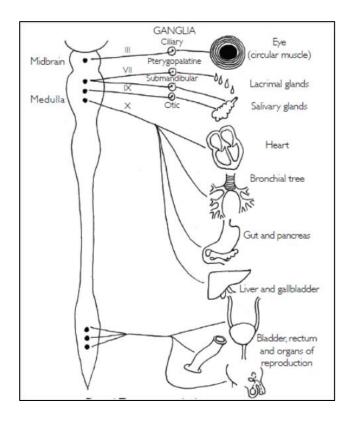


Figure 2.3. Distribution of the PNS (Reprinted with permission from O'Donnell, A., & Glasgow, B. (2011). The autonomic nervous system. *New Zealand Medical Student Journal*, (13)).

B. Measures of Autonomic Functions

When choosing a tool in measuring the impact of an intervention upon ANS regulation, the primary consideration is whether it measures an activity which will be sensitive to the expected change. The instrumentation must be responsive to minute changes in activation and allow precise measurement. Stable baseline measurements are equally important to assess the patterns and intensity of changes. ANS activity is most susceptible to various environmental and biological influences (e.g. temperature, blood pressure) and may provide vague information about the impact of the technique. In such cases, non-invasive cardiac and skin conductance measures offer the most promising and most frequently used measures of the effect of regulation of the ANS (Gomez et al., 2017; Gomez et al., 2018).

i. Heart Rate Variability

a. Overview of Heart Rate Variability

Heart rate variability (HRV) refers to the beat-to-beat fluctuations in heart rate (Berntson et al., 1997). This beat-to-beat variation in HR can reflect cardiac autonomic activity and has been considered as a useful non-invasive index of neuropathy of cardiac autonomic functions (Cornforth, Jelinek & Tarvainen, 2015; von Borell et al. 2007). Figure 2.4 illustrates a typical R-R interval.

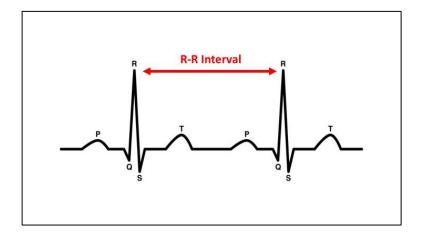


Figure 2.4. R-R interval.

HRV is the degree of fluctuation variances in the length of a heartbeat to heartbeat intervals (Malik & Camm, 2004). To an extent, HRV reflects the regularity of heartbeats where the larger the regularity, the lower the HRV, and vice versa. HRV is computed from elapsed time differences in consecutive R-R intervals, measured in millisecond (ms). This data is commonly obtained using ECG or plethysmography. Figure 2.5 illustrates a typical R-R interval tachogram.

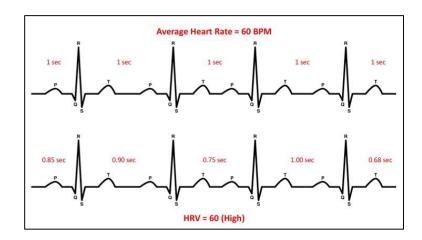


Figure 2.5. R-R interval tachogram.

b. Physiological Mechanisms of HRV

The physiological mechanism responsible for HRV entails a series of events controlled by the sympathetic and parasympathetic branches of the ANS. At the resting phase, heartbeat frequency is produced by the primary impulse generating tissue (pacemaker), the Sino-atrial node (SA-node). The rate of the non-innervated SA-node in itself is variable and ranges between 60-80 bpm (Sammito et al., 2015; Weaver & Polosa, 2006). It is usually higher in children (Shim, Park, Moon, Lee, Kim, Han & Lee 2014). The SA-node also has other subordinate nodes that are capable of spontaneous depolarisation with lower rates – AV-node, Bundle of His, & Purkinje fibres. The pacemaker primarily facilitates in the autonomic modulation of HR by the SNS and PNS. Such dual control by the ANS was demonstrated in various experiments, such as the use of sympathetic blockade by propranolol, as well as vagus blockade by atropine (Castiglioni, Parati, Di Rienzo, Carabalona, Cividjian & Quintin, 2011; Reyes del Paso, Langewitz, Mulder, van Roon & Duschek, 2013). Figure 2.6 illustrates a summary of the physiological mechanism of HRV.

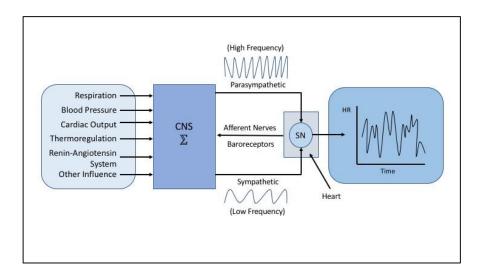


Figure 2.6. Physiologic mechanisms of HRV.

The sympathetic branch typically decreases the absolute HRV by shortening the NN-intervals, with noradrenaline as its neurotransmitter at the SA-node. In contrast, the parasympathetic branch typically increases absolute HRV, with acetylcholine as its neurotransmitter (Andreassi, 2013). The parasympathetic (vagal) control outweighs the sympathetic effect on the HR at rest and during mild exertion, leading to increased variability of the heartbeat. Thus, the gap difference between two consecutive heartbeats increases.

Cardiac response to sympathetic stimulation is relatively slow, taking about 5 seconds post-stimulation before an increase in HR is seen, peaking at 30 seconds. In comparison, parasympathetic response to HR is instantaneous, taking one or two heartbeats before slowing down to its minimum in proportion to stimulation level, and dependent on the heart cycle's actual phase.

Normalising of HRV data (normalized units) has previously been suggested to as a reliable marker of autonomic cardiac control (Malliani et al., 1991; Malliani, Lombardi & Pagani, 1994; Montano et al., 2009). Sympathetic activation has been associated with a decrease in total HRV power (including the Low Frequency (LF) component), while the opposite occurs with vagal activation. Changes in the Total spectral power results in a distorted estimate of

LF and HF power when spectral components are expressed in absolute units (ms²). In order to avoid this, the LF/HF ratio or normalized units have been suggested when examining sympathetic cardiac tones. To derive the normalized units, the power of a given component (HF or LF) is divided by the total power minus the Very Low-Frequency (VLF) power.

HRV indices present a wide array of applications the requires easy and non-invasive methods of investigation of autonomic functions. It has previously been shown that altered HRV is present in various cardiovascular diseases implicating its role in diagnostics and prognostics (Thayer & Lane, 2007). However, the validity of using the LF or the LF/HF ratio as an index of sympathetic functions I debatable regardless of adjustment in the total power or normalization (Eckberg, 1997; Goldstein et al., 2011; Kleiger, Stein, & Bigger, 2005; Malliani, Pagani, Montano, & Mela, 1998; Malpas, 2002; Parati, Mancia, Di Rienzo, & Castiglioni, 2006; Taylor & Studinger, 2006). Nevertheless, the LF band is still used extensively to index the sympathetic autonomic tone.

c. HRV Analysis: Time Domain Methods

There are different methods to analyse HRV (i.e. time domain, frequency domains, non-linear). The simplest form is the time domain analysis, wherein the normal to normal interval sequence is treated as an unordered set of intervals (or pair of intervals) and different techniques are employed to express the variance of such data. Table 2.3 summarises the different HRV time-domain variables.

The SDNN, computed as the standard deviation of all normal R-R intervals (between two consecutive heartbeats), is considered as a straightforward useful metric of HRV. SDNN is reported in units of milliseconds (ms) and is usually measured over 24 hours. Since the values for the accepted normal range are based on the 24-hour record, this should not be the comparison for the results derived from

shorter (or longer) periods of SDNN. The SDNN index and the SDANN index (both with units in ms) are two variants of the SDNN. These arise from the total 5-min segments within the 24-hour monitoring period. The SDANN is the standard deviation of the mean of normalised R-R (NN) intervals in all the 5-min segments that constitute the entire heart rate monitoring period. On the other hand, the SDNN index represents the average of these 5-minute standard deviations of normal R-R (NN) intervals within the same 24-hour monitoring period (i.e. mean of 288 normalized R-R(NN) standard deviations).

The r-MSSD and the pNN50 are other noteworthy time-domain indices. The root-mean-square successive difference (r-MSSD) represents the long-term (24 hrs) square root of the averaged squared differences between successive normal R-R (NN) intervals. On the other hand, the pNN50 is calculated from the percentage of differences that are greater than 50 ms between successive normal R-R (NN) intervals over a 24-hour monitoring period. Measuring comparisons between consecutive heartbeats, the r-MSSD and the pNN50 measures short-term variations in the NN intervals.

Table 2.3. HRV Time-Domain Methods (Reprinted with Permission from Task Force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology. (1996). Heart rate variability. Standards of measurement, physiological interpretation, and clinical use. *European Heart Journal*, *17*(3), 354–81).

Variable	Units Description						
Statistical measures							
SDNN	ms	Standard deviation of all NN intervals.					
SDANN	ms	Standard deviation of the averages of NN intervals in all 5 min segments of the entire recording.					
RMSSD	ms	The square root of the mean of the sum of the squares of differences between adjacent intervals.					
SDNN index	ms	Mean of the standard deviations of all NN intervals for all 5 min segments of the entire recording.					
SDSD	ms	Standard deviation of differences between adjacent NN intervals.					
NN50 count		Number of pairs of adjacent NN intervals differing by more than 50 ms in the entire recording.					
		Three variants are possible counting all such NN intervals pairs or only pairs in which the first or the second interval is longer.					
pNN50	%	NN50 count divided by the total number of all NN intervals.					
		Geometric measures					
HRV triangular index		Total number of all NN intervals divided by the height of the histogram of all NN intervals measured on a discrete scale with bins of 7.8125 ms (1/128 s). (Details in Fig. 2)					
TINN	ms	Baseline width of the minimum square difference triangular interpolation of the highest peak of the					
		histogram of all NN intervals (Details in Fig. 2.)					
Differential index	ms	Difference between the widths of the histogram of differences between adjacent NN intervals measured at selected heights (e.g. at the levels of 1000 and 10 000 samples) ^[21] .					
Logarithmic index		Coefficient φ of the negative exponential curve $k \cdot e^{-\varphi t}$ which is the best approximation of the histogram of absolute differences between adjacent NN intervals ^[22] .					

d. HRV Analysis: Frequency Domain Methods

By analysing the fluctuations in the frequency domain, additional information on the nature of HR fluctuations may be collected. HRV can likewise be decoded into frequency components that comprise the overall variability. Knowledge of how the power (variance) of the ordered NN intervals allocates the frequency functions provided by power spectral density (PSD) analysis. Table 2.4 summarises the various HRV frequency domain variables.

Table 2.4. HRV Frequency Domain Methods (Reprinted with permission from Task Force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology. (1996). Heart rate variability. Standards of measurement, physiological interpretation, and clinical use. *European Heart Journal*, *17*(3), 354–81.).

Variable	Units	Description Analysis of short-term recordings (5 min)	Frequency range
5 min total power	ms ²	The variance of NN intervals over the temporal segment	approximately ≤0-4 Hz
VLF	ms^2	Power in very low frequency range	≤0.04 Hz
LF	ms^2	Power in low frequency range	0.04-0.15 Hz
LF norm	n.u.	LF power in normalised units LF/(Total Power-VLF) × 100	
HF	ms ²	Power in high frequency range	0·15-0·4 Hz
HF norm	n.u.	HF power in normalised units HF/(Total Power-VLF) × 100	
LF/HF		Ratio LF [ms ²]/HF [ms ²]	
		Analysis of entire 24 h	
Total power	ms ²	Variance of all NN intervals	approximately ≤0.4 Hz
ULF	ms^2	Power in the ultra low frequency range	≤0.003 Hz
VLF	ms^2	Power in the very low frequency range	0.003-0.04 Hz
LF	ms ²	Power in the low frequency range	0-04-0-15 Hz
HF	ms^2	Power in the high frequency range	0-15-0-4 Hz
а		Slope of the linear interpolation of the spectrum in a log-log scale	approximately ≤0.04 Hz

There are several spectral methods that have been suggested analysing HRV tachograms. The PSD method of analysis offers information on the spectral distribution of power as a unit frequency. Frequency domain analysis can be achieved through the Fourier transform (i.e. acquiring a series of numbers from the temporal axis). Since specific physiologic mechanisms mediate fluctuations within each band, the PSD is commonly incorporated within a frequency range. The LF band (0.04-0.15 Hz) is has been suggested to be related to both SNS and PNS modulation, while HF band (0.15-0.40 Hz) is almost entirely controlled the PNS. The LF to HF ratio power is often used as a metric of sympathovagal balance, although this remains debatable.

Various authors, initiated by Pagani and his group (Colombo et al., 2014, Malliani, et al., 1991; Montano et al., 2009; Pagani et al., 1986, Reyes del Paso, et al., 2013; Toninelli et al., 2012) have

presented a model in which HRV analysis is applied to evaluate the autonomic balance between the PNS and SNS. The model comprises three core statements: (1) the HF component power can be considered as an index of cardiac vagal tone; (2) the LF component as an index of sympathetic outflow; and (3) the LF/HF ratio as an index of sympathovagal balance (Pagani et al., 1986). Such reflects the relative SNS contribution to HR regulation. However, it has likewise been suggested that the LF may embody both SNS and PNS influences (Berntson et al., 1997; Hakim, Gozal & Gozal, 2012; Japundzic, Grichois, Zitoun, Laude, & Elghozi, 1990; Reyes del Paso et al., 2013; Randall, Brown, Raisch, Yingling, & Randall, 1991; Task Force, 1996). Given these issues, additional information from other measures of SNS (i.e. electrodermal activity) may provide more information on SNS functioning.

ii. Electrodermal Activity

a. Overview of Electrodermal Activity

Electrodermal activity (EDA) represents the variations in the skin's electrical properties consequent to sweat secretion. When a low constant voltage is applied to the skin, changes in skin conductance (SC) that can be measured non-invasively occurs (Fowles et al., 1981). The ease of data acquisition that can index exclusive sympathetic innervation of the eccrine sweat glands contributes to the acceptance and wide use of SC measures within basic and clinical research (Dawson & Schell, 2012; Dawson, Schell & Filion, 2007).

b. The Electrodermal System

The electrodermal system is basically composed of the skin, it's supporting layers and structures (i.e. glands) found within these layers. The three layers (epidermis, dermis, subdermis) that composes the skin provides an adaptive barrier that supports the body in the maintenance of water balance and core temperature. Figure 2.8 illustrates the electrodermal system.

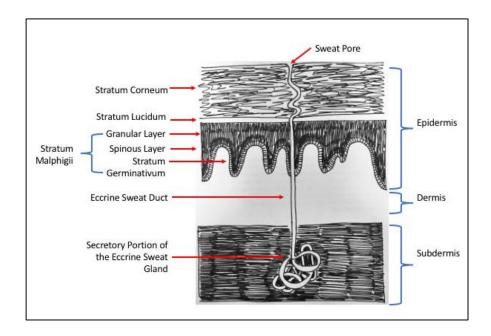


Figure 2.7. The electrodermal system.

There are two forms of sweat glands in the human body: the eccrine, which have been the primary interest in electrodermal activity, and the apocrine. The distinction between these two is usually made based on location and function (Cacioppo, Tassinary & Berntson, 2007). Whereas the apocrine sweat glands are typically found in the armpits and genital areas opening into hair follicles, eccrine glands are found in dense abundance at palms, soles of the feet and throughout the body.

Thermoregulation is the primary function of most eccrine sweat glands. However, those located on the palmar and plantar surfaces have been thought of as being more concerned with explaining behaviour (Cacioppo, Tassinary & Berntson, 2007) and has been suggested to be more responsive to significant or emotional stimuli. Emotion-evoked sweating is usually most evident in these areas primarily because of high gland density (Shields et al., 1987). EDA measurement by researchers in the field of psychophysiology is primarily concerned with these psychologically induced sweating functions of the eccrine glands.

Edelberg (1983, 1992) proposed the sweat circuit model, commonly referred to as the skin conductance model. In this model, sweat ducts found in the epidermis produce phasic changes when filled sweat. Alternately, when sweat is reabsorbed back into the system, a recovery back to the basal tonic levels is observed. Depending on the amount of sweat produced and secreted in the sweat ducts, variations in the amplitude of change in skin conductance is seen. Activation of the sweat gland is a basic physiological mechanism related to adaptation, which has been a keen interest in psychology because it is interpreted as a neurophysiological response to challenges. The mechanism of control is largely attributed to sympathetic functions.

c. EDA Components

The EDA complex is comprised of various components. A summary of the components can be found in Table 2.5.

Table 2.5. EDA Components

Common EDA Components				
EDA Measure	Definition			
Skin Conductance Level	Tonic level of electrical conductivity of the skin			
(SCL)				
Skin Conductance	Phasic changes in electrical conductivity of the			
Response (SCR)	skin			
Non-specific SCR (NS-	SCRs that occur in the absence of an identifiable			
SCRs)	eliciting stimuli			
Frequency of NS-SCRs	Rate of NS-SCRs that occur in the absence of an			
	identifiable stimuli over time			
Event-related SCR (ER-	SCRs that can be attributed to a specific eliciting			
SCR)	stimulus			

The EDA complex has two main components: a tonic component (SCL) and a phasic component (SCR). The SCL is representative of the background EDA, characterised to be slow-acting and commonly seen in resting states. That SCL constantly changes within an individual and can markedly differ between individuals. Baseline skin conductance would be varied between the individual and the changes in their psychological states. The SCL develops a constantly

moving and changing baseline that differs between individuals. The typical tonic basal levels range from 10-50µS (Boucsein, 2012).

The phasic SC changes in response to environmental stimuli are known as SCR. The presentation of a discrete stimulus results to a consequent phasic response that is represented by the SCR. The SCR is described to be a rapid change from the resting state cause by activation of sympathetic activity. Examples of stimuli that may cause an SCR includes movement, odour, sounds or startling visuals. The amplitude of SCR summed with the background of tonic baseline levels (SCL) becomes the phasic skin conductance level. While both components represent importance in autonomic regulation, each is supported by different neural mechanism (Dawson et al., 2002; Nagai et al., 2004).

The microsiemens (μ S) or the micromho (μ hmo) represents the typical EDA units. Both units are considered to be equivalent to one another (i.e. 1μ S = 1μ mho).

d. Standards of EDA Skin Measurement

The Society for Psychophysiological Research recommended standards of electrodermal measurements (Boucsein et al., 2012; Fowles et al., 1981). Table 2.6 summarises the basic EDA standards.

Table 2.6. Standards in EDA (source: Fowles et al., 1981).

Selected Standards of Electrodermal Activity Measurement				
Component	Recommendation			
Measurement of Choice	Skin conductance			
Electrodes	Silver-Silver Chloride			
	Sodium Chloride Paste			
Area of Skin Contact	Double-sided adhesive collars ensure contact			
	are equal to the diameter of the hole in the			
	collar			
Electrode Placement	Thenar or hypothenar eminences of one hand			
	or			
	Medial and distal phalanges of one hand			
Signal Conditioning	Apply constant 0.5 volts across 2 electrodes			
Tonic Level of Control	Subtract portion of tonic SCL to increase			
	sensitivity to smaller phasic responses			

C. Implications of ANS on the Regulation of Response to Sensory Stimuli

The first physiological axis that is activated as a response to environmental challenges is the ANS (Cacioppo, Tassinary, & Berntson, 2017; Everly & Lating, 2012). HRV can reflect activities of PNS, SNS, and their interactions. EDA is a measure of SNS. Issues surrounding the representativeness of the LF bands of HRV as an accurate measure of SNS functions has led this research to use EDA as an adjunct measure.

VII. Summary of Literature and Knowledge Gaps

There is evidence by previous researchers that a child's ethnicity and their living environments can influence how children regulate responses to sensory stimuli. However, methodological variations and the limited inquiry on using more objective measures of the underlying mechanism that enables adaptation through regulation were suggested.

Previous research suggested that the environment could override biologically-embedded traits, possibly related to one's ethnicity, and shape the child's physiological regulation. However, it was rarely examined whether this can be applied to inquiries associated with a child's ability to regulate responses to sensory stimuli.

Prior research evidence suggested the ability of the environment to shape a child's ability to adapt to external challenges. However, there is scarcity in the available research to support whether migrant children may respond to sensory stimuli compared to their peers habituating in their environment of origin.

Earlier findings have presented conflicting conclusions on the role of ethnicity in influencing the regulation of response to sensory stimuli, with rare explorations on. Methodological caveats related to the primary use of behavioural measures, uncontrolled experimental paradigm, and uncontrolled participant characteristics may likely confound former research. Employing mainly a neurophysiological perspective, this research hypothesizes that children from different ethnicities and geographic environments living in similar physical environments have significantly different adeptness in their regulation of response to sensory stimuli.

While evidence supports the claim that the environment may likely influence a child's regulation of response to sensory stimuli, it remains unknown whether certain features of the physical living environment (i.e. urban and rural environments) may further explain such differences. Applying neurophysiological perspectives, this research hypothesizes significant differences in the regulation of response to sensory stimuli among children from similar ethnicities and geographic environments living in different physical environments.

Chapter 3: Conceptual Framework

In this chapter, this research defines and discusses the concepts involved, their relationships and how it forms the research question. The concept of allostasis is used to demonstrate the influence of a child's ethnicity and environments on how they regulate responses to sensory stimuli. This section ends with the research questions, hypothesis, and aims of this thesis.

The conceptual framework of this research is depicted in Figure 3.1. This research primarily aims to identify the influence of ethnicity, environment or their interaction on the adaptation to regulate responses to external challenges (i.e. sensory stimuli) among children using a neurophysiologic perspective. In this framework, adaptation is depicted as a mechanism of responding to external challenges in the form of a sensory stimulus (e.g. auditory stimuli) arising from the external milieu. The mechanism that enables the response to external challenges (i.e. sensory stimuli) is supported by neurophysiological systems whose underlying regulation is under the control and dynamics of the sympathetic and parasympathetic branches of the autonomic nervous system. The consequent response may be categorised into behavioural and physiological. In this research, the focus is on the physiological response. The preceding sections further explain the conceptualisations and the relationships of the variables involved in this study.

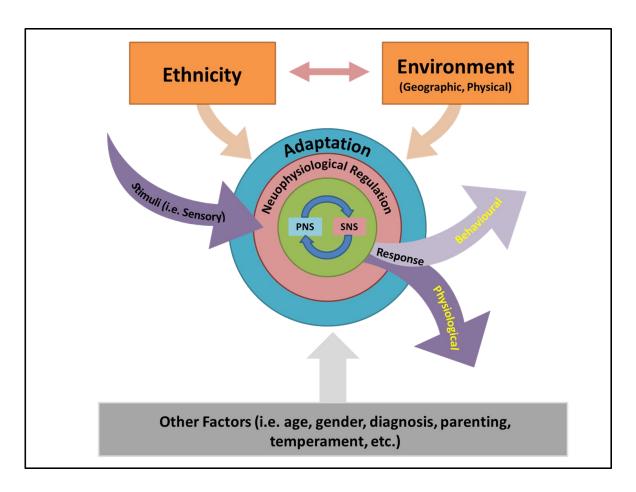


Figure 3.1. Conceptual Framework: Adaptation as an approach to examine the influence of ethnicity and the environment on the neurophysiologic regulation of response to sensory stimuli among children.

I. Allostasis as an Adaptive Mechanism

Allostasis allows individuals to adapt to the demands of the environment through physiological internal regulation (McEwen, 2008). In the face of environmental challenges, an individual's physiological systems respond by altering the organism's resources to create new set points allowing for adaptation (Ganzel & Morris, 2011; Juster et al., 2011). Allostasis is responsible for both in situ and long-term adaptive mechanisms that allow individuals to respond perturbation in their internal and external environments (Karlamangla et al., 2002; McEwen & Wingfield, 2003). Mediators of allostasis include, but are not limited to, the nervous, immunological and endocrine systems (McEwen, 1999, 2000; McEwen & Wingfield, 2003). Distinct neurophysiological circuits enable the regulation of response to external challenges and changes in the environment. These include the SNS and PNS branches of the autonomic nervous system (McEwen & Wingfield, 2003, 2010).

II. Adaptation is Influenced by Individual Differences

Adaptation to sensory stimuli (i.e. auditory stimuli) may be influenced by a number of factors (i.e. age, gender, diagnosis, parenting, temperament, etc.). Individual differences shape how individuals respond to external challenges (McEwen, 1999, 2008). A biological perspective is employed in this research to approach the definition of individual differences. In this research, such individual differences have been categorized to involve environment-dependent information appraisal and biological embedding in ethnicity (Danese & McEwen, 2012). Other factors that may influence adaptation to sensory stimuli are controlled at participant selection and/or statistical analysis to account for the variance it may contribute.

A. Environment-dependent Information Processing

Environment-dependent information processing suggests that one's life history, which may include environmental, cultural, social or economic backgrounds, has shaped their abilities to make sense of situations and instilled in us a set of codes that determines how we process or respond to stimuli (Mims & Olden, 2012; Templer, 2008; Wolf et al., 2007). In this research, the environment is further classified between geographical (i.e. country of origin, country of habitation) and physical (i.e. physical characteristics of urban and rural contexts).

B. Biological Embedding in Ethnicity

An individual's genetic trait as expressed in the body's physiology is represented by biological embedding (McEwen, 2008). Individual differences as a result of biological embedding influences the ability to adapt to environmental stimulation or challenges. In this research, ethnicity is viewed as a representation of biological embedding.

III. Adaptation Involves Regulation

Regulation entails some form of change or alterations in parameters to respond to an external challenge. Regulation to adapt to external challenges involves: 1) the readiness (capacity) to receive stimuli as represented by resting baseline (Berntson et al., 1991); and the reactivity to the stimuli measured in the magnitude of response (Andreassi 2013), measured by the response pattern across conditions noting changes in physiologic parameters and the levels of physiologic reactivity at each condition.

Regulation shapes neurophysiological functions to match the external environment (McEwen, 2000, 2003; McEwen & Wingfield, 2003; Sterling & Eyer, 1998).

IV. Adaptation in Facing Challenges

A. Response to Sensory Stimuli

Sensory information is the most basic form of challenge individuals face on a daily basis (Strickland, 2001). Sensory information from the environment is processed and regulated by the individual to support behaviours in everyday situations and development (Dunn, 1999; Goldstein et al., 2014). In this research, the response to sensory stimuli entails adjustments of internal measures as a reaction to a simple form of external challenge. This includes the child's physiological capacity to receive, react and consequently recover from the sensory stimuli. The resting condition represents the child's capacity to receive sensory information. Other authors have referred to the resting condition basal or baseline condition. In this research, the resting condition refers to the child's capacity to monitor and maintain internal homeostasis in response to environmental sensory challenges that reflect the dynamic relationship of PNS and SNS activity (Lai, 2013).

The stimulation condition refers to the reactivity of the child's physiological responses in the face of a block of passive auditory stimuli. The stimulation condition represents physiological reactions in response to sensory stimulation. Herein, magnitude changes in the parameters are observed as a result of stimulation from a resting condition (Cui et al., 2015).

The recovery condition refers to the point in time that stimulation is stopped or withdrawn, resulting in similarity to the resting condition. It is considered as a change from one condition (presentation of stimulus) to another (cessation of stimulus), and thus still represents an external challenge that needs adaptation. In this research, the recovery condition signifies the adaptive response to environmental challenges, such as in the case of sensory stimuli, that reflects perturbations in the internal states (Lai, 2013; Miller et al., 1999). The autonomic response of dynamic changes in lieu of the said internal state perturbations is aimed at meeting the challenges of the environment (Danese & McEwen, 2012; McEwen & Wingfield, 2010).

B. Auditory Stimuli as a Representative Sensory Stimuli

Different methods have been suggested to elicit physiological responses to sensory stimuli. The most common method in the literature is the use of a multi-sensory stimuli laboratory paradigm. The consequent delivery of different sensory stimuli may result in habituation (Andreassi, 2013). In this research, a single sensory stimulus was used to elicit physiological responses. Specifically, an auditory stimulus was used.

The auditory stimuli have previously been suggested in various research as a good representation of external challenge that children adapt to (Andreassi, 2013). Unlike other forms of sensory stimuli, an auditory stimulus taps into a subcortical route to the limbic system that elicits a rapid and stimulus-specific physiological autonomic (i.e. HRV, EDA) response (Andreassi, 2013; Garrido et al., 2012). Purposely, this thesis used a non-meaningful pure tone as an auditory stimulus in order to address the confounding effects of stimuli perceptual sensitivity (Andreassi, 2007; 2013).

The auditory system is one of the basic sensory systems. There are different sensations in the living environment, and an auditory stimulus can be considered as a representation of one of the sensory stimuli. In this thesis, an auditory stimulus has been applied as a sensory stimulus to elicit autonomic responses measured by activities of the PNS and SNS.

C. Neurophysiological Regulation of Response to Sensory Stimuli

As mentioned earlier, regulation involves a change of neurophysiological parameters in response to external challenges in order to adapt. The concept of allostasis explains how individuals adapt to challenges imposed by external environments (McEwen, 2000, 2003; McEwen & Wingfield, 2003; Sterling & Eyer, 1998). In allostasis, the body responds to external challenges mediated by various neurophysiological systems (i.e. PNS, SNS) to achieve an internal state of homeostasis (McEwen, 1998, 2003). In animals, it was found that responding to external challenges result to alteration of physiological parameters (i.e. increase in heart rate, respiration) in order to adapt (Berthold et al., 2013; Jachowski & Singh, 2015; Nathan et al., 2008). Among humans, regulation in the PNS (Matsushima et al., 2016; Schaaf et al., 2003; Schaaf et al., 2010) and SNS (Schoen et al., 2009;

Miller et al., 1999, McIntosh et al., 1999) occur in response to external challenges (i.e. sensory stimuli).

Manifestations of atypical sensory-related behaviours have been linked to underlying irregularities of autonomic regulation (Chang et al., 2012; Daluwatte, Miles, Sun & Yao, 2015; Matsushima et al., 2016; Schoen et al., 2009; Woodard et al., 2012). Measurement of mechanisms underlying the neurophysiological regulation of response to sensory information is recommended (Gomez et al., 2017, 2018). Changes in environmental situations should be indexed and include a resting, stimulation and recovery conditions (Lai, 2013; Miller et al., 1999).

Physiological activity may offer a sensitive and objective measure of the underlying adaptation mechanisms related to the regulation of response to external sensory challenges (Appelhans & Luecken, 2006). Previous research has found that sympathetic (SNS; McIntosh et al., 1999) and parasympathetic (PNS; Schaaf et al., 2003) autonomic functions are associated with the regulation of response to sensory stimuli. However, most of the previous the neurophysiological studies on the regulation of response to sensory stimuli has a limitation as they examined either SNS or PNS only (Gomez et al., 2017; Gomez et al., 2018; Schaaf et al, 2015). Examining from both systems (PNS and SNS) provides a clearer picture of the neurophysiological regulation of ANS responses.

V. Ethnicity Influences Adaptation

A. Biological-embedding of Ethnicity Influences Adaptation

Ethnicity is defined to be the biological origins and genetic similarities among individuals (Ali-Khan et al., 2011; Jorde & Wooding, 2004). Ethnicity is a representation of biologically-embedded genetic traits and consequent expressions, previously recommended to have an influence on the behavioural and physiological responses related to adaptation (Danese & McEwen, 2012). Thus, ethnicity may be a key variable that can profoundly modify the maturation of the neurophysiological systems responsible for adaptation to external challenges (Rutter & Krepner, 2007).

B. Ethnicity and Adaptive Responses to Sensory Stimuli

Behaviours are influenced by an individual's genetic makeup, and behaviours could likewise moderate transmission of the behavioural and genetic makeup of a child (Klahr & Burt, 2014). However, conflicting results have been suggested. Some researchers found that ethnicity effects can explain differences in the behavioural response to sensory stimuli, while others suggested otherwise (Caron et al., 2012; Royeen & Mu, 2000; Tirosh, et al., 2003). Whether the ability to regulate responses to sensory stimuli is influenced by a child's ethnicity remains inconclusive, or at best, inconsistent. As previously argued, the regulation of response to sensory stimuli entails an adaptive mechanism. The available evidence has used behavioural outcomes and testing in a non-controlled environment, which may likely have impacted the results. While behavioural measures may still be of some use to describe the regulation of response to sensory stimuli, this research adopts a more neurophysiological perspective. It is unknown whether differences in the response to sensory stimuli can be explained by a child's ethnicity or the environment in context. In this research, the researcher looks at how ethnicity may explain variations in a child's ability to adapt to sensory events in their external environment. This research uses a neurophysiological perspective in attempting to describe the regulation of response by indexing the interrelated operation of the PNS and SNS among a group of children with distinct (i.e. Chinese and Filipino) or similar ethnic origins, living in similar or and different geographic environments (i.e. Hong Kong and Philippines) or physical environments (i.e. urban and rural areas).

VI. Environment Influences Adaptation

A. Two Levels of Environment

In this research, the environment is classified on two levels: geographic and physical environment. Because the socio-cultural influences may likely confound the effects of the features within the geographic environment, the physical features of such an environment need to be further examined. The physical environment is referring to empirical aspects of the physical setting, which includes tangible natural or man-made structures and/or components (McDonell & Pickett, 1990). The physical environment is represented in this thesis by the contrasting

characteristics of urban and rural settings. Exposure to various experiences in different physical environment landscapes may influence how children adapt to external challenges in their environment.

Environment-dependent information appraisal suggests that the responses of an individual may likely have been shaped by similar socio-cultural traits and experiences shared by a category of people that set apart one group of people from another, embedded within similar geographic environments (Danese & McEwen, 2003; Robinson-Wood, 2016; Zimmerman & Woolf, 2014).

B. Environment and Adaptive Responses to Sensory Stimuli

The socio-cultural contexts and experiences within the geographic environments have been implicated to account for differences or associations between individuals from different or similar geographic environments (Caprio et al., 2008). Thus, adaptive behavioural response to sensory stimuli among children is likely influenced by their geographic environment habitat. The geographic environment where a child develops has the ability to override ethnic influences on adaptive sensory behaviours (Gunn et al., 2009). The various sensory experiences within the geographic environment can likewise shape how they respond to sensory stimuli. However, the experiences related to the sensory information may be different from one contextual geographic environment to another.

The urban context is described to be set in a metropolitan densely populated within a number of man-made human structures (i.e. houses, buildings, bridges, railways, etc.). On the other hand, a rural environment may be viewed as an open strip of natural environment (i.e. mountainous, coastal, agricultural) populated with fewer human inhabitants and man-made establishments. The physical environment thus refers to the physical contextual characteristics of the living environment. The physical environment where children dwell can greatly impact how individuals regulate responses to the external world (Laumann, Garling, & Stormark, 2003; Rutter et al., 2001; Ulrich et al., 1991). Children with similar ethnicities living in natural environments (i.e. rural context) have been shown to have different behavioural responses to sensory stimuli (Brown & Dunn, 2010; Lin et al., 2013). The sensory environment and the opportunities in the environment generally affect the brain and its functions (Kempermann, van Praag & Gage, 1999). However, up

to this point, whether the physical environment or other factors influence the variations in response toward sensory stimuli among individuals sharing similar ethnicities across environments as reflected by neurophysiological parameters is still uncertain.

The finding of previous research on the influence of environments and its levels is still inconclusive in explaining variations in the adaptive abilities of children in response to a sensory stimulus. The context upon experiencing the sensory stimulation may be varied between levels of the environment and may influence how children adapt to sensory stimuli. But the findings of previous studies have a limitation on explaining the influence of the environment. For example, the methods mainly used behavioural outcomes, which may reflect a lesser objective measure that can provide precise information about the regulation of response to sensory stimuli (Schoen et al., 2009). In this thesis, the researcher recruited children from similar ethnic origins (i.e. Filipino) from different geographic environments (i.e. Hong Kong and Philippines) and physical environments (i.e. urban and rural Philippines). Neurophysiological measures of HRV and EDA will be primarily employed to index the interrelated functions of the PNS and SNS, respectively. Behavioural measures of adaptive regulation of response to sensory stimuli in children will likewise be employed.

VII. Ethnicity x Environment Interaction and its Influence on Adaptation

It has been suggested that the interaction between ethnicity and environment has a moderating role in the activity of a child's autonomic activity that is further related to their ability to adapt to external stressors (Dieleman et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012). The interaction between genetics and the environment might have a stronger role in the regulation of response to sensory stimuli (Gomez, Lai & Tsang, under review). Such gene x environment interaction may be a stronger model to explain its role in neurophysiological mechanisms underlying the behaviours. Perturbations in the environment influence physiological regulation in migrating animal models (Jachowski & Singh, 2015; Harms et al., 2015; Nardocci et al., 2014). Genetic markers have likewise been shown to be associated with autonomic activity in children (Bortoluzzi et al., 2015; Pagliaccio et al., 2015a,b; Roberts et al., 2015).

As mentioned previously, there are two levels of environment. The first level is described to be the geographic environment. In this research, *ethnicity x environment* is represented by a group of children of migrant origins (Filipino children living in Hong Kong). This group was selected because their ethnicity (Filipino) was not formed in their current host country (Hong Kong) but in their country of origin (Philippines). This research will examine whether these children (Filipino children living in Hong Kong) has an adaptation to their current environment of habitat by comparing their regulation of response to sensory stimuli to their peers from their host country and country of origin (Filipino children living in the Philippines).

VIII. Knowledge Gap

A. Knowledge Gap 1

There is evidence by previous researchers that a child's ethnicity and their living environments can influence how children regulate responses to sensory stimuli. However, methodological variations and the limited inquiry on using more objective measures of the underlying mechanism that enables adaptation through regulation was suggested. Thus, it is unknown whether ethnicity shapes the capacity and reactivity of physiological response to sensory stimuli.

B. Knowledge Gap 2

Previous research suggested that the environment could override biologically-embedded traits, possibly related to one's ethnicity, and shape the child's physiological regulation. However, it was rarely examined whether this can be applied to inquiries associated with a child's ability to regulate responses to sensory stimuli. Thus, it is unknown whether among children of similar ethnicities but living in different the geographic environment shapes the capacity and reactivity of physiological response to sensory stimuli.

C. Knowledge Gap 3

Prior research evidence suggested the ability of the environment to shape a child's ability to adapt to external challenges. However, there is scarcity in the available research to support whether migrant children may respond to sensory stimuli compared to their peers habituating in their environment of origin. Thus, it is

unknown whether among children of different ethnicities but living in different the geographic environment shapes the capacity and reactivity of physiological response to sensory stimuli.

D. Knowledge Gap 4

Earlier findings have presented conflicting conclusions on the role of ethnicity in influencing the regulation of response to sensory stimuli, with rare explorations on. Methodological caveats related to the primary use of behavioural measures, uncontrolled experimental paradigm, and uncontrolled participant characteristics may likely confound former research. Employing mainly a neurophysiological perspective, this research hypothesizes that children from different ethnicities and geographic environments living in similar physical environments have significantly different adeptness in their regulation of response to sensory stimuli. Thus, it is unknown whether living in an urban or rural physical environment shapes the capacity and reactivity of physiological response to sensory stimuli.

IX. Research Question/Motivation

To achieve adaptation, regulation of responses towards external challenges in the environment is essential in daily activities. The research question in this thesis is: "do ethnicity and environments influence the regulation of response to sensory stimuli in children?" To answer this research question, this thesis identifies the role of ethnicity and environments on the regulation of response towards sensory stimuli among children from a neurophysiological perspective. The findings of this study may increase understanding of the possible effects of migration among children and informing policies and programs on the possible health support these children needs.

X. Research Aims

This research aims to identify the role of ethnicity and environments in the regulation of response to sensory stimuli in children, from a neurophysiological perspective.

Specifically, this research's objective is to: 1) compare the physiological (HRV and EDA) regulation of response towards sensory stimulus between children from different ethnicities living within the same geographic and physical environments; 2) compare the physiological (HRV and EDA) regulation of response towards sensory stimulus between children from similar ethnicities and physical environments living in different geographic environments;

3) compare the physiological (HRV and EDA) regulation of response towards sensory stimulus between children from different ethnicities and geographic environments living in similar physical environments; 4) compare the physiological (HRV and EDA) regulation of response towards sensory stimuli between children from similar ethnicities and geographic environments living in different physical environments.

XI. Research Hypothesis

This thesis has four main research hypotheses.

- A. *Hypothesis 1*: Chinese children living in Hong Kong and Filipino children living in Hong Kong will have significantly different regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions.
- B. *Hypothesis* 2: Filipino children living in Hong Kong and Filipino children living in urban Philippines will have significantly different regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions.
- C. *Hypothesis 3*: Chinese children living in Hong Kong and Filipino children living in urban Philippines will have significantly different regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions.
- D. *Hypothesis 4*: Filipino children living in urban Philippines and Filipino children living in rural Philippines will have significantly different regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions.

XII. Significance of this Study

The significance of this study lies in being able to further understand the neurophysiological mechanism and the influence of a child's ethnicity and their living environments on the ability to cope with external challenges, specifically with sensory stimuli. Specifically, the findings in this study have implications on research, practice and policy development relevant to childhood populations. Firstly, this research demonstrates a reconceptualised understanding of how neurophysiological mechanisms support the regulation of response to sensory stimuli. Understanding such mechanisms will help in better understanding of factors that may influence the development of children. Secondly, clinical practice will be informed towards the development of relevant assessment tools

that address physiological components; and in initiating innovative and novel interventions that addresses the underlying physiologic mechanisms supporting behaviours related to the dysregulation of response to sensory stimuli. Likewise, this thesis has implications on using auditory stimuli as an illustration of response toward sensory stimuli. Lastly, the findings of this research will support the development of policies that address the health and well-being needs of children who are transitioning from one environment to another.

Chapter 4: Methods

This section of the thesis describes the specific methodologies undertaken to answer the research question that examines the role of ethnicity and environments of the regulation of response to sensory stimuli among children. This section outlines the research design, description of participants, experimental protocol and paradigm, specific procedures, instrumentation measures used, and the analysis of data. The methods reported in this section of this thesis have already been published. The reader is advised to see Gomez et al., 2017 and Gomez et al., 2018.

I. Research Design

This thesis employed the use of a cross-sectional observational quantitative research design across different groups of participants at a specific time point (Mann, 2003). Participants were recruited and grouped based on specific characteristics that represents the aims of this study and were subjected to an experimental laboratory paradigm to determine their neurophysiological regulation of response towards sensory stimulation. Figure 4.1 provides a visual representation of the groupings of the participants based on the variables to be tested.

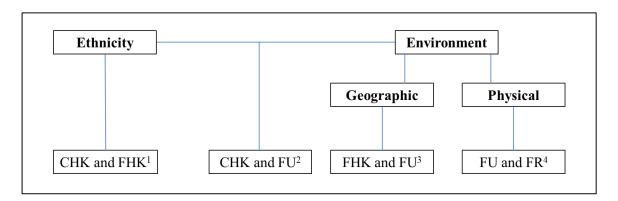


Figure 4.1. Summary of the groupings of the participants based on the variables in this thesis. (Note: ¹refers to the pairwise combinations for hypothesis testing 1; ²refers to the pairwise combinations for hypothesis testing 2; ³refers to the pairwise combinations for hypothesis testing 3; and ⁴refers to the pairwise combinations for hypothesis testing 4).

II. Participants

This research consists of four groups of participants divided into four subgroup combinations. They are (a) typically developing Chinese children living in Hong Kong (CHK group; n=31); (b) typically developing Filipino children in HK (FHK group; n=28); (c) typically developing Filipino children who are living at Urban area (FU group; n=54); and (d) typically developing Filipino children who are living at Philippines-Rural area (FR group; n=43).

The recruited children, boys and girls, were deemed as typically developing after screening for sensory modulation problems by using the Sensory Profile; and neurodevelopmental/ orthopaedic/ cardiorespiratory/ psychological/ neurobehavioural conditions by parent interview and questionnaire. For the CHK, FU and FR groups, they were born and raised in their current geographical and physical landscapes. Participants were screened for any maladaptive behaviour in sensory processing in daily activities using the Sensory Profile (Dunn, 1999) and parent-reported pertinent medical history. Participants who exhibited probable sensory processing issues (>3SD) were excluded from the study. Among the FU groups, migration history and immigrational generation were noted. Socio-economic data was gathered and matched based on the classifications by the Philippine Statistics Authority (2015) and the Hong Kong Census and Statistics Department (2016), which can be seen as an appendix at the end of this section. All children were enrolled in regular classrooms (public and primary schools) with no history of grade-level repetition and special education support. At the time of the testing procedures, all children were between the ages of 7-12 years. Table 4.1 presents the inclusion and exclusion criteria used for the participants in this thesis. Table 4.3 and 4.4 found at the end of this chapter outlines the classification for income and socio-economic class.

Table 4.1. Inclusion and Exclusion Criteria.

Inclusion Criteria	Exclusion Criteria		
Males and Females	Known history of developmental		
• 7-12 years old	disabilities, medical history of cardiac		
• For the CHK, FU and FR groups: were	or pulmonary problem, medical history		
born and raised in their current	of diabetes, or having any sensory		
geographical and physical landscapes	deficits		
• For the FU group: at least a period of	History of grade-level failure or		
three years (recent years) living in	repetition		
their current geographic environment	History special education support		
post-migration			

The sample size of the four groups: CHK, FHK, FU and FR are n=31, n=28, n=54 and n=43 respectively. This is deemed as ample based on the initial sample size calculations of a moderately large effect size of $f^2=0.625$, a power of 0.80 (Gomez et al., 2018). Table 4.2 presents the specific subgroups and the actual sample size gathered for this thesis.

Table 4.2. Specific Subgroups in this Thesis and the Gathered Sample Size.

Ethnicity	Chinese	Filipino		
Geographic Environment	Hong Kong	Hong Kong	Philippines	
Environment Landscape	Urban	Urban	Urban	Rural
Group 1	CHK (n=31)		FU (<i>n</i> = 54)	
Group 2	CHK (<i>n</i> = 31)	FHK (n= 28)		
Group 3		FHK (<i>n</i> = 28)	FU (<i>n</i> = 54)	
Group 4			FU (<i>n</i> = 54)	FR (<i>n</i> = 43)

The HK-Chinese (CHK) participants were recruited from mainstream primary schools in Hong Kong. Filipino children living in HK (FHK) were recruited from Filipino community group organizations as recommended by the Philippine consulate in HK. Filipino participants (FU and FR) were recruited from primary schools of urban (Manila, Marikina, Taguig, Quezon City) and rural (La Union) Philippines.

Recruitment was done through letters of invitation to the schools, community board posters, parent groups, and social media. Written informed consent was obtained prior to commencement of the experiment.

Ethical approval was obtained from the Hong Kong Polytechnic University, Human Subjects Ethics Sub-committee, with reference number HSEARS20150316001; and the University of Santo Tomas- College of Rehabilitation Sciences Ethics Review Committee with reference number FI-2015-021 and FI-2017-021. Participants were asked to complete the assent parental consent forms prior to laboratory testing. Testing was stopped when the participant expressed discomfort or displays an inability to tolerate the procedures.

III. Experimental Protocol

different sensory stimuli.

The experimental protocol used in this thesis was an adaptation of the original from the Sensory Challenge Protocol (SCP)² employed by Miller et al. (1999) and further modified by Lai (2013)³. In the studies of Miller et al. (1999) and Lai (2013), ANS response was measured across several sensory modalities.

In this thesis, to represent a sensory stimulus, an auditory stimulus was used to elicit ANS responses measured by PNS and SNS activity. Thus, only the part of the auditory stimuli of the Sensory Experiment (SE; Lai, 2013) was retained. Figure 4.2 represents the

² There are four phases in the manually-operated SCP: (1) a baseline period of three min where the child is resting and seated quietly in a chair; (2) the administration of a set of sensory challenges (consists of different sensory stimuli in seven domains, consecutively repeated for eight times each), (3) a recovery phase where the child returns to quiet sitting for a period of three minutes, and (4) an exposure to a two minute prolonged auditory tone (post-recovery condition). The different stimuli include: auditory tones (84 dB), visual (strobe lights of 20 watts at 10 Hz), auditory siren sound (78dB), olfactory (oil of wintergreen scent passed under the nose), tactile (moving touch along the jawline from one right to left using a feather), and vestibular (chair is suddenly tilted back to a 30° angle). After recovery, a two-minute prolonged auditory tone (75 dB) was administered. All stimuli, except for the prolonged auditory stimuli, occurred for three seconds with a pseudorandom interval of 12–17 second of interstimuli interval. By using the SCP, researchers can describe the PNS

activity levels at resting, during stimulation and recovery phases while determining associations between

³ The Sensory Experiment (SE) by Lai (2013) was a modified version of the SCP (Miller et al., 1999). There is a total of eight phases in the experiment that is operated by Labview: 1) initial resting period; 2) auditory processing; 3) first resting period; 4) visual processing; 5) second resting period; 6) tactile processing; 7) third resting period; and 8) anticipatory processing. The resting periods lasted for 2 minutes, while the initial resting period lasted for 5 minutes. There are three blocks of sensory tasks: auditory, visual and tactile. The auditory stimuli were presented as a burst of sound (i.e. smoke detector) at 84 dB with 10 trials, each lasting for 3 sec and an ISI of 15-19 s. The visual stimuli were carried out using bursts of light flashes at 10 Hz, 10 trials, lasting for 3 s with an ISI of 15-19 s. The tactile stimuli were a computer-operated vibration on the forearm at 142 hz, 10 trials each lasting for 3 s and an ISI of 15-19s. Another modification by the SE from Miller et al. (1999) was the use of an anticipatory task, a computer-generated active sensory processing task. By using the SE, autonomic activity was measured.

experimental paradigm of this research. Figure 4.3 describes the stimulus presentation in the experimental paradigm.

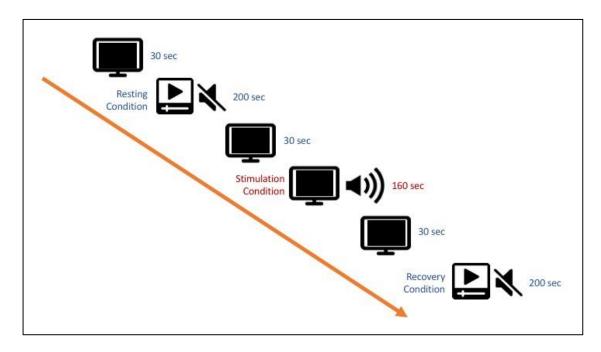


Figure 4.2. Experimental paradigm.

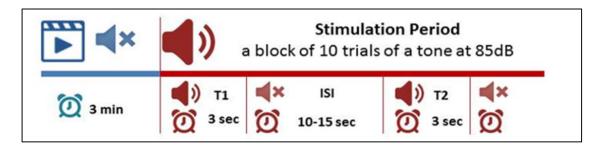


Figure 4.3. Experimental paradigm stimulus.⁴

The experimental paradigm used in this thesis lasted for an approximated 11 minutes, consisting of three discrete conditions: resting (200 s), stimulation (160 s), and recovery (200 s) conditions. This three-condition phase was similar to the SE (Lai, 2013) except for the use of a singular representative sensory stimulus (i.e. auditory stimuli). A three-minute resting period was employed to allow sufficient data time points necessary for the analysis of autonomic responses (Roth, Dawson & Filion, 2012; Task Force, 1996). At the resting condition, a 200-second clip of a silent cartoon movie was shown. This was similar to the SE by Lai (2013), however, the silent cartoon movie was changed to

⁴ T1 refers to the first-time point (i.e. first trial) where a 3-second auditory stimulus is presented. **ISI** refers to the pseudo-randomized inter-stimulus interval which lasts for 10-15 seconds. T2 refers to the second time point (second trial).

another cartoon movie clip. Allowing the child to watch a silent cartoon movie during resting condition was suggested by Andreassi (2013) for psychophysiological studies in paediatric population. During stimulation condition, the computer screen turns black and the participants are exposed to an auditory stimulus (presented as a block of 10 trials of a 4 kHz pure tone at 85dB; each trial lasting for 3 seconds, and a pseudorandomized inter-stimulus-interval in 10-15 seconds). The stimulation condition lasted approximately 160 seconds. A pure tone was used instead of suggestions from previous researchers (i.e. Miller et al.'s (1999) fire engine sound or Lai's (2013) smoke detector alarm), in an attempt of reducing stimuli-associated emotional responses. Specification for the loudness of the auditory stimuli (set at 85dB) allows for salient capturing of physiological responses and avoiding other internal physiological responses (Turpin, 1986). A pseudo-randomized inter-stimulus-interval in 10-15 seconds was selected in place to avoid participant adaptation to the stimuli and to allow just enough time to recover from the stimulus, without re-setting to baseline levels (Benedek & Kaernbach, 2010; Perakakis, Joffily, Taylor, Guerra & Jaime Vila, 2009). During the stimulation condition, a black blank screen was continuously displayed on the computer monitor screen. After the stimulation condition, a recovery condition ensues. During the recovery condition, the same silent cartoon movie (played during resting condition) was played for 200 seconds. A transition phase of 30 seconds in between condition was placed where written instructions were shown on the screen during the 11th-20th second⁵ within this transition period. Figure 4.4 shows screenshots of the instructions given to the participants. 6 LabVIEW (National Instrument, 2014) was used to develop and execute the experimental paradigm, which includes generation of the pure tone used as auditory stimuli. LabView program scripts provide objective, standardized and computerized administration of the auditory stimulus. Furthermore, the program can record real-time and elapsed time event markings necessary for data analysis in psychophysiological recordings.

⁵ The written instructions were shown in the middle of the 30 second transition phase in order to provide ample amount of time for the researcher to explain and reinforce the instructions, check for understanding and allow influence on the ANS response.

⁶ The instructions shown on the screen are as follow: 1) Before the resting condition: "You will be watching a silent cartoon movie shortly. Please sit still and look at the screen;" 2) Before the auditory condition: "You will be hearing some sounds shortly. Please sit still and look at the screen."

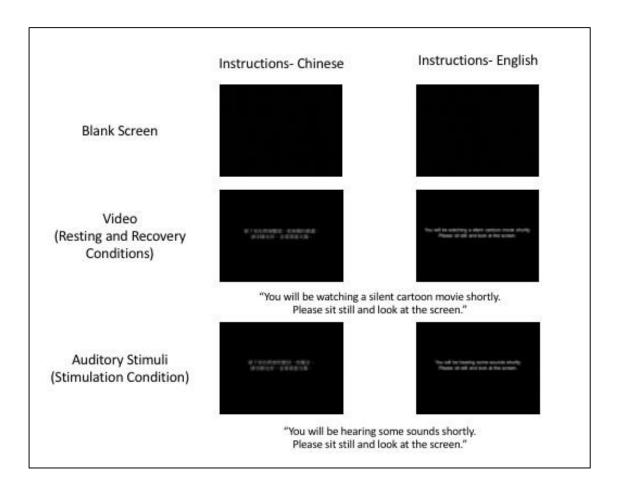


Fig. 4.4. Screenshot of the instructions on the computer monitor for the laboratory paradigm.

IV. Procedures

The procedures conducted in this study for the four groups, specifically the conduct of the experimental protocol was adapted from the study of Lai (2013). In that study (Lai, 2013), the protocol was developed to mitigate and lessen the possible influence of other factors on the ANS response of the participants. Thus, the same procedures were strictly implemented for the four groups at a controlled laboratory. The parents of the participants were instructed with several experiment preparatory reminders (Andreassi, 2013; Lai, 2013): no caffeinated drinks intake (up to 4 hrs before testing); no food intake (up to 1.5 hrs before testing); no rigorous physical exercise (before testing); no treatments/interventions (i.e.)such as sensory integration therapy, craniosacral therapy, acupuncture) or any other procedures that could influence ANS activity (up to 24 hours before testing); if the child is currently under medication, parents are to inform the tester to provide further information related to the medication. These measures were placed to control for possible confounding factors related to autonomic regulation.

Participants who are unable to concede and follow through with the given reminders were excluded from the final data analysis.

During the set day for experimental procedures, the participants were greeted by the researcher, led inside the testing room, initially accompanied by the parent where general instructions were given. Body mass index (BMI) was ascertained through measurement of the participants' height and weight and further transformed into BMI measurement. With the help of the parent or assistant, the participant wore the Polar H2 chest strap. Two finger cuffs were then fixed around the index and middle finger of the non-dominant hand of the participants. The testing set-up is depicted in Fig. 4.4. The researcher asked the child to sit on a comfortable child-sized chair, facing a 19-inch computer monitor, 24-36 inches (arm's length) away. A set of headphones were placed over the child's ear.

Autonomic physiological processes are easily affected by external environmental conditions. In this research, the testing environment has been controlled to reflect a non-overtly stimulating sensory environment. The testing room environment was dimmed to 10 lux (illumination level), set at 23-25 deg Celsius (temperature), controlled at 60-80% humidity and 40-45dB (noise level). At an arm's length of 60-80 cm away to the left of the participant was the researcher keeping minimal interaction and distraction. The procedures described in this thesis was based and adapted from that of Lai (2013) and mentioned by Gomez et al. (2017, 2018). These conditions are kept constant across all testing. Environmental conditions were recorded prior to and after the experimental procedures. These conditions were kept constant across all four groups. Figure 4.5 and 4.6 shows a typical set-up of the experiment.



Figure 4.5. Experimental set-up.



Figure 4.6. Sample set-up of the experiment with a participant.

During the actual experimental procedures, the researcher kept an observation log to record behavioural data from the child. Any movement, abnormal breathing patterns, sneeze, yawn, cough, external noise or technical problems with the neurophysiological equipment were recorded.

A 30-second transition phase from one condition to the next was included in the experimental paradigm for the purpose of taking a short rest period as needed. Gentle stretches, soft speaking or slow eye opening and closing were allowed (Brett-Green, Miller, Gavin & Davies, 2008). Similarly, during this time window, the participants were given gentle verbal reminders and reinforcements.

The parents were asked to complete the parent-answered questionnaires: Sensory Profile. The Chinese version was administered to the HK-Chinese participants, while the English version was administered to the Filipino participants. In cases that participants needed clarifications from the questionnaires, the researcher was present to answer their queries.

V. Instrumentation Measures

In this research, neurophysiological measures to index autonomic regulation were used to measure the regulation of response to sensory stimulus. Neurophysiological measures to index autonomic regulation used in this research were heart rate variability (which measures parasympathetic and sympathetic nervous systems activity levels) and electrodermal activity (which measure sympathetic nervous system activity levels).

A. Heart Rate Variability

This research uses heart rate variability (HRV) as measures of PNS and SNS functions across different conditions i.e. resting, stimulation, recovery). Specifically, this thesis used the Polar H2 Heart Rate Monitor (Polar, Finland) to measure HRV (Fig. 4.7). The equipment consists of a heart rate monitor that is attached to a chest belt and transmits heart rate signals via an infrared device to a computer notebook. Heart rate monitors such as the Polar H2 allows efficient data acquisition of heart rate variability and is suitable for the paediatric population. There is evidence that supports the use of Polar heart rate monitors as a valid instrument to measure heart rate variability (De Rooij, van Eijsden, Roseboom & Vrijkotte, 2013; Barbosa, Azevedo, Pastre & Vanderlie, 2016; Gomez et al., 2018; Plews, Scott, Altini, Wood, Kidling & Laursen, 2017; Williams, Jaczok, Ellis, Hillecke, Thayer & Koenig, 2016).



Figure 4.7. Polar H2 chest strap.

B. Electrodermal Activity

Electrodermal activity (EDA) is an autonomic measure that of the SNS. Issues on the sympathetic representativeness of the LF bands of HRV has led this research to use EDA as an adjunct SNS measure. The eSense Skin Response- GSR sensor (Mindfield, Germany; Fig. 4.8) measures SCL and SCR using a direct current (0.50 volts) at a resolution of 18-bit and 5 Hz/sec sampling rate. Exosomatic EDA measurement is achieved through two 5 mm Ag/AgCl electrode. While fairly new, the specifications of eSense meet the standard equipment guidelines set by Fowles et al. (1981) and downloaded data can be analysed following the guidelines for EDA publication (Roth, Dawson & Filion 2012). There are preliminary evidence supporting the reliability, validity and utility of using eSense in indexing electrodermal activity (Aymerich-Franch, Petit, Ganesh, & Kheddar, 2017; Chatterjee, Sinha, Sinha & Saha, 2016; Gomez et al., 2018; Hörmann, et al., 2016; Liapis et al., 2017).



Figure 4.8. eSense GSR sensor.

VI. Data Analysis

The data analysis that this research implemented are generally categorized into two: 1) neurophysiological data analysis; and 2) statistical analysis. It is imperative to properly analyse the neurophysiological data that indexes autonomic regulation (LF n.u., HF n.u. and EDA) using existing standards and guidelines prior to further statistical analysis. Each type of data analysis is discussed in the following subsections.

A. Neurophysiological Signal Processing

Two neurophysiological measures were used in this study. The methods of processing and analysis of 1) heart rate variability measures and 2) electrodermal activity measures are described below.

i. Heart Rate Variability Data Processing

In this study, a Polar H2 heart rate monitor was used to measure HRV. Online recording and raw data storage through infrared signals were done using Polar Trainer 5. No editing or correction was done to the original data stored. Windows PC-compatible computers were used for signal acquisition, storage, and processing. To analyse HRV data, aHRV (Nevrokard, Slovenia) was used. aHRV employs the current guidelines for HRV reporting (Task Force of The European Society of Cardiology, 1996).

Raw data from the Polar Trainer files were converted into HRV tachogram files. Prior to HRV analysis, tachograms were subjected to succinct preprocessing. Artefacts in HRV seem inevitable in psychophysiological studies (Berntson, Quigley, Jang, and Boysen, 1990). Arrhythmia occurrences, ectopic beats, missing data, movement artefacts, noise effects and skipped beats can inadvertently adjust the HRV data. Raw HRV data was transformed into tachograms, which were then subjected to visual analysis, and identification of artefacts, ectopic beats, and abnormal noise signals guided by the researcher's observation notes for each individual participant. Tachograms were epoched into specific time events corresponding to the experimental conditions resting, stimulation and recovery. Each subject was then left with a raw 200-second resting and

recovery period, and 160-second stimulation HRV time-epoched file. As recommended by the guidelines (Task Force 1996), comparison in HRV cannot be performed among varying time periods; hence the researcher analysed the signal with the length of 150-seconds for each condition. For the resting (original 200 s) and recovery (original 200 s) periods, the signals were extracted from the 20th-170th second of the corresponding time segment. For the stimulation condition, the 5th-155th second time segment was extracted for analysis. These time points were considered based to accommodate for considerable up and down-regulation effects. Thus, the epoched segments should represent better quality of recordings. Figure 4.8 shows a representation of the data epochs for HRV and EDA processed and analysed in this thesis.

The epoched HRV files were then subjected to correction of artefacts following the guidelines used by Task Force of The European Society of Cardiology (1996) as employed by the aHRV software. Artefact identification was done by comparing values to 20% under/over the mean of the preceding 25 beats (Task Force, 1996). Interpolation was performed to correct noise artefacts, thereby preserving optimum data sample integrity. A correction threshold of 3% from the composite HRV normalised data was set. Samples with >3% data correction were discarded from the final HRV analysis.

Analysis of the frequency domain component included: total power (TP; 0.0–0.5 Hz), low (LF; 0.04–0.15 Hz) and high (HF; 0.15–0.40 Hz) frequency components in their raw and normalized units (n.u.). The LF n.u. was used to represent sympathetic modulation activity (predominantly), while the HF n.u. represented parasympathetic modulation activity (Eckberg, 1997; Goldstein et al., 2011; Kleiger, Stein, & Bigger, 2005; Malliani et al., 1998; Malpas, 2002; Parati et al., 2006; Task Force, 1996; Taylor & Studinger, 2006). The meaning and calculations of parameters used are described in the earlier chapter of this report. The frequency-domain analysis was performed using the nonparametric method of the Fast Fourier transform (FFT). The direct current component was deleted, and a Hamming window was used to

reduce the effect of leakage (Kuo & Chan, 1993). For each data epochs, the analysis software was able to estimate the PSD based on FFT, which was then corrected for attenuation resulting. The power spectrum was subsequently quantified into various frequency-domain measurements, including total variance, high-frequency power, low-frequency power, and the LF/HF ratio. In particular, LF power was normalized by using the following formula: LF n.u.= LF/ (total power - VLF) x 100 to index sympathetic effects on HRV (Task Force, 1996). A similar procedure was also applied to HF where the following formula: HF n.u.= HF/ (total power - VLF) x 100 to assess parasympathetic effects (Task Force, 1996). To demonstrate and correct for possible skewness in the data, normalisation was performed.

ii. Electrodermal Activity Data Processing and Analysis

Electrodermal activity (EDA) was collected using eSense GSR (Mindfield, Germany). eSense GSR is composed of two Velcro-fastened electrodes that measure the skin conductance of the middle phalanx of the 2nd and 3rd digits of the non-dominant hand. Measurement was at 10 Hz, while five values per second (5Hz) are stored in the eSense GSR propriety application and can be exported via e-mail. Resolution is at 18 bits, rounded up to 2 decimal points. The real-time online recording used an iPad Mini and MS-Excel output file was downloaded for offline processing. Ledalab (www.ledalab.de), version v.3.2.9, a computer program set in a MATLAB environment, was used to process EDA data using the continuous decomposition method (CDA; Benedek & Kaernbach, 2010). Raw data from the eSense output (5Hz sampling frequency) were first converted into a spreadsheet file where discrete events based on the LabVIEW output were marked (i.e. start of resting condition, end of resting condition, start of stimulation block, the 10 events within the stimulation condition, etc) and then converted into a .txt file before being exported into Ledalab. EDA data were epoched into three distinct events: resting, stimulation and recovery conditions. Figure 4.8 shows a representation of the data epochs for HRV and EDA processed and analysed in this thesis.

For all three epoched conditions, a 2-sec pre-event and 9-sec post-event period were taken into consideration to compensate for the delay in the researcher's reaction time (Wilkes et al., 2009). The epoched EDA data were then pre-processed individually. The EDA data were then groomed to reduce noise, which includes manual smoothing using a 5-sec Hann window (Christou et al., 2011; Lajante et al., 2012) and a unidirectional first-order Butterworth low pass filter at 5 Hz frequency cut-off (Bach, 2014; Staib et al., 2015). After identifying EDA artefacts, a spline interpolation within a 5-sec pre/post parameter was done to correct the data (Dibbets et al., 2011). The groomed signals were then analysed using the CDA method (Benedek & Kaernbach, 2010) by optimally fitting the data to a bi-exponential. For the resting and recovery condition, the skin conductance level (SCL) was extracted by using the CDA.Tonic parameter.⁷

The software can identify significant peaks of >0.05 μ S (Boucsein, 2012), within a response window of 1-4 seconds post-stimulus (Bach, 2014; Lapate, 2014) for the stimulation condition which consisted of 10 trials. SCR components across trials were averaged (zero responses not included) within the stimulation condition block to represent an estimate of the mean SNS activity that reflects the responsiveness to sensory stimulus. A z-score transformation of individual SCR data was performed to remove within and between-subject variance (Bach, 2014). Normalization of the data was achieved through square root transformation of the SCR to correct skewness and meet parametric statistical assumptions (Kohrs, 2014; Krabs et al., 2015; Lajante et al.,

7

⁷ The CDA.Tonic parameter computes for the mean tonic EDA within the epoched response window (180-second block), in an aggregated 10-second within-window averaging method. The CDA.Tonic parameter represents the" most accurate representation of the deconvoluted underlying skin conductance level within the specified block expressed in μS" (Benedek & Kaernbach, 2010). This parameter has been logarithmically transformed to correct for skewed distribution using the following formula: √(CDA.Tonic). For the processing of SCR of the resting, stimulation and recovery condition, a similar CDA method was used (Bateman functions comprising onset, amplitude, T1 and T2 parameters. The T1 and T2 functions represent the value Tau in their computational analysis to perform the CDA).

 $^{^8}$ To represent the sympathetic activity in the form of EDA, the SCR was identified using the CDA.SCR parameter (Bach, 2014; Chen et al., 2013; Lapate, 2014; Seidel et al., 2013). The CDA.SCR is considered as the "average phasic driver within the specified response window initially set, and represents the phasic EDA most accurately expressed in units of μS " (Benedek & Kaernbach, 2010). Along with the data provided by the Ledalab output under the CDA.nSCR (identifies the number of significant, above the set >0.05 μS threshold, skin conductance responses) parameter, significant amplitude responses identified.

2012; Lapate, 2014). A logarithmic transformation using the following formula was employed: √(CDA.SCR+1). However, in order to compare the SCR across conditions, this research also utilized the same CDA.SCR parameters as a block whenever necessary. Similar normalization was performed for the block design parameters of its tonic counterpart. Figure 4.9 shows the representation of time event epochs processed and analysed in the experimental laboratory paradigm.

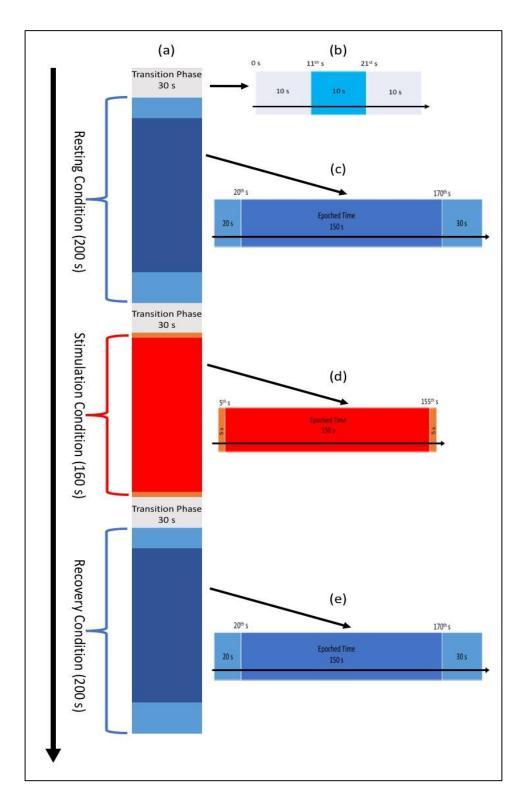


Figure 4.9. Representation of time event epochs processed and analysed in the experimental laboratory paradigm. (a) represents the timeline of the entire laboratory paradigm divided into events. (b) represents the breakdown of the transition phase. (c) to (d) represents the specific time epochs used for data processing for the resting, stimulation and recovery conditions, respectively.

B. Statistical Analysis

A series of similar statistical tests were employed to test hypotheses 1-4, where independent variables differed between hypotheses testing to reflect their specific objectives. To test hypotheses 1, that children from different ethnicities living within the same geographic and physical environments will have significant differences in their regulation of response to sensory stimuli, group differences were examined between CHK and FHK participants. To test hypothesis 2, that there are significant differences in the regulation of response to sensory stimuli among children from similar ethnicities and physical environments living in different geographic environments, group differences were examined using data from FHK and FU participants. To test hypothesis 3, that children from different ethnicities and geographic environments living in similar physical environments have significantly different adeptness in their regulation of response to sensory stimuli, CHK and FU groups were compared. To test hypothesis 4, that there are significant differences in the regulation of response to sensory stimuli among children from similar ethnicities and geographic environments living in different physical environments, data from FU and FR groups were examined.

A mixed factorial ANOVA and MANOVA tests were done among different pairwise combinations of the participants (Hypothesis 2: CHK and FHK; Hypothesis 3: FHK and FU; Hypothesis 4: CHK and FU; Hypothesis 5: FU and FR) across Hypotheses testing 1 to 4 on the neurophysiological measures of regulation to sensory stimuli (LF n.u., HF n.u. and SCL/SCR) across laboratory conditions (resting, stimulation, recovery). Univariate and multivariate tests were performed to determine group and subgroup baseline similarities at α = 0.05. Variables deemed significantly different were used as covariate/s in general linear modelling and are included in the report. A paired sample t-test was done between the groups among the behavioural measures at a critical value of α = 0.05. The effect size was interpreted using Cohen's d.

MANOVA tests, specifically the Pillai's V test (1), were conducted to determine whether effects of ethnicity or environment (used as the Independent Variables (IV) in this study) can explain differences in the multivariate set of dependent variables (LF n.u., HF n.u. and SCL/SCR), across the different studies, with different

participant matches (Hypothesis 1: CHK and FHK; Hypothesis 2: FHK and FU; Hypothesis 3: CHK and FU; Hypothesis 4: FU and FR); during three separate experimental conditions (resting, stimulation and recovery). Likewise, follow-up individual ANOVAs were also conducted. There are several MANOVA tests, and Pillai's V (Λ) test is one of them, considered to be the most powerful and robust statistic for general use and when there are departures from traditional assumptions (Pillai, 2004). A traditional critical value of α =0.05 was set across hypotheses 2 to 5. However, a Bonferroni adjusted p-value of 0.017 (0.05/3) was likewise set for the multivariate tests. The effect size was interpreted using Cohen's d.

Table 4.3 Income Classification*

Income Class	Description	Hong Kong	Philippines
Poor	Average income <	< HKD 4,000	< PHP 7,890 per
	official poverty	per month	month
	threshold		
Low Income	Average income	HKD 4,000 to	PHP 7,890 to
	between poverty line -	HKD 9,999 per	PHP 15,780 per
	2 x poverty line	month	month
Lower Middle Income	Average income	HKD 10,000 to	PHP 15,780 to
	between 2 x poverty	HKD 24,999 per	PHP 31,560 per
	line - 4 x poverty line	month	month
Middle Class	Average income	HKD 25,000 to	PHP 31,560 to
	between 4 x poverty	HKD 39,999 per	PHP 78,900 per
	line - 10 x poverty line	month	month
Upper Middle Income	Average income	HKD 40,000 to	PHP78,900 to
	between 10 x poverty	HKD 59,999 per	PHP 118,350
	line – 15 x poverty line	month	per month
Upper Income	Average income	HKD 60,000 to	PHP 118,350 to
	between 15 x poverty	HKD 99,000 per	PHP 157,800
	line – 20 x poverty line	month	
Rich	Average income at	HKD 100, 000	At least PHP
	least equal to 20 x	and over	157,800
	poverty line		

^{*}Based on the classifications by the Philippine Statistics Authority (2015) and the Hong Kong Census and Statistics Department (2016).

Table 4.4. Socio-economic Classification*

Class	% Share in Total Income	Income Class
AB	9%	Between Rich and Upper Income
С	26%	Between Upper Middle Income and Middle Class
D	56%	Between Lower Middle

		Income and Low Income
Е	9%	Poor

^{*}Based on the classifications by the Philippine Statistics Authority (2015) and the Hong Kong Census and Statistics Department (2016).

Chapter 5: Results

In this study, the role of ethnicity and environment in the regulation of response to sensory stimuli in children was examined from a neurophysiological perspective. The research question of this thesis is: "do ethnicity and environments have an influence on the regulation of response to sensory stimuli in children. In this thesis, to represent a sensory stimulus, an auditory stimulus was used to elicit ANS responses measured by PNS and SNS activity. This section is divided into four sections, each one describing the results of each of the specific hypothesis testing in this thesis.

I. Research Hypothesis 1

A. Overview

Hypothesis 1 was tested. The influence of ethnicity on the adaptation to sensory stimuli using an auditory stimulus was examined. This thesis hypothesizes that Chinese children living in Hong Kong and Filipino children living in Hong Kong will have significantly different neurophysiological regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions. A baseline-corrected 2x3 mixed factorial ANOVA with follow-up Bonferroni correction (α = 0.05) was conducted with follow-up MANOVA test differences between CHK and FHK group's autonomic activity (LF n.u., HF n.u., SCR/SCL) at each condition (resting, stimulation and recovery).

B. Summary of Participant Characteristics

Table 5.1.1 presents the summary of participant characteristics between typically developing Chinese (CHK; n= 31) and Filipino (FHK; n=28) children living in Hong Kong (n=59). 64.30% among the FHK group were native-born compared to 100% native-born CHK children. For both groups, the majority were males (65.63% in CHK and 85.71% in FHK). On average, the FHK group is slightly older M= 9.57 yr. (SD= 1.91) compared to the CHK children at M= 8.59 yr. (SD= 1.58). BMI was slightly different between groups where the CHK children lower at M= 17.09 (SD= 2.72) compared to their FHK counterparts who registered at M= 18.83 (SD= 3.54). All of the FHK children attend public schools, while a majority of the CHK participants (63.33%) attends private school. In both groups, both parents were working while the mother was the identified primary caregiver who has at

least a college level of education. Among the CHK group, 38.70% were classified to be upper middle income and upper income, while 71.40% were upper middle income in the FHK group. Majority of the families for both groups belong to socioeconomic class AB.

Table 5.1.1. Summary of Participant Characteristics Between CHK and FHK Groups (n= 59).

	СНК	FHK
Characteristics	n=31	n=28
Age in yrs. $M(SD)$	8.59 (1.58)	9.57 (1.91)
BMI in $Kg/M^2 M(SD)$	17.09 (2.72)	18.83 (3.54)
Migration Status ^a	100% Native Born	64.30% Native Born
Gender ^a	65.63% Male	85.71% Male
School Type	63.33% Private School	100% Public School
No. of Parents Working	100% 2 Parents	100 % 2 Parents
Primary Caregiver	100% Mother	100% Mother
Educational Level of Primary Caregiver	100% College Level	100% College Level
Income	38.70% Upper Middle	71.40% Upper
Classification ^b	Income	Middle Income
Socio-Economic Classification ^{b,c}	93.5% Class AB	96.4% Class AB

Note: anot similar at baseline p>0.05, thus used as a covariate in subsequent MANOVA tests; for the classification of income and socio-economic status, see Appendix 4.A; socio-economic Status ranges from Class A-E, with Class A representing richer families and Class E representing poorer families

C. Differences in the Patterns of Neurophysiological Regulation Between CHK and FHK Groups

To answer the question on whether ethnicity influences the regulation of response to sensory stimuli, a baseline-corrected 2x3 mixed factorial ANCOVA (using migration status and age as covariates) with follow-up Bonferroni correction (α =0.05) was conducted to determine the effect of ethnicity and environments on neurophysiological measures of regulation of response to sensory stimuli (LF n.u., HF n.u., SCR, SCL) across experimental conditions (resting, stimulation, recovery). Differences in the change of neurophysiological parameters related to the regulation of response to an auditory stimulus between CHK and FHK groups were examined. This group represents children from different ethnicities living in similar

geographic environments (i.e. Hong Kong). Table 5.1.2 summarises the main effects of pairwise (CHK and FHK) baseline-corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter. Table 5.1.2 summarises the condition pairwise (CHK and FHK) baseline-corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter.

i. LF n.u.

Change in the mean LF n.u. significantly varied across conditions (F(2,110)= 74.97, MS= 499.31, p= 0.01). When factored using age and migration status as a covariate, the model still yields significant differences of p = 0.05. There was no significant interaction between condition and participant grouping for the LF n.u. measure (F(1, 110) = 0.75, MS = 75.51, p = 0.47). The results thus only suggest within-group differences in the change of the LF n.u. across conditions. Condition pairwise comparisons did suggest significant differences in the direction of change of LF n.u. values from resting to recovery conditions (MD = -5.62, p = 0.01). The LF n.u. did not significantly increase during stimulation condition but continued to increase during the recovery condition. At recovery conditions, LF n.u. levels were significantly higher compared to resting values. Figure 5.1.1 displays the covariate (age and migration status) adjusted estimated marginal means of the LF n.u. for this 2x3 mixed factorial ANCOVA between CHK and FHK groups. The regulation of response to sensory stimuli as represented by the pattern of change in the LF n.u. did not have significant interaction between condition and participant grouping among children having different ethnicities but living in similar geographic environments (CHK and FHK).

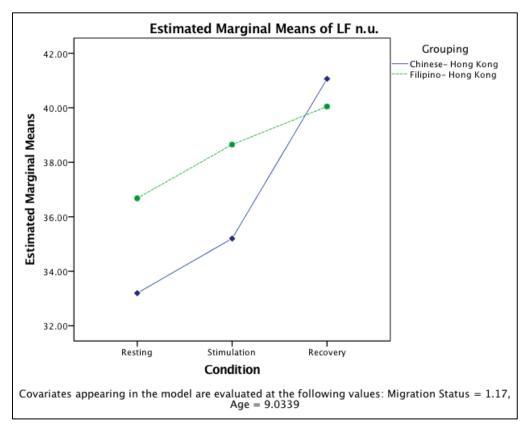


Figure 5.1.1. Covariate (age and migration status) adjusted estimated marginal means for LF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FHK).

ii. HF n.u.

Changes in the mean HF n.u. significantly varied across conditions (F(2, 110) = 4.19, MS = 453.35, p = 0.02) within CHK and FHK groups. There was no significant interaction between condition and participant grouping for the HF n.u. measure (F(1, 110) = 0.42, MS = 45.36, p = 0.67). The results thus only suggest within-group differences on the mean change of the HF n.u. across conditions. Condition pairwise comparisons did suggest significant differences in the direction of change of HF n.u. values only for resting to recovery conditions (MD = 5.37, p = 0.02). At stimulation condition, HF n.u. levels were not significantly different during resting conditions but continued to show a significant pattern of decrease measured during the recovery condition. At recovery conditions, the HF n.u. was significantly lower when compared to the resting values. Figure 5.1.2 displays the covariate (age and migration status) adjusted estimated marginal means of the HF n.u. for this 2x3 mixed factorial ANCOVA between CHK and FHK

groups. The regulation of response to sensory stimuli as represented by the pattern of change in the HF n.u. did not have significant interaction between condition and participant grouping among children having different ethnicities but living in similar geographic environments (CHK and FHK).

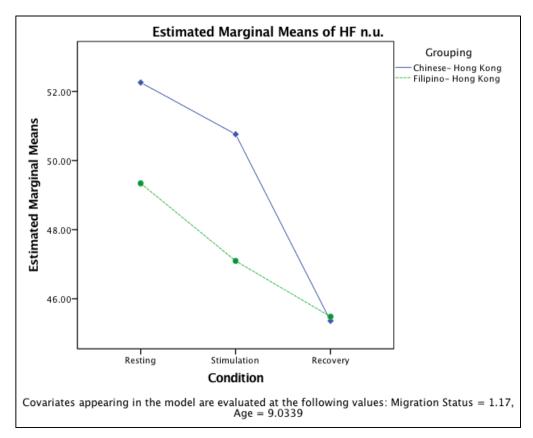


Figure 5.1.2. Covariate (age and migration status) adjusted estimated marginal means for HF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FHK).

iii. SCR

A baseline-corrected 2x3 mixed factorial ANCOVA (using migration status and BMI as covariates) was carried out for the neurophysiological variable, SCR, and as Mauchly's test was significant (p<0.05), the Greenhouse-Geisser correction was applied. Changes in the mean SCR values significantly varied across conditions (F(1.11, 110)= 16.89, MS= 0.33, p<0.00). The results suggest that the mean SCR values within CHK and FHK groups significantly varied across conditions. However, within groups interaction between condition and grouping for the SCR measure did not reach significance thresholds (F(1.11, 110)= 3.15, MS= 0.06, p= 0.08).

Nevertheless, condition pairwise comparisons did suggest significant differences in the direction of change of SCR values for resting to stimulation condition (MS= -0.11, p<0.00) and resting to recovery condition (MD= -0.06, p<0.00). Thus, SCR levels were significantly increased from resting condition upon auditory stimuli presentation but were not significantly different to SCR levels at recovery conditions. At recovery conditions, the SCR levels were significantly lower when compared to resting values. Figure 5.1.3 displays the covariate (age and migration status) adjusted estimated marginal means of the SCR for this 2x3 mixed factorial ANCOVA between CHK and FHK groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCR showed a significant interaction between condition and participant grouping among children having different ethnicities but living in similar geographic environments (CHK and FHK).

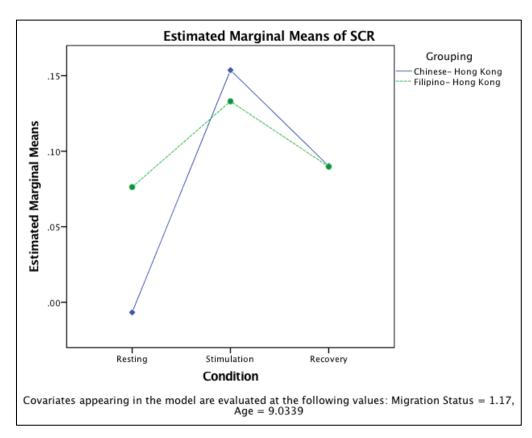


Figure 5.1.3. Covariate (Age and migration status) adjusted estimated marginal means for SCR using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FHK).

iv. SCL

The mean change in SCL values of CHK and FHK groups significantly varied across conditions (F(2, 110) = 11.75, MS = 0.13, p < 0.00). There was significant interaction between condition and participant grouping for the SCL measure (F(2, 110) = 10.61, MS = 0.12, p < 0.00). The results suggest that the mean change in SCL values within both participant groups (CHK and FHK) significantly varied across conditions. Follow-up tests on the condition pairwise comparisons did suggest significant differences in SCL change from stimulation to recovery (MD = -0.09, p < 0.00) and resting to recovery conditions (MD= -0.06, p= 0.01). The results suggest significant differences in the patterns for regulation between CHK and FHK groups. For the CHK group, SCL values significantly decreased during auditory stimulation condition, while the FHK group showed a significant increase in SCL values into the stimulation condition. Both groups showed significant SCL level differences from resting and recovery conditions. At recovery conditions, the SCL level was significantly higher than in the resting condition for both groups. Figure 5.1.4 displays the covariate (age and migration status) adjusted estimated marginal means of the SCL for this 2x3 mixed factorial ANCOVA between CHK and FHK groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCL did not have significant interaction between condition and participant grouping among children having different ethnicities but living in similar geographic environments (CHK and FHK).

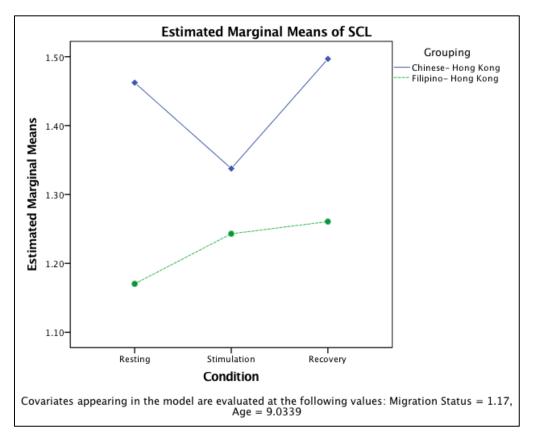


Figure 5.1.4. Covariate (age and migration status) adjusted estimated marginal means for SCL using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FHK).

Table 5.1.2. Summary of Differences in the Patterns of Neurophysiological Regulation Between CHK and FHK groups using Main Effects of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

		Uncorrected Main Effects						Baseline Corrected Main Effects								
Outcomes		SS	df	MS	F	Sig	ES	SS	df	MS	F	Sig	ES			
LF n.u.	Within															
	Condition	802.01	2	401.00	3.99	0.02	0.07	998.61	2	499.31	4.97	0.01	0.08			
	Condition * Migration	678.95	2	339.48	3.38	0.04	0.06	617.67	2	308.84	3.07	0.05	0.05			
	Condition * Age	301.68	2	150.84	1.50	0.23	0.03	314.49	2	157.24	1.56	0.21	0.03			
	Condition * Grouping	124.55	2	62.28	0.62	0.54	0.01	11056.20	110	100.51						
	Error	11056.20	110	100.51												
	Between															
	Migration	655.96	1	655.96	1.00	0.32	0.02	491.03	1	491.03	0.75	0.39	0.01			
	Age	28.47	1	28.47	0.04	0.84	0.00	35.18	1	35.18	0.05	0.82	0.00			
	Grouping	26.05	1	26.05	0.04	0.84	0.00	131.48	1	131.48	0.20	0.66	0.00			
	Error	36189.46	55	657.99				36189.46	55	657.99						
HF n.u.	Within															
	Condition	1276.21	2	638.10	5.90	<0.00	0.10	906.70	2	453.35	4.19	0.02	0.07			
	Condition * Migration	454.87	2	227.43	2.10	0.13	0.04	325.17	2	162.59	1.50	0.23	0.03			
	Condition * Age	68.83	2	34.42	0.32	0.73	0.01	70.67	2	35.33	0.33	0.72	0.01			
	Condition * Grouping	11.96	2	5.98	0.06	0.95	0.00	90.71	2	45.36	0.42	0.66	0.01			
	Error	11900.12	110	108.18				11900.12	110	108.18						
	Between															
	Migration	1108.15	1	1108.15	1.36	0.25	0.02	629.98	1	629.98	0.77	0.38	0.01			
	Age	107.53	1	107.53	0.13	0.72	0.00	138.78	1	138.78	0.17	0.68	0.00			
	Grouping	6.57	1	6.57	0.01	0.93	0.00	156.99	1	156.99	0.19	0.66	0.00			
	Error	44769.04	55	813.98				44769.04	55	813.98						

SCR Within

	Condition	0.21	2	0.11	9.77	<0.00	0.15	0.36	1.11	0.33	16.89	<0.00	0.23
	Condition * Migration	0.00	2	0.00	0.23	0.80	0.00	0.03	1.11	0.03	1.45	0.24	0.03
	Condition * Age	0.05	2	0.02	2.09	0.13	0.04	0.04	1.11	0.04	2.07	0.15	0.04
	Condition * Grouping	0.01	2	0.01	0.61	0.54	0.01	0.07	1.11	0.06	3.15	0.08	0.05
	Error	1.19	110	0.01				1.19	110	0.01			
	Between												
	Migration	0.00	1	0.00	0.13	0.71	0.00	0.07	1	0.07	2.34	0.13	0.04
	Age	0.00	1	0.00	0.07	0.79	0.00	0.00	1	0.00	0.00	0.95	0.00
	Grouping	0.06	1	0.06	2.08	0.15	0.04	0.01	1	0.01	0.49	0.49	0.01
	Error	1.63	55	0.03				1.63	55	0.03			
SCL	Within												
	Condition	0.19	2	0.10	8.92	<0.00	0.14	0.26	2	0.13	11.75	<0.00	0.18
	Condition * Migration	0.13	2	0.06	5.87	<0.00	0.10	0.22	2	0.11	10.20	<0.00	0.16
	Condition * Age	0.06	2	0.03	2.64	0.08	0.05	0.04	2	0.02	2.01	0.14	0.04
	Condition * Grouping	0.01	2	0.01	0.56	0.57	0.01	0.23	2	0.12	10.61	<0.00	0.16
	Error	1.19	110	0.01				1.21	110	0.01			
	Between												
	Migration	13.96	1	13.96	10.27	<0.00	0.16	11.65	1	11.65	8.57	<0.00	0.13
	Age	3.72	1	3.72	2.74	0.10	0.05	3.95	1	3.95	2.91	0.09	0.05
	Grouping	0.38	1	0.38	0.28	0.60	0.01	1.46	1	1.46	1.07	0.30	0.02
-	Error	74.72	55	1.36				74.81	55	1.36			

Note: LF n.u.= HRV- Low Frequency normalized unit; HF n.u.= HRV- High Frequency normalized unit; SCR- EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; SS= Sum of Squares; df= Degrees of Freedom; MS= Mean Squared; ES= effect size (Cohen's d); Sig= significant difference at p < 0.05; asignificant (p < 0.05) Mauchly's test

Table 5.1.3. Summary of Differences in the Patterns of Neurophysiological Regulation Between CHK and FHK Group Using Condition Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

	Baseline Corrected Condition Pairwise Comparisons															
		Rest-S	Stim				St	im-Rec	ov			Rest-Recov				
				95%	6 CI				95%	6 CI				95%	6 CI	
Outcomes	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	
LF n.u.	-1.99	1.71	0.75	-6.20	2.23	-3.63	1.94	0.20	-8.41	1.15	-5.62*	1.90	0.01	-10.30	-0.93	
HF n.u.	1.87	1.81	0.92	-2.61	6.35	3.50	2.05	0.28	-1.57	8.57	5.37*	1.88	0.02	0.73	10.02	
SCR	-0.11*	0.02	<0.00	-0.16	-0.05	0.05	0.02	0.08	0.00	0.11	-0.06*	0.01	<0.00	-0.07	-0.04	
SCL	0.03	0.02	0.61	-0.02	0.08	-0.09*	0.02	< 0.00	-0.13	-0.05	-0.06*	0.02	0.01	-0.11	-0.01	

Note: LF n.u.= HRV- Low frequency normalized unit; HF n.u.= HRV- High frequency normalized unit; SCR: EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; MD= Mean Difference; SE= Standard Error; LB= Lower Bound limit; UB: Upper Bound limit; Sig= significant difference at p < 0.05; * significant at p < 0.05; * significant (p < 0.05) Mauchly's test

D. Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between CHK and FHK Groups

i. Resting Condition

For the resting condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (CHK: n= 31, age= 8.59 yr., BMI= 17.09 kg/m²; FHK: n= 28, age= 9.57 yr., BMI= 18.83 kg/m²) by MANOVA, and migration status and age as covariates (see Table 5.1.1).

There is no significant multivariate group mean difference between the two groups (Λ = 0.01, F(3,11)= 0.20, p= 0.89, d= 0.21) on LF n.u. (CHK: M= 36.94; SD= 15.71; FHK: M= 34.85; SD= 15.29), HF n.u. (CHK: M= 50.87; SD= 19.13; FHK: M= 53.81; SD= 16.60), and SCL (CHK: M= 1.32; SD= 0.48; FHK: M= 1.49; SD= 0.94). Table 5.1.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is no significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at resting condition among children having different ethnicities but living in similar geographic environments (CHK and FHK).

ii. Stimulation Condition

For the stimulation condition, LF n.u., HF n.u. and SCR were compared between the two groups of participants (CHK: n= 31, age= 8.59 yr., BMI= 17.09 kg/m²; FHK: n= 28, age= 9.57 yr., BMI= 18.83 kg/m²) by MANOVA, and migration status and age as covariates (see Table 5.1.1).

There is no significant multivariate group mean difference between the two groups (Λ = 0.03, F(3,11)= 0.52, p= 0.67, d= 0.34) on LF n.u. (CHK: M= 37.36; SD= 14.57; FHK: M= 36.84; SD= 20.09), HF n.u. (CHK: M= 49.02; SD= 18.15; FHK: M= 52.70; SD= 20.59), and SCR (CHK: M= 0.48; SD= 0.59; FHK: M= 0.65; SD= 0.67). Table 5.1.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is no significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCR at stimulation condition among children having different ethnicities but living in similar geographic environments (CHK and FHK).

iii. Recovery Condition

For the recovery condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (CHK: n= 31, age= 8.59 yr., BMI= 17.09 kg/m²; FHK: n= 28, age= 9.57 yr., BMI= 18.83 kg/m²) by MANOVA, and migration status and age as covariates (see Table 5.1.1).

There is no multivariate significant group mean difference between the two groups (A= 0.00, F(3,11)= 0.05, p= 0.99, d= 0.10) on LF n.u. (CHK: M= 41.25; SD= 15.99; FHK: M= 40.58; SD= 19.56), HF n.u. (CHK: M= 45.42; SD= 17.75; FHK: M= 46.58; SD= 18.12), and SCL (CHK: M= 1.39; SD= 0.52; FHK: M= 1.52; SD= 0.86). Table 5.1.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is no significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at recovery condition among children having different ethnicities but living in similar geographic environments (CHK and FHK).

Table 5.1.4. Summary Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between CHK and FHK Groups Using MANOVA Tests (n= 59).

	CI	CHK FHK Total			M		ANOVA						
		31	n=	28	n=	59	Pillai's V	F		J	F		
	M	SD	M	SD	M	SD	Filial S V	Г	p	d		p	d
Resting- LF n.u.	36.94	15.71	34.85	15.29	35.95	15.41					0.10	0.76	0.08
Resting- HF n.u	50.87	19.13	53.81	16.6	52.27	17.89	0.01	0.20	0.89	0.21	0.00	1.00	0.00
Resting- SCL	1.32	0.48	1.49	0.94	1.40	0.73					0.20	0.66	0.12
Stimulation- LF n.u.	37.36	14.57	36.84	20.09	37.11	17.26					0.32	0.58	0.15
Stimulation- HF n.u.	49.02	18.15	52.7	20.59	50.77	19.27	0.03	0.52	0.67	0.34	0.00	1.00	0.00
Stimulation-SCR	0.19	0.25	0.14	0.11	0.17	0.20					0.04	0.84	0.05
Recovery- LF n.u.	41.25	15.99	40.58	19.56	40.93	17.62			•	•	0.10	0.76	0.08
Recovery- HF n.u.	45.42	17.75	46.58	18.12	45.97	17.78	0.00	0.05	0.99	0.10	0.06	0.81	0.06
Recovery- SCL	1.39	0.52	1.52	0.86	1.45	0.70					0.07	0.79	0.07

Note: Pillai's V refers to MANOVA test statistics; F refers to the F-statistics; p refers to the significance level set at p < 0.05; d refers to the effect size expressed in Cohen's d; *significant after Bonferroni Adjusted p = 0.017

E. Summary of Results for Hypothesis Testing 1

The influence of ethnicity on the adaptation to sensory stimuli using an auditory stimulus was examined by looking at the neurophysiological regulation of the ANS using HRV and EDA. There was significant interaction between condition and participant grouping (CHK and FHK) only for the mean change in SCL. The results of this thesis suggest that ethnicity may have influenced the direction of change in sympathetic response to sensory stimuli among children. However, individual group characteristics reflecting the regulation of response to sensory stimuli can be noted.

Using the HF n.u. and SCR as representative measures of PNS and SNS activity respectively, there are interesting patterns of change in the neurophysiological regulation of response to sensory stimuli can be seen exhibited by each group. There is a significant increase in SCR activity with the maintenance of the HF n.u. activity from resting to stimulation conditions seen in the FHK group. This is in contrast to the CHK group who exhibited maintenance of both HF n.u. and SCR activity.

Indexing the change in the activity of the HF n.u. and SCR when auditory stimulation is removed, from stimulation to recovery conditions, suggests another set of interesting results. The FHK group showed a significant decrease in their SCR while the HF n.u. is maintained. In contrast, the CHK groups' data showed maintenance of HF n.u. and SCR activity at baseline with no significant pattern of change.

To determine the overall autonomic activity of the HF n.u. and SCR, this thesis looks at the patterns of change from resting to recovery conditions. The FHK group showed baseline control of HF n.u. and SCR, which suggests that at recovery conditions, autonomic activity was the same at resting condition. On the other hand, the CHK group displays a significant change in the HF n.u. with a significant increase of SCR seen at recovery conditions compared to the resting conditions.

This section compared the ANS activity across conditions between CHK and FHK groups. This thesis found significant differences between these two groups that have different ethnicities but living in the same geographic environments in their SCL patterns of change from one condition to another. Furthermore, this thesis found no significant differences between CHK and FHK groups on the level of

ANS activity in the three experimental conditions. Ethnicity alone may not be able to explain the differences in the neurophysiological regulation of response to sensory stimuli in children.

II. Research Hypothesis 2

A. Overview

Hypothesis 2 was tested. The influence of geographic environments on the adaptation to sensory stimuli using an auditory stimulus was examined. This thesis hypothesizes that Filipino children living in Hong Kong and Filipino children living in urban Philippines will have significantly different neurophysiological regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions. A baseline-corrected 2x3 mixed factorial ANOVA with follow-up Bonferroni correction (α = 0.05) was conducted with follow-up MANOVA test differences between FHK and FU group's autonomic activity (LF n.u., HF n.u., SCR/SCL) at each condition (resting, stimulation and recovery).

B. Summary of Participant Characteristics

Table 5.2.1 presents the summary of participant characteristics (n=82) between typically developing Filipino children living in Hong Kong (FHK; n=28) and Filipino children living in urban Philippines (FU; n=54). 64.30% among the FHK group were native-born compared to 100% native-born FU children. For both groups, the majority were males (85.71% in FHK and 61.02% in FU). On average, the FHK group is slightly older M=9.57 yr. (SD=1.91) compared to the FU children at M=8.92 yr. (SD=1.71). BMI was slightly different between groups where the FU children were slightly lower at M=18.41 (SD=4.78) compared to their FHK counterparts who registered at M=18.4118.83 (SD= 3.54). All of the FHK children attend public schools, while a majority of the FU participants (70.40%) attends private school. Both parents are working for the FHK group, while only 66.70% was for the FU children. The mothers were identified as the primary caregiver of the participating children (100% for FHK; 98.10% for FU). 100% of the mothers of the FHK group attended collegiate education, while 98.10% of the identified primary caregivers for the FU group. There were more families in the FHK group (71.40%) classified as upper middle-income households compared to 14.80% in the FU families. Majority of the FHK children belong to families in the socio-economic class AB (96.40%) compared to the FU group whose majority belongs to class C (48.10%).

Table 5.2.1. Summary of Participant Characteristics Between FHK and FU Groups (*n*= 82).

D. A'.' and Classical distance	FHK	FU
Participant Characteristics	n=28	n=54
Age in yrs. $M(SD)$	9.57 (1.91)	8.91 (1.71)
BMI in $Kg/M^2 M(SD)$	18.83 (3.54)	18.41 (4.78)
Migration Status ^a	64.30% Native Born	100% Native Born
Gender ^a	85.71% Male	61.02% Male
School Type	100% Public School	70.40% Private School
No. of Parents Working	100 % 2 Parents	66.70% 2 Parents
Primary Caregiver	100% Mother	98.10% Mother
Educational Level of Primary Caregiver	100% College Level	94.40% College Level
Income Classification ^b	71.40% Upper Middle Income	48.10% Middle Income
Socio-Economic Classification ^{b,c}	96.4% Class AB	48.10% Class C

Note: anot similar at baseline p>0.05, thus used as a covariate in subsequent MANOVA tests; for the classification of income and socio-economic status, see Appendix 4.A; socio-economic Status ranges from Class A-E, with Class A representing richer families and Class E representing poorer families

C. Differences in the Patterns of Neurophysiological Regulation Between FHK and FU Groups

To answer the question on whether geographic environments influence the regulation of response to sensory stimuli, a baseline-corrected 2x3 mixed factorial ANCOVA (using migration status and gender as covariates) with follow-up Bonferroni correction (α =0.05) was conducted to determine the effect of ethnicity and environments on neurophysiological measures of regulation of response to sensory stimuli (LF n.u., HF n.u., SCR, SCL) across conditions (resting, stimulation, recovery). Differences in the change of neurophysiological parameters related to the regulation of response to an auditory stimulus between FHK and FU groups were examined. This group (FHK and FU) represents children from similar ethnicities living in different geographic environments but of similar urbanization characteristics. Table 5.2.2 summarises the main effects of pairwise (FHK and FU) baseline-corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter. Table 5.2.3 summarises the condition pairwise (FHK and FU) baseline-corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter.

i. LF n.u.

Changes in the mean LF n.u. significantly varied across conditions (F(2,156)= 9.01, MS= 910.08, p<0.00). When factored using gender as a covariate, the model still yields significant differences of p = 0.05. There was no significant interaction between condition and participant within the FHK and FU groups for the changes in the LF n.u. (F(1, 110) = 0.38, MS =150.89, p = 0.54). Within-group condition pairwise comparisons did suggest significant differences in the direction of change of LF n.u. values for stimulation to recovery conditions (MD = -5.61, p < 0.00). While LF n.u. values decreased during stimulation condition, the change from resting condition in LF n.u. levels were not significant. However, there is a significant continuing increase of the LF n.u. from auditory stimulation condition to recovery condition. Generally, between FHK and FU groups, the LF n.u. levels at recovery condition were not significantly increased from resting condition even after stimulation. Figure 5.2.1 displays the covariate (migration status and gender) adjusted estimated marginal means of the LF n.u. for this 2x3 mixed factorial ANCOVA between FHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the LF n.u. did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different geographic environments (FHK & FU).

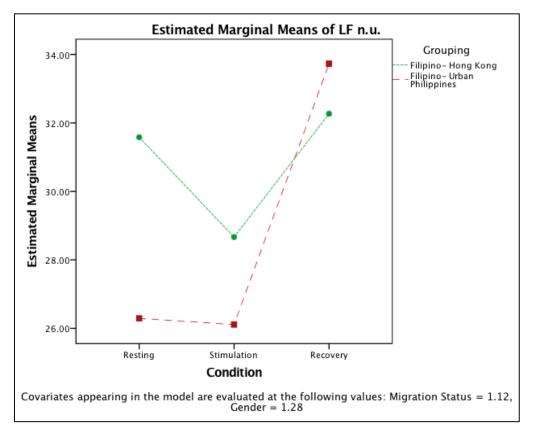


Figure 5.2.1. Covariate (migration status and gender) adjusted estimated marginal means for LF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (FHK and FU).

ii. HF n.u.

Changes in the mean HF n.u. significantly varied across conditions (F(2, 110) = 11.88, MS = 1243.48, p < 0.00). There was no significant interaction between condition and participant grouping for the HF n.u. measure (F(1, 110) = 0.84, MS = 596.20, p = 0.36). Condition pairwise comparisons did suggest significant differences in the direction of change of HF n.u. values in stimulation to recovery conditions (MD = 5.45, p = 0.01) and resting to recovery conditions (MD = 6.44, p < 0.00). The results suggest that HF n.u. significantly decreased during the stimulation condition. At recovery conditions, the HF n.u. levels were significantly decreased from the resting condition. Figure 5.2.2 displays the covariate (migration status and gender) adjusted estimated marginal means of the HF n.u. for this 2x3 mixed factorial ANCOVA between FHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the HF n.u. did not have significant interaction between condition and participant

grouping among children having similar ethnicities living in different geographic environments (FHK and FU).

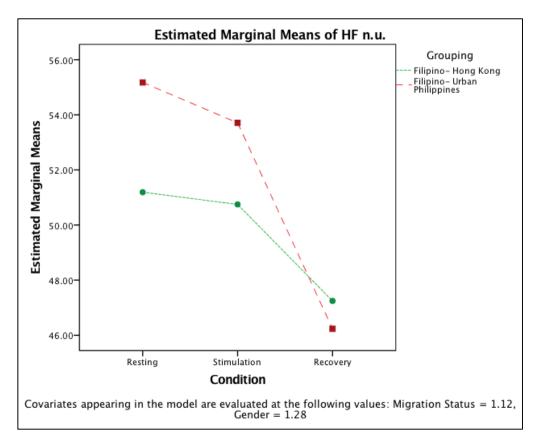


Figure 5.2.2. Covariate (migration status and gender) adjusted estimated marginal means for HF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (FHK and FU).

iii. SCR

A baseline-corrected 2x3 mixed factorial ANCOVA (using migration status and BMI as covariates) was carried out for the neurophysiological variable, SCR, and as Mauchly's test was significant (p<0.05), the Greenhouse-Geisser correction was applied. Mean changes in the SCR values significantly varied across conditions (F(1.67, 110)= 50.53, MS= 0.11, p<0.00) within-group (FHK and FU). However, within groups interaction between experimental condition and participant grouping for the SCR measure did not reach significance thresholds (F(1.67, 110)= 1.11, MS= 0.02, p= 0.30). Nevertheless, condition pairwise comparisons did suggest significant differences in the direction of change of SCR values only for resting to stimulation condition (MD= -0.06, p<0.00) and stimulation to

recovery conditions (MD= 0.05, p<0.00). The results suggest an interesting difference in the regulation of response where there is a significant increase of the SCR upon auditory condition; and consequent decrease of the same SCR parameter after auditory stimulation as measured during the recovery condition. At recovery conditions, SCR values were not significantly increased from the resting conditions. Figure 5.2.3 displays the covariate (migration status and gender) adjusted estimated marginal means of the SCR for this 2x3 mixed factorial ANCOVA between FHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCR did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different geographic environments (FHK and FU).

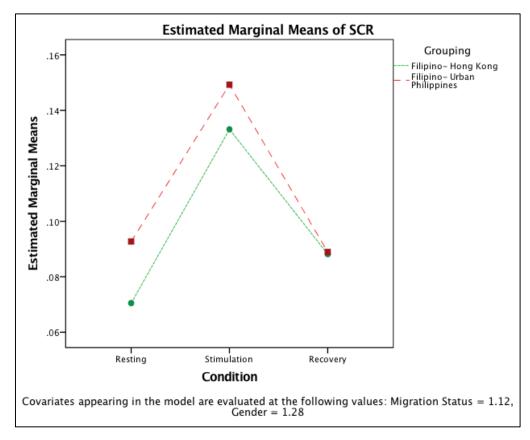


Figure 5.2.3. Covariate (migration status and gender) adjusted estimated marginal means for SCR using baseline-corrected 2x3 mixed factorial ANCOVA (FHK and FU).

iv. SCL

The Greenhouse-Geisser correction was employed due to a significant Mauchly's test (p< 0.05) for a baseline-corrected 2x3 mixed factorial ANCOVA (using migration status and gender as covariates) in exploring the SCL measure. Changes in the mean SCL values significantly varied across conditions (F(1.28, 99.92) = 7.85, MS = 1.28, p < 0.00) within-group (FHK and FU). There was likewise consequent significant within-group differences on the changes in the SCL parameter when gender was factored as a covariate (p= 0.01). However, within groups interaction between condition and participant grouping (FHK and FU) for the changes in the SCL measure did not reach significance thresholds (F(1.28, 99.92) = 1.11, MS=0.06, p=0.16. Furthermore, there was a significant increase in the SCL direction of change from resting to stimulation (MD= -0.10, p<0.00), and decrease stimulation to recovery (MD=0.04, p=0.02) after the follow-up condition pairwise comparisons. This suggests that there was a pattern of increasing SCL after auditory stimulation from resting, and a consequent decreasing pattern (down-regulation) of the SCL parameter as the child recovers after the said sensory stimulation. At recovery, the SCL level was not significantly different from the resting condition. Figure 5.2.4 displays the covariate (migration status and gender) adjusted estimated marginal means of the SCL for this 2x3 mixed factorial ANCOVA between FHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCL did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different geographic environments (FHK and FU).

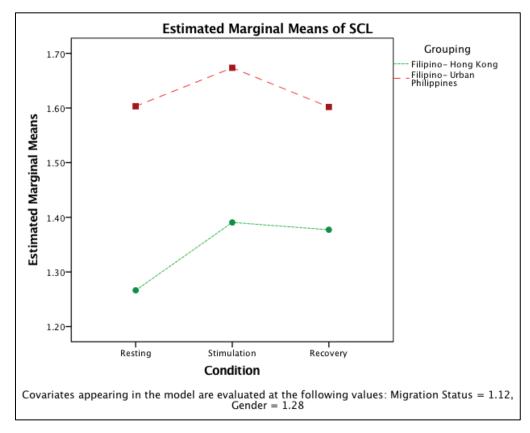


Figure 5.2.4. Covariate (migration status and gender) adjusted estimated marginal means for SCL using baseline-corrected 2x3 mixed factorial ANCOVA (FHK and FU).

Table 5.2.2. Summary of Differences in the Patterns of Neurophysiological Regulation Between FHK and FU Groups Using Main Effects of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

			Unco	rrected Ma	in Effec	ts]	Baseline	Corrected	Main Ef	fects	
	Outcomes	SS	df	MS	F	Sig	ES	SS	df	MS	F	Sig	ES
LF n.u.	Within												
	Condition	1573.43	2	786.71	7.79	<0.00	0.09	1820.16	2	910.08	9.01	<0.00	0.10
	Condition * Migration	549.84	2	274.92	2.72	0.07	0.03	583.75	2	291.88	2.89	0.06	0.04
	Condition * Gender	709.23	2	354.62	3.51	0.03	0.04	623.69	2	311.85	3.09	<0.00	0.04
	Condition * Grouping	344.64	2	172.32	1.71	0.18	0.02	296.45	2	148.23	1.47	0.23	0.02
	Error	15750.68	156	100.97				15750.68	156	100.97			
	Between												
	Migration	84.44	1	84.44	0.18	0.67	0.00	464.03	1	464.03	1.00	0.32	0.01
	Gender	138.26	1	138.26	0.30	0.59	0.00	0.47	1	0.47	0.00	0.97	0.00
	Grouping	3943.92	1	3943.92	8.46	<0.00	0.10	174.67	1	174.67	0.37	0.54	0.00
	Error	36369.15	78	466.27				36369.15	78	466.27			
HF n.u.	Within												
	Condition	1542.34	2	771.17	7.37	<0.00	0.09	2486.97	2	1243.48	11.88	<0.00	0.13
	Condition * Migration	483.99	2	241.99	2.31	0.10	0.03	307.66	2	153.83	1.47	0.23	0.02
	Condition * Gender	412.58	2	206.29	1.97	0.14	0.02	394.31	2	197.16	1.88	0.16	0.02
	Condition * Grouping	60.66	2	30.33	0.29	0.75	0.00	178.93	2	89.47	0.85	0.43	0.01
	Error	16323.83	156	104.64				16323.83	156	104.64			
	Between												
	Migration	52.92	1	52.92	0.07	0.79	0.00	596.20	1	596.20	0.84	0.36	0.01
	Gender	357.65	1	357.65	0.50	0.48	0.01	162.19	1	162.19	0.23	0.63	0.00
	Grouping	1595.49	1	1595.49	2.23	0.14	0.03	150.89	1	150.89	0.21	0.65	0.00
	Error	55686.01	78	713.92				55686.01	78	713.92			

SCR^a Within

	Condition	0.16	1.67	0.10	45.92	<0.00	0.37	0.18	1.67	0.11	50.53	<0.00	0.39
	Condition * Migration	0.00	1.67	0.00	1.07	0.33	0.01	0.00	1.67	0.00	0.53	0.56	0.01
	Condition * Gender	0.00	1.67	0.00	0.96	0.37	0.01	0.00	1.67	0.00	0.81	0.43	0.01
	Condition * Grouping	0.00	1.67	0.00	0.24	0.75	0.00	0.00	1.67	0.00	0.89	0.40	0.01
	Error	0.27	130.50	0.00				0.27	130.50	0.00			
	Between												
	Migration	0.00	1	0.00	0.04	0.85	0.00	0.02	1	0.02	1.11	0.30	0.01
	Gender	0.00	1	0.00	0.10	0.75	0.00	0.00	1	0.00	0.00	0.99	0.00
	Grouping	0.08	1	0.08	3.74	0.06	0.05	0.01	1	0.01	0.30	0.59	0.00
	Error	1.73	78	0.02				1.73	78	0.02			
SCL^a	Within												
	Condition	0.58	1.28	0.45	13.93	<0.00	0.15	0.33	1.28	0.25	7.84	<0.00	0.09
	Condition * Migration	0.19	1.28	0.15	4.64	0.02	0.06	0.09	1.28	0.07	2.12	0.14	0.03
	Condition * Gender	0.33	1.28	0.26	8.03	<0.00	0.09	0.28	1.28	0.22	6.82	0.01	0.08
	Condition * Grouping	0.00	1.28	0.00	0.11	0.80	0.00	0.08	1.28	0.06	1.95	0.16	0.02
	Error	3.24	99.92	0.03				3.24	99.92	0.03			
	Between												
	Migration	5.37	1	5.37	3.83	0.05	0.05	9.78	1	9.78	6.97	0.01	0.08
	Gender	1.48	1	1.48	1.06	0.31	0.01	0.86	1	0.86	0.61	0.44	0.01
	Grouping	9.32	1	9.32	6.65	0.01	0.08	3.06	1	3.06	2.18	0.14	0.03
	Error	109.35	78	1.40				109.35	78	1.40			

Note: LF n.u.= HRV- Low Frequency normalized unit; HF n.u.= HRV- High Frequency normalized unit; SCR- EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; SS= Sum of Squares; df= Degrees of Freedom; MS= Mean Squared; ES= effect size (Cohen's d); Sig= significant difference at p < 0.05; asignificant (p < 0.05) Mauchly's test

Table 5.2.3. Summary of Differences in the Patterns of Neurophysiological Regulation Between FHK and FU Groups Using Condition Pairwise (FHK and FU) baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

					Basel	line Corre	cted Co	ndition l	Pairwise	Compa	risons				
]	Rest-Stin	1			S	tim-Reco	V]	Rest-Rec	OV	
				95%	6 CI				95%	6 CI				95%	% CI
Outcomes	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB
LF n.u.	1.55	1.57	0.99	-2.30	5.40	-5.61*	1.59	<0.00	-9.49	-1.72	-4.06	1.89	0.10	-8.69	0.56
HF n.u.	0.95	1.61	1.00	-2.98	4.88	5.49*	1.68	<0.00	1.38	9.59	6.44*	1.87	<0.00	1.87	11.01
SCR^a	-0.06*	0.01	<0.00	-0.08	-0.04	0.05*	0.01	<0.00	0.03	0.07	-0.01	0.01	0.59	-0.02	0.01
SCL^a	-0.10*	0.02	<0.00	-0.15	-0.04	0.04*	0.01	0.02	0.01	0.08	-0.05	0.03	0.26	-0.13	0.02

Note: LF n.u.= HRV- Low frequency normalized unit; HF n.u.= HRV- High frequency normalized unit; SCR: EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; MD= Mean Difference; SE= Standard Error; LB= Lower Bound limit; UB: Upper Bound limit; Sig= significant difference at p < 0.05; *significant at p < 0.05; *significant (p < 0.05) Mauchly's test

D. Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between FHK and FU groups

i. Resting Condition

For the resting condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (FHK: n= 28, age= 9.57 yr., BMI= 18.826 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA, and migration status and gender as covariates (see Table 5.2.1).

There is significant multivariate group mean difference between the two groups (A= 0.23, F(3,32)= 7.42, p<0.00, d= 1.08) on LF n.u. (FHK: M= 34.85; SD= 15.29; FU: M= 28.10; SD= 14.10), HF n.u. (FHK: M= 53.81; SD= 16.60; FU: M= 57.91; SD= 17.72), and SCL (FHK: M= 1.49; SD= 0.94; FU: M= 1.64; SD= 0.54). Table 5.2.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at resting condition among children having similar ethnicities living in different geographic environments (FHK and FU).

ii. Stimulation Condition

For the stimulation condition, LF n.u., HF n.u. and SCR were compared between the two groups of participants (FHK: n= 28, age= 9.57 yr., BMI= 18.826 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA, and migration status and gender as covariates (see Table 5.2.1).

There is significant multivariate group mean difference between the two groups (Λ = 0.14, F(3,32)= 4.26, p= 0.01, d= 0.82) on LF n.u. (FHK: M= 36.84; SD= 20.09; FU: M= 26.98; SD= 12.74), HF n.u. (FHK: M= 52.70; SD= 20.59; FU: M= 55.93; SD= 17.87), and SCR (FHK: M= 0.65; SD= 0.67; FU: M= 0.43; SD= 0.57). Table 5.2.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is no significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCR at

stimulation condition among children having similar ethnicities living in different geographic environments (FHK and FU).

iii. Recovery Condition

For the recovery condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (FHK: n= 28, age= 9.57 yr., BMI= 18.826 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA, and migration status and gender as covariates (see Table 5.2.1).

The multivariate group mean difference between the two groups approached statistical thresholds (Λ = 0.10, F(3,32)= 2.712, p= 0.05, d= 0.65) on LF n.u. (FHK: M= 40.58; SD= 19.56; FU: M= 33.23; SD= 11.97), HF n.u. (FHK: M= 46.58; SD= 18.12; FU: M= 52.62; SD= 15.34), and SCL (FHK: M= 1.52; SD= 0.86; FU: M= 1.78; SD= 0.67). Table 5.2.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is no significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at recovery condition among children having similar ethnicities living in different geographic environments (FHK and FU).

Table 5.2.4. Summary of Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between FHK and FU groups Using MANOVA Tests (n= 82).

	FH	łK	F	U	To	tal		MAN(OVA			ANOVA	
	n=	-28	n=	54	n=	82	- Pillai's V	F		J	F		
	M	SD	M	SD	M	SD	- Pillat S V	Г	p	d	Г	p	d
Resting- LF n.u.	34.85	15.29	28.1	14.1	30.4	14.78					9.34	<0.00*	0.69
Resting- HF n.u	53.81	16.6	57.91	17.72	56.51	17.35	0.23	7.42	<0.00*	1.08	2.79	0.10	0.38
Resting- SCL	1.49	0.94	1.64	0.54	1.59	0.7					7.30	0.01*	0.61
Stimulation- LF n.u.	36.84	20.09	26.98	12.74	30.35	16.21					8.25	0.01*	0.65
Stimulation- HF n.u.	52.7	20.59	55.93	17.87	54.83	18.78	0.14	4.26	0.01*	0.82	1.40	0.24	0.27
Stimulation-SCR	0.14	0.11	0.17	0.10	0.16	0.11					1.09	0.30	0.24
Recovery- LF n.u.	40.58	19.56	33.23	11.97	35.74	15.28					1.95	0.17	0.32
Recovery- HF n.u.	46.58	18.12	52.62	15.34	50.56	16.49	0.10	2.71	0.05	0.65	1.20	0.28	0.25
Recovery- SCL	1.52	0.86	1.78	0.67	1.69	0.75					5.98	0.02*	0.55

Note: Pillai's V refers to MANOVA test statistics; F refers to the F-statistics; p refers to the significance level set at p < 0.05; d refers to the effect size expressed in Cohen's d; *significant after Bonferroni Adjusted p = 0.017

E. Summary of Hypothesis Testing 2

The influence of geographic environments on the adaptation to sensory stimuli using an auditory stimulus was examined by looking at the neurophysiological regulation of the ANS using HRV and EDA. There was no significant interaction between condition and participant grouping (FHK and FU) across neurophysiological measures of regulation of response to sensory stimuli. The results of this thesis suggest that geographic environments may not be sufficient to influence the direction of change in the neurophysiological measures in response to sensory stimuli among children.

Using the HF n.u. and SCR as representative measures of PNS and SNS activity respectively, there are interesting patterns of change in the neurophysiological regulation of response to sensory stimuli can be seen exhibited by each group. There is a significant increase in SCR activity with the maintenance of the HF n.u. activity from resting to stimulation conditions seen in the FHK and FU groups.

Indexing the change in the activity of the HF n.u. and SCR when auditory stimulation is removed, from stimulation to recovery conditions, suggests another set of interesting results. Both the FHK and FU groups showed a significant decrease in their SCR while the HF n.u. is maintained.

To determine the overall autonomic activity of the HF n.u. and SCR, this thesis looks at the patterns of change from resting to recovery conditions. Both the FHK and FU groups were comparable in the fact that there was baseline control of HF n.u. and SCR.

This section compared the ANS activity across conditions between FHK and FU groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis finds differences between the two groups having similar ethnicities but living in different geographic environments on their levels of autonomic activity, but not on the autonomic patterns of change from one condition to another. Results suggest that the geographic environments may have the ability to override ethnicity effects and influence the neurophysiological regulation of response to sensory stimuli in children.

III. Research Hypothesis 3

A. Overview

Hypothesis 3 was tested. The influence of ethnicity and geographic environments on the adaptation to sensory stimuli using an auditory stimulus was examined. This thesis hypothesizes that Chinese children living in Hong Kong and Filipino children living in urban Philippines will have significantly different neurophysiological regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions. A baseline-corrected 2x3 mixed factorial ANOVA with follow-up Bonferroni correction (α = 0.05) was conducted with follow-up MANOVA test differences between CHK and FU group's autonomic activity (LF n.u., HF n.u., SCR/SCL) at each condition (resting, stimulation and recovery).

B. Summary of Participant Characteristics

Table 5.3.1 presents the summary of participant characteristics (n=85) between typically developing Chinese children living in Hong Kong (CHK; n=31) and Filipino children living in urban Philippines (FU; n=54). All children in this pairwise combination was native-borne. For both groups, the majority were males (65.63% in CHK and 61.02% in FU). On average, the FU group is slightly older at M = 8.92 yr. (SD = 1.71) compared to the CHK children at M = 8.59 yr. (SD = 1.58). BMI was slightly different between groups where the CHK children lower at M=17.09 (SD = 2.72) compared to their FU counterparts who registered at M = 18.41(SD = 4.78). Majority of the children attended private school with the FU children slightly higher at 70.40% than the 63.33% of the CHK children. In both groups, majority of the children belonged to a two-parent working household (100% for CHK, 66.70% for FU), the mother was identified as the primary caregiver (100%) for CHK, 98.10% for FU) with at least a college-level educational attainment (100% for CHK, 94.40% FU). Among the CHK group, 38.70% were classified to be upper middle income and upper income, while a majority of 48.80% in the FU families were deemed to be middle income. Majority of the CHK families belong to socio-economic class AB, while 48.10% of the FU families at class C.

Table 5.3.1. Summary of Participant Characteristics Between CHK and FU Groups (n=85).

Participant	СНК	FU
Characteristics	n=31	n=54
Age in yrs. $M(SD)$	8.59 (1.58)	8.91 (1.71)
BMI in Kg/M^2 M(SD)	17.09 (2.72)	18.41 (4.78)
Migration Status	100% Native Born	100% Native Born
Gender	65.63% Male	61.02% Male
Sahaal Tura	63.33% Private	70.40%
School Type	School	Private School
No. of Parents	100% 2 Parents	66.70% 2
Working	100% 21 drents	Parents
Primary Caregiver	100% Mother	98.10%
Timary Caregiver	100% Woller	Mother
Educational Level of	100% College	94.40%
Primary Caregiver	Level	College Level
Income	29.700/ Hansa	48.10%
Income Classification ^b	38.70% Upper	Middle
Classification	Middle Income	Income
Socio-Economic	02 50/ Cl AP	48.10% Class
Classification ^{b,c}	93.5% Class AB	C

Note: anot similar at baseline p>0.05, thus used as a covariate in subsequent MANOVA tests; for the classification of income and socio-economic status, see Appendix 4.A; socio-economic Status ranges from Class A-E, with Class A representing richer families and Class E representing poorer families

C. Differences in the Patterns of Neurophysiological Regulation Between CHK and FU Groups

To answer the question on whether geographic environments and ethnicity influence the regulation of response to sensory stimuli, a baseline-corrected 2x3 mixed factorial with follow-up Bonferroni correction (α = 0.05) was conducted to determine the effect of ethnicity and environments on neurophysiological measures of regulation of response to sensory stimuli (LF n.u., HF n.u., SCR, SCL) across conditions (resting, stimulation, recovery). Differences in the change of neurophysiological parameters related to the regulation of response to an auditory stimulus between CHK and FU groups were examined. This group represents children from different ethnicities living in different geographic environments but of similar urbanization characteristics. Table 5.3.2 summarises the main effects of pairwise (CHK and FU) baseline-corrected 2x3 mixed factorial ANOVA by

neurophysiological parameter. Table 5.3.3 summarises the condition pairwise (CHK and FU) baseline-corrected 2x3 mixed factorial ANOVA by neurophysiological parameter.

i. LF n.u.

Changes in the mean LF n.u. significantly varied across conditions (F(2, 156)= 9.01, MS= 910.08, p<0.00). When factored using gender as a covariate, the model still yields significant differences of p=0.05. There was no significant interaction between condition and participant grouping for the LF n.u. measure (F(1, 110) = 0.38, MS = 150.89, p = 0.54). Condition pairwise comparisons did suggest significant differences in the direction of change of LF n.u. values only for stimulation to recovery conditions (MD = -5.601, p < 0.00). Thus, there is a significant increase in the LF n.u. after auditory stimulation when measured during the recovery condition. LF n.u. values did not significantly decrease during the stimulation condition. At recovery conditions, LF n.u. values were not significantly increased compared to resting conditions. Figure 5.3.1 displays the estimated marginal means of the LF n.u. for this 2x3 mixed factorial ANOVA between CHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the LF n.u. did not have significant interaction between condition and participant grouping among children having different ethnicities living in different geographic environments (CHK and FU).

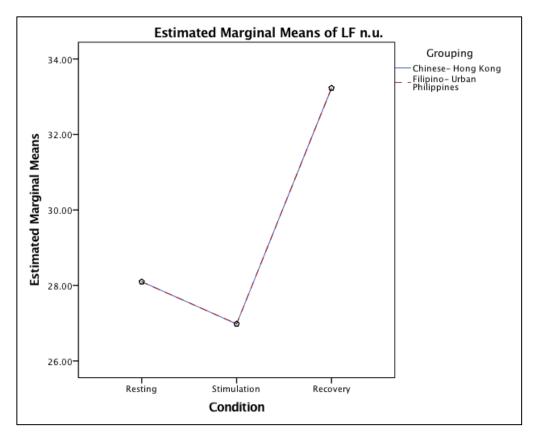


Figure 5.3.1. Estimated marginal means for LF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FU).

ii. HF n.u.

The changes in the mean HF n.u. significantly varied across conditions (F(2, 110)= 11.88, MS= 1243.48, p<0.00). There was no significant interaction between condition and participant grouping for the LF n.u. measure (F(1, 110)= 0.84, MS= 596.20, p= 0.36). Condition pairwise comparisons did suggest significant differences in the direction of change of HF n.u. values for stimulation to recovery conditions (MD= 5.448, p= 0.01) and resting to recovery conditions (MD= 6.44, p<0.00). Thus, there is a significant and continuous decrease in the HF n.u. during auditory stimulation when measured during recovery conditions. At stimulation conditions, HF n.u. was not significantly decreased from resting condition values. However, at recovery conditions, there was a significant decrease in HF n.u. values from resting and stimulation conditions after auditory stimulation. Figure 5.3.2 displays the estimated marginal means of the HF n.u. for this 2x3 mixed factorial ANOVA between CHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the HF n.u. did not have significant

interaction between condition and participant grouping among children having different ethnicities living in different geographic environments (CHK and FU).

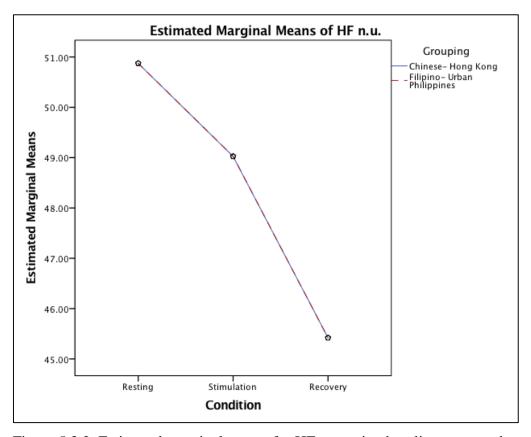


Figure 5.3.2. Estimated marginal means for HF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FU).

iii. SCR

A baseline-corrected 2x3 mixed factorial ANOVA was carried out for the physiological variable, SCR, and as Mauchly's test was significant (p<0.05), the Greenhouse-Geisser correction was applied. Mean changes in the SCR values significantly varied across conditions (F(1.67, 110)= 50.53, MS= 0.11, p<0.00. The results suggest that the mean SCR values within-participant group (CHK and FU) significantly varied across conditions. However, within groups interaction between experimental condition and participant grouping for the SCR measure did not reach significance thresholds (F(1.67, 110)= 1.11, MS= 0.02, p= 0.30). Nevertheless, condition pairwise comparisons did suggest significant differences in the change of SCR values only for resting to stimulation condition (MD= -0.06, p<0.00) and stimulation to recovery conditions (MD= 0.05, p<0.00). The results suggest an interesting difference in

the regulation response where there is a significant increase of the SCR during auditory stimulation condition; and consequent decrease of the same SCR parameter at the recovery condition. Resting and recovery condition SCR values were not significantly different. Figure 5.3.3 displays the estimated marginal means of the SCR for this 2x3 mixed factorial ANOVA between CHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCR did not have significant interaction between condition and participant grouping among children having different ethnicities living in different geographic environments (CHK and FU).

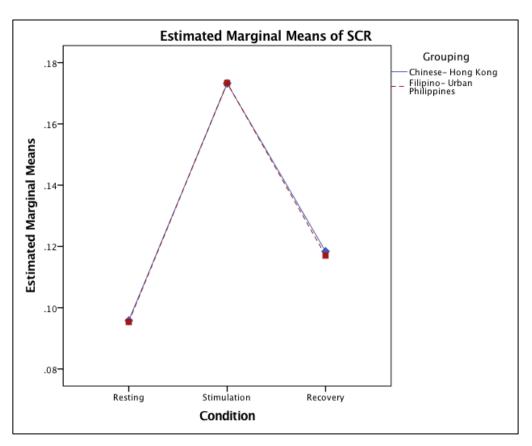


Figure 5.3.3. Estimated marginal means for SCR using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FU).

iv. SCL

The Greenhouse-Geisser correction was employed due to a significant Mauchly's test (p<0.05) for a baseline-corrected 2x3 mixed factorial ANOVA in exploring changes in the SCL measure. Changes in the mean SCL values significantly varied across conditions (F(1.28, 99.92)= 7.85, MS= 1.28, p<0.00) within CHK and FU groups. There was likewise consequent significant within-

group differences on SCL parameters when gender was factored as a covariate (p=0.01). However, within groups (CHK and FU) interaction between condition and participant grouping for the SCR measure did not reach significance thresholds (F(1.28, 99.92) = 1.12, MS = 0.06, p = 0.16). The results suggest that the mean SCR values within the two participant groups did not significantly vary across conditions. Furthermore, there was a significant difference in the SCL change from resting to stimulation (MD = -0.10, p < 0.00), stimulation to recovery (MD= 0.04, p= 0.02) after the follow-up condition pairwise comparisons. This suggests an interesting regulation of response pattern, where there was a significant increase of the SCL during auditory stimulation from resting; and a consequent significant decrease of the SCL parameter as the child recovers after the said sensory stimulation. There was no significant difference between SCL values at resting and recovery conditions. Figure 5.3.4 displays the estimated marginal means of the SCL for this 2x3 mixed factorial ANOVA between CHK and FU groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCL did not have significant interaction between condition and participant grouping among children having different ethnicities living in different geographic environments (CHK and FU).

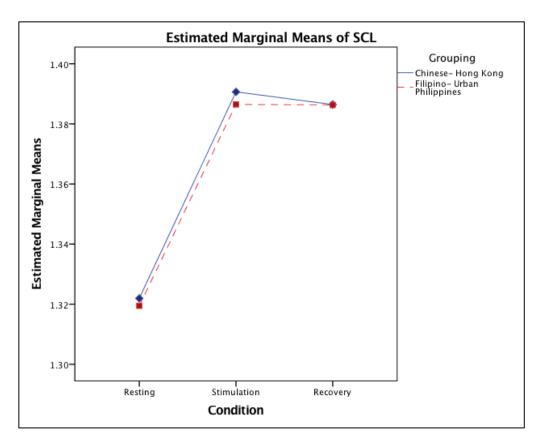


Figure 5.3.4. Estimated marginal means for SCL using baseline-corrected 2x3 mixed factorial ANCOVA (CHK and FU).

Table 5.3.2. Summary of Differences in the Patterns of Neurophysiological Regulation Between CHK and FU Groups Using Main Effects of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

			Unco	rrected Ma	in Effect	ts		H	Baseline C	orrected l	Main Eff	fects	
	Outcomes	SS	df	MS	F	Sig	ES	SS	df	MS	F	Sig	ES
LF n.u.	Within												
	Condition	1493.25	2	746.63	7.92	<0.00	0.09	1888.06	2.00	944.03	10.02	<0.00	0.11
	Condition * Grouping	56.38	2	28.19	0.30	0.74	0.00	0.00	2.00	0.00	0.00	1.00	0.00
	Error	15642.89	166	94.23				15642.89	166.00	94.23			
	Between												
	Grouping	4870.34	1	4870.34	12.43	< 0.00	0.13	0.00	1.00	0.00	0.00	1.00	0.00
	Error	32528.86	83	391.91				32528.86	83.00	391.91			
HF n.u.	Within												
	Condition	1246.13	2	623.07	6.07	<0.00	0.07	1308.26	2.00	654.13	6.37	<0.00	0.07
	Condition * Grouping	0.87	2	0.43	0.00	1.00	0.00	0.00	2.00	0.00	0.00	1.00	0.00
	Error	17051.10	166	102.72				17051.10	166.00	102.72			
	Between												
	Grouping	2932.07	1	2932.07	4.10	0.05	0.05	0.00	1.00	0.00	0.00	1.00	0.00
	Error	59319.62	83	714.69				59319.62	83.00	714.69			
SCRa	Within												
	Condition	0.25	1.11	0.22	15.35	< 0.00	0.16	0.27	1.11	0.25	17.02	< 0.00	0.17
	Condition * Grouping	0.02	1.11	0.01	1.02	0.32	0.01	0.00	1.11	0.00	0.00	0.98	0.00
	Error	1.33	91.91	0.01				1.33	91.91	0.01			
	Between												
	Grouping	0.00	1	0.00	0.02	0.90	0.00	0.00	1.00	0.00	0.00	0.98	0.00
	Error	2.26	83	0.03				2.26	83.00	0.03			

SCL^a	Within												
	Condition	0.59	1.35	0.44	12.45	<0.00	0.13	0.25	1.35	0.19	5.36	0.01	0.06
	Condition * Grouping	0.04	1.35	0.03	0.89	0.38	0.01	0.00	1.35	0.00	0.00	0.98	0.00
	Error	3.92	112.06	0.03				3.92	112.06	0.03			
	Between												
	Grouping	8.28	1	8.28	8.75	<0.00	0.10	0.00	1.00	0.00	0.00	0.99	0.00
	Error	78.59	83	0.95				78.59	83.00	0.95			

Note: LF n.u.= HRV- Low Frequency normalized unit; HF n.u.= HRV- High Frequency normalized unit; SCR- EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; SS= Sum of Squares; df= Degrees of Freedom; MS= Mean Squared; ES= effect size (Cohen's d); Sig= significant difference at p<0.05; *asignificant (p<0.05) Mauchly's test

Table 5.3.3. Summary of Differences in the Patterns of Neurophysiological Regulation Between CHK and FU Groups Using Condition Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

					Base	line Corre	cted Co	ondition 1	Pairwise	Compa	risons				
]	Rest-Stin	1			S	tim-Reco	V			R	est-Reco	v	
				95%	6 CI				95%	6 CI				95%	6 CI
Outcomes	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB
LF n.u.	1.12	1.61	1.00	-2.82	5.05	-6.25*	1.38	<0.00	-9.62	-2.88	-5.13*	1.64	0.01	-9.13	-1.13
HF n.u.	1.85	1.61	0.76	-2.08	5.77	3.61	1.54	0.06	-0.15	7.36	5.45*	1.70	0.01	1.30	9.60
SCR^a	-0.08*	0.02	<0.00	-0.12	-0.04	0.06*	0.02	0.01	0.01	0.10	-0.02*	0.00	<0.00	-0.03	-0.01
SCL^a	-0.07*	0.03	0.03	-0.13	-0.01	0.00	0.01	1.00	-0.03	0.04	-0.07	0.03	0.10	-0.14	0.01

Note: LF n.u.= HRV- Low frequency normalized unit; HF n.u.= HRV- High frequency normalized unit; SCR: EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; MD= Mean Difference; SE= Standard Error; LB= Lower Bound limit; UB: Upper Bound limit; Sig= significant difference at p < 0.05; * significant at p < 0.05; asignificant (p < 0.05) Mauchly's test

D. Differences in the Neurophysiological Response to Sensory Stimuli Across
 Conditions Between CHK and FU Groups

i. Resting Condition

For the resting condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (CHK: n= 31, age= 8.55 yr., BMI= 17.23 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA, (see Table 5.3.1).

There is significant multivariate group mean difference between the two groups (A= 0.17, F(3,38) = 5.60, p<0.00, d= 0.91) on LF n.u. (CHK: M= 36.94; SD= 15.71; FU: M= 28.10; SD= 14.10), HF n.u. (CHK: M= 50.87; SD= 19.13; FU: M= 57.91; SD= 17.72), and SCL (CHK: M= 1.32; SD= 0.48; FU: M= 1.64; SD= 0.54). Table 5.3.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at resting condition among children having different ethnicities living in different geographic environments (CHK and FU).

ii. Stimulation Condition

For the stimulation condition, LF n.u., HF n.u. and SCR were compared between the two groups of participants (CHK: n= 31, age= 8.55 yr., BMI= 17.23 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA (see Table 5.3.1).

There is significant multivariate group mean difference between the two groups (A= 0.15, F(3,38) = 4.66, p= 0.01, d= 0.83) on LF n.u. (CHK: M= 37.36; SD= 14.57; FU: M= 26.98; SD= 12.74), HF n.u. (CHK: M= 49.02; SD= 17.87; FU: M= 55.93; SD= 17.87), and SCR (CHK: M= 0.48; SD= 0.59; FU: M= 0.43; SD= 0.57). Table 5.3.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCR at

stimulation condition among children having different ethnicities living in different geographic environments (CHK and FU).

iii. Recovery Condition

For the recovery condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (CHK: n= 31, age= 8.55 yr., BMI= 17.23 kg/m²; FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²) by MANOVA (see Table 5.3.1).

The multivariate group mean difference between the two groups approached traditional significance thresholds (Λ = 0.14, F(3,38) = 4.44, p= 0.001, d= 0.81) on LF n.u. (CHK: M= 41.25; SD= 15.99; FU: M= 33.23; SD= 11.97), HF n.u. (CHK: M= 45.42; SD= 17.75; FU: M= 52.62; SD= 15.34), and SCL (CHK: M= 1.39; SD= 0.52; FU: M= 1.78; SD= 0.67). Table 5.3.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCR at stimulation condition among children having different ethnicities living in different geographic environments (CHK and FU).

Table 5.3.4. Summary of Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between CHK and FU Groups Using MANOVA Tests (n= 85).

	CI	HK	F	U	To	tal		MANO)VA			ANOVA	
	<u>n=</u>	:31	n=	54	n=	85	Pillai's V	$oldsymbol{F}$	n	d	${m F}$	n	d
	M	SD	M	SD	M	SD	T mai s v	ľ	p	и	ľ	p	и
Resting- LF n.u.	36.94	15.71	28.1	14.1	31.32	15.23					7.13	0.01*	0.59
Resting- HF n.u	50.87	19.13	57.91	17.72	55.34	18.45	0.17	5.60	<0.00*	0.91	2.93	0.09	0.38
Resting- SCL	1.32	0.48	1.64	0.54	1.52	0.54					7.38	0.01*	0.60
Stimulation- LF n.u.	37.36	14.57	26.98	12.74	30.77	14.26					11.76	<0.00*	0.75
Stimulation- HF n.u.	49.02	18.15	55.93	17.87	53.41	18.18	0.15	4.66	0.01*	0.83	2.91	0.09	0.37
Stimulation-SCR	0.19	0.25	0.17	0.10	0.18	0.17					0.14	0.71	0.08
Recovery- LF n.u.	41.25	15.99	33.23	11.97	36.16	14.03					6.89	0.01*	0.58
Recovery- HF n.u.	45.42	17.75	52.62	15.34	49.99	16.53	0.14	4.44	0.01*	0.81	3.86	0.05	0.43
Recovery- SCL	1.39	0.52	1.78	0.67	1.64	0.65					7.86	0.01*	0.62

Note: Pillai's V refers to MANOVA test statistics; F refers to the F-statistics; p refers to the significance level set at p < 0.05; d refers to the effect size expressed in Cohen's d; *significant after Bonferroni Adjusted p = 0.017

E. Summary of Hypothesis Testing 3

The influence of ethnicity and geographic environments on the adaptation to sensory stimuli using an auditory stimulus was examined by looking at the neurophysiological regulation of the ANS using HRV and EDA. There was no significant interaction between condition and participant grouping (CHK and FU) across neurophysiological measures of regulation of response to sensory stimuli. The results of this thesis suggest that ethnicity and geographic environments may not be sufficient to influence the direction of change in the neurophysiological measures in response to sensory stimuli among children.

Using the HF n.u. and SCR as representative measures of PNS and SNS activity respectively, there are interesting patterns of change in the neurophysiological regulation of response to sensory stimuli can be seen exhibited by each group. There is a significant increase in SCR activity with the maintenance of the HF n.u. activity from resting to stimulation conditions seen in the FU group. This is in contrast to the CHK group who exhibited maintenance of both HF n.u. and SCR activity.

Indexing the change in the activity of the HF n.u. and SCR when auditory stimulation is removed, from stimulation to recovery conditions, suggests another set of interesting results. The FU group showed a significant decrease in their SCR while the HF n.u. is maintained. In contrast, the CHK group's data showed maintenance of HF n.u. and SCR activity at baseline with no significant pattern of change.

To determine the overall autonomic activity of the HF n.u. and SCR, this thesis looks at the patterns of change from resting to recovery conditions. The FU group displayed baseline control of HF n.u. and SCR, which suggests that at recovery conditions, autonomic activity was the same at resting condition. On the other hand, the CHK group no significant change in the HF n.u. with a significant increase of SCR seen at recovery conditions compared to the resting conditions.

This section compared the ANS activity across conditions between CHK and FU groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis found differences between the two groups having

different ethnicities and geographic environments but living in similar physical environments. Results found that differences in ethnicity and geographic environment significantly influences the neurophysiological regulation of response to sensory stimuli in children.

IV. Research Hypothesis 4

A. Overview

Hypothesis 4 was tested. The influence of physical environments on the adaptation to sensory stimuli using an auditory stimulus was examined. This thesis hypothesizes Filipino children living in urban Philippines and Filipino children living in rural Philippines will have significantly different neurophysiological regulation of responses (HRV and EDA) to sensory stimuli at resting, stimulation and conditions. A baseline-corrected 2x3 mixed factorial ANOVA with follow-up Bonferroni correction (α = 0.05) was conducted with follow-up MANOVA test differences between FU and FR group's autonomic activity (LF n.u., HF n.u., SCR/SCL) at each condition (resting, stimulation and recovery).

B. Summary of Participant Characteristics

Table 5.4.1 presents the summary of participant characteristics (n=97) between typically developing Filipino children living in urban (FU; n=54) and rural (FR; n=43) Philippines. All children in this pairwise combination were native-borne. For both groups, the majority were males (61.02% in FU, 54.30% in FR). On average, the FU group is slightly younger at M = 8.92 yr. (SD = 1.71) compared to the FR children at M = 9.40 yr. (SD = 1.64). BMI was different between groups where the FR children are lower at M=15.64 (SD=3.01) compared to their FU counterparts who registered at M=18.41 (SD=4.78). Majority of the children attended a private school for the FU children at 70.40, while 97.70% of the FR children attended public school systems. 66.70% of the FU household have two parents working compared to 44.20% in the FR group. Majority of the identified primary caregiver was the mother in both groups (98.10% in FU, 93.00% in the FR), with at least a college-level educational attainment (94.40% in FU, 55.80% in FR). A majority of 48.80% in the FU families were deemed to be middle income compared to the 60.50% of families in the FR group that belong to lower middle income. Among the FU group, a majority of 48.10% belong to socio-economic class, while 60.50% of the FR group belong to class D.

Table 5.4.1. Summary of Participant Characteristics Between FU and FR Groups (n=97).

Participant	FU	FR
Characteristics	n=54	n=43
Age in yrs. $M(SD)$	8.91 (1.71)	9.39 (1.64)
BMI in $Kg/M^2 M(SD)^a$	18.41 (4.78)	15.64 (3.01)
Migration Status	100% Native Born	100% Native Born
Gender	61.02% Male	54.35% Male
School Type	70.40% Private School	97.70% Public School
No. of Parents Working	66.70% 2 Parents	44.20% 2 Parents
Primary Caregiver	98.10% Mother	93.00% Mother
Educational Level of Primary Caregiver	94.40% College Level	55.80% College Level
Income Classification ^b	48.10% Middle Income	60.50% Lower Middle Income
Socio-Economic Classification ^{b,c}	48.10% Class C	60.50% Class D

Note: anot similar at baseline p>0.05, thus used as a covariate in subsequent MANOVA tests; bfor the classification of income and socio-economic status, see Appendix 4.A; socio-economic Status ranges from Class A-E, with Class A representing richer families and Class E representing poorer families

C. Differences in the Patterns of Neurophysiological Regulation Between FU and FR Groups

To answer the question on whether physical environments influence the regulation of response to sensory stimuli, a baseline-corrected 2x3 mixed factorial ANCOVA (using BMI as a covariate) with follow-up Bonferroni correction (α=0.05) was conducted to determine the effect of physical environments on neurophysiological measures of regulation of response to sensory stimuli (LF n.u., HF n.u., SCR, SCL) across conditions (resting, stimulation, recovery). Differences in the change of neurophysiological parameters related to the regulation of response to auditory stimuli between FU and FR groups were examined. This group represents children from similar ethnicities living in the same geographic environments but of similar physical environment characteristics (i.e. urban vs rural). Table 5.4.2 summarises the main effects of pairwise (FU and FR) baseline-corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter. Table 5.4.3 summarises the condition

pairwise (FU and FR) baseline corrected 2x3 mixed factorial ANCOVA by neurophysiological parameter.

i. LF n.u.

Changes in the mean LF n.u. significantly varied across conditions (F(2,188)= 11.11, MS= 1076.60, p<0.00) within groups. There was no significant interaction between condition and participant grouping (FU and FR) for the LF n.u. measure (F(2, 188) = 0.10, MS = 9.22, p = 0.91). Condition pairwise comparisons did suggest significant differences in the direction of change of LF n.u. values only for stimulation to recovery conditions (MD= -6.17, p<0.00) and resting to recovery conditions (MD= -5.10, p=0.01). Thus, there is a significant increase in LF n.u. upon auditory stimulation when measured during the recovery condition. At stimulation and recovery conditions, the values of LF n.u. did not significantly differ from the resting condition. Figure 5.4.1 displays the covariate (BMI) adjusted estimated marginal means of the LF n.u. for this 2x3 mixed factorial ANCOVA between FU and FR groups. The regulation of response to sensory stimuli as represented by the pattern of change in the LF n.u. did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different physical environments (FU and FR).

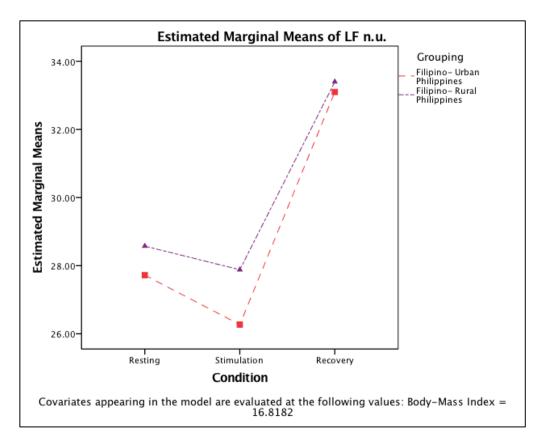


Figure 5.4.1 Covariate (BMI) adjusted estimated marginal means for LF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (FU and FR).

ii. HF n.u.

The Greenhouse-Geisser correction was employed due to a significant Mauchly's test (p<0.05) for a baseline-corrected 2x3 mixed factorial ANCOVA (using BMI as a covariate) in exploring changes in the HF n.u. measure. Changes in the mean HF n.u. did not significantly varied across conditions (F(1.86, 174.94)= 0.42, MS= 44.02, p= 0.65) within group. There was no significant interaction between condition and participant grouping (FU and FR) for the LF n.u. measure (F(186, 174.94)= 0.22, MS= 22.96, p= 0.79). Follow-up condition pairwise comparisons did suggest similar results where no significant differences in the change of HF n.u. values were likewise seen. Thus, HF n.u. seems to be maintained even after sensory stimulation. Figure 5.4.2 displays the covariate (BMI) adjusted estimated marginal means of the HF n.u. for this 2x3 mixed factorial ANCOVA between FU and FR groups. The regulation of response to sensory stimuli as represented by the pattern of change in the HF n.u. did not have significant interaction between condition and participant grouping among children

having similar ethnicities living in different physical environments (FU and FR).

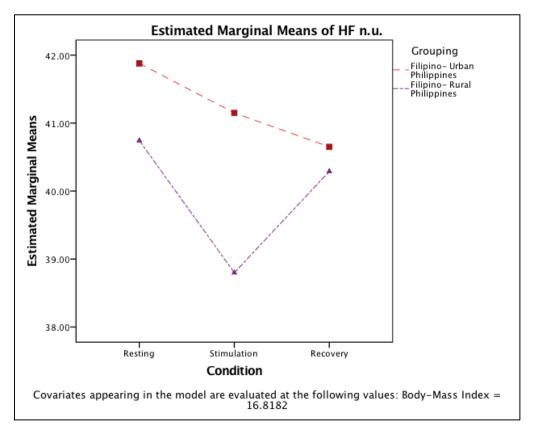


Figure 5.5.2. Covariate (BMI) adjusted estimated marginal means for HF n.u. using baseline-corrected 2x3 mixed factorial ANCOVA (FU and FR).

iii. SCR

A baseline-corrected 2x3 mixed factorial ANCOVA (using BMI as a covariate) was carried out for the physiological variable, SCR, and as Mauchly's test was significant (p<0.05), the Greenhouse-Geisser correction was applied. Mean changes in the SCR values significantly varied across conditions (F(1.59, 188)= 14.51, MS= 0.02, p<0.00) within groups. The results suggest that the mean SCR values within all participant groups (FU and FR) significantly varied across conditions. However, within groups interaction between experimental condition and participant grouping for the SCR measure did not reach significance thresholds (F(1.59, 188)= 0.19, MS= 0.00, p= 0.78). Condition pairwise comparisons suggest significant differences in the change of SR values in resting to stimulation condition (MD= -0.03, p<0.00), stimulation to recovery conditions (MD= 0.02, p=

0.03) and resting to recovery conditions (MD= -0.01, p<0.00). The results suggest an interesting difference in the regulation response where there is a significant increase of the SCR upon auditory stimulation from resting conditions; and consequent decrease of the same SCR parameter after said sensory stimulation as measured during the recovery condition. At recovery conditions, SCR values were significantly increased from resting conditions due to auditory stimulation. Figure 5.4.3 displays the covariate (BMI) adjusted estimated marginal means of the SCR for this 2x3 mixed factorial ANCOVA between FU and FR groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCR did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different physical environments (FU and FR).

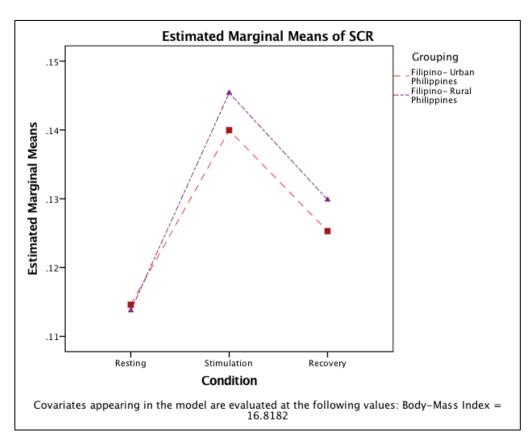


Figure 5.4.3. Covariate (BMI) adjusted estimated marginal means for SCR using baseline-corrected 2x3 mixed factorial ANCOVA (FU and FR).

iv. SCL

The Greenhouse-Geisser correction was employed due to a significant Mauchly's test (p< 0.05) for a baseline-corrected 2x3 mixed factorial ANCOVA (using BMI as a covariate) in exploring the changes in the SCL measure. Mean SCL values significantly varied across conditions (F(1.26,118.06)= 24.71, MS= 0.78, p<0.00) within groups. However, within groups interaction between condition and participant grouping for the SCR was not statistically significant (F(1.26, 118.06) = 0.19, MS = 0.02, p = 0.52). There was a significant difference in the SCL change from resting to stimulation (MD = -0.12, p < 0.00) and resting to recovery conditions (MD = -0.13, p < 0.00)p<0.00) after the follow-up condition pairwise comparisons. This suggests the significant increase of the SCL parameter after auditory stimulation from resting as a measure of sensory response and a continuous significant increase of the SCL when changes in the values are compared from resting and recovery conditions. Thus, at recovery conditions, there is a significant increase in SCL values from resting conditions after auditory stimulation. Figure 5.4.4 displays the covariate (BMI) adjusted estimated marginal means of the SCL for this 2x3 mixed factorial ANCOVA between FU and FR groups. The regulation of response to sensory stimuli as represented by the pattern of change in the SCL did not have significant interaction between condition and participant grouping among children having similar ethnicities living in different physical environments (FU and FR).

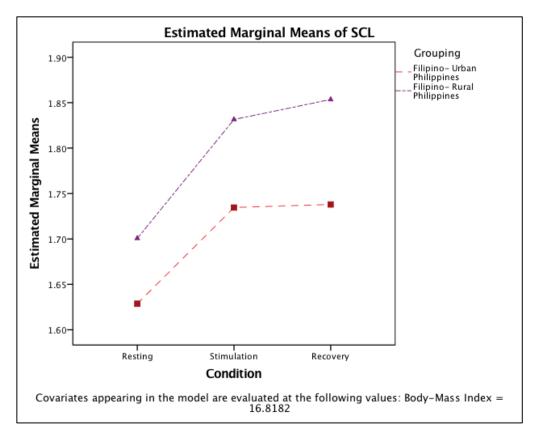


Figure 5.4.4. Covariate (BMI) adjusted estimated marginal means for SCL using baseline-corrected 2x3 mixed factorial ANCOVA (FU and FR).

Table 5.4.2. Summary of Differences in the Patterns of Neurophysiological Regulation Between FU and FR Groups Using Main Effects of Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

			Unco	rrected Ma	in Effect	ts			Baseline	Corrected	Main E	ffects	
	Outcomes	SS	df	MS	F	Sig	ES	SS	df	MS	F	Sig	ES
LF n.u.	Within												
	Condition	1074.90	2	537.45	5.55	<0.00	0.06	2153.20	2.00	1076.60	11.11	< 0.00	0.11
	Condition * BMI	21.13	2	10.57	0.11	0.90	0.00	145.73	2.00	72.86	0.75	0.47	0.01
	Condition * Grouping	740.56	2	370.28	3.82	0.02	0.04	18.44	2.00	9.22	0.10	0.91	0.00
	Error	18221.89	188	96.92				18221.89	188.00	96.92			
	Between												
	BMI	59.19	1	59.19	0.17	0.68	0.00	427.67	1.00	427.67	1.25	0.27	0.01
	Grouping	7621.29	1	7621.29	22.28	<0.00	0.19	54.14	1.00	54.14	0.16	0.69	0.00
	Error	32148.12	94	342.00				32148.12	94.00	342.00			
HF n.u.**	Within												
	Condition	539.18	1.86	289.71	2.73	0.07	0.03	81.93	1.86	44.02	0.42	0.65	0.00
	Condition * BMI	229.18	1.86	123.15	1.16	0.31	0.01	339.39	1.86	182.36	1.72	0.18	0.02
	Condition * Grouping	420.21	1.86	225.79	2.13	0.13	0.02	42.74	1.86	22.96	0.22	0.79	0.00
	Error	18547.53	174.94	106.02				18547.53	174.94	106.02			
	Between												
	BMI	172.89	1	172.89	0.27	0.61	0.00	833.94	1.00	833.94	1.28	0.26	0.01
	Grouping	16552.03	1	16552.03	25.47	<0.00	0.21	104.97	1.00	104.97	0.16	0.69	0.00
	Error	61080.68	94	649.79				61080.68	94.00	649.79			

SCR^a	Within												
	Condition	0.10	1.59	0.06	38.89	<0.00	0.29	0.04	1.59	0.02	14.51	<0.00	0.13
	Condition * BMI	0.01	1.59	0.01	3.32	0.05	0.03	0.00	1.59	0.00	1.14	0.31	0.01
	Condition * Grouping	0.01	1.59	0.01	3.85	0.03	0.04	0.00	1.59	0.00	0.19	0.78	0.00
	Error	0.25	149.54	0.00				0.25	188.00	0.00			
	Between												
	BMI	0.02	1	0.02	0.93	0.34	0.01	0.01	1.00	0.01	0.57	0.45	0.01
	Grouping	0.00	1	0.00	0.17	0.68	0.00	0.00	1.00	0.00	0.03	0.86	0.00
	Error	1.75	94	0.02				1.75	94.00	0.02			
SCL^a	Within												
	Condition	0.73	1.26	0.58	18.48	<0.00	0.16	0.98	1.26	0.78	24.71	<0.00	0.21
	Condition * BMI	0.17	1.26	0.13	4.22	0.03	0.04	0.13	1.26	0.11	3.37	0.06	0.03
	Condition * Grouping	0.02	1.26	0.01	0.41	0.57	0.00	0.02	1.26	0.02	0.51	0.52	0.01
	Error	3.73	118.06	0.03				3.73	118.06	0.03			
	Between												
	BMI	3.66	1	3.66	4.25	0.04	0.04	4.15	1.00	4.15	4.81	<0.00	0.05
	Grouping	1.22	1	1.22	1.41	0.24	0.01	0.57	1.00	0.57	0.67	0.42	0.01
	Error	80.96	94	0.86				80.96	94.00	0.86			

Note: LF n.u.= HRV- Low Frequency normalized unit; HF n.u.= HRV- High Frequency normalized unit; SCR- EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; SS= Sum of Squares; df= Degrees of Freedom; MS= Mean Squared; ES= effect size (Cohen's d); Sig= significant difference at p < 0.05; asignificant (p < 0.05) Mauchly's test

Table 5.4.3. Summary of Differences in the Patterns of Neurophysiological Regulation Between FU and FR Groups Using Condition Pairwise Baseline Corrected 2x3 Mixed Factorial ANCOVA by Neurophysiological Parameter.

	Baseline Corrected Condition Pairwise Comparisons															
	Rest-Stim						Stim-Recov					Rest-Recov				
		95% CI							95%	6 CI				95%	% CI	
Outcomes	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	MD	SE	Sig	LB	UB	
LF n.u.	1.07	1.42	1.00	-2.38	4.53	-6.17*	1.28	<0.00	-9.29	-3.06	-5.10*	1.56	<0.00	-8.91	-1.29	
HF n.u ^a	1.34	1.40	1.00	-2.07	4.74	-0.49	1.28	1.00	-3.62	2.63	0.84	1.61	1.00	-3.09	4.77	
SCR^a	-0.03*	0.01	<0.00	-0.04	-0.01	0.02*	0.01	0.03	0.00	0.03	-0.01*	0.00	<0.00	-0.02	0.00	
SCL^a	-0.12*	0.02	<0.00	-0.17	-0.07	-0.01	0.01	0.87	-0.04	0.02	-0.13*	0.03	<0.00	-0.20	-0.07	

Note: LF n.u.= HRV- Low frequency normalized unit; HF n.u.= HRV- High frequency normalized unit; SCR: EDA- Skin Conductance Response; SCL: EDA- Skin Conductance Level; MD= Mean Difference; SE= Standard Error; LB= Lower Bound limit; UB: Upper Bound limit; Sig= significant difference at p < 0.05; * significant at p < 0.05; asignificant (p < 0.05) Mauchly's test

D. Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between FU and FR Groups

i. Resting Condition

For the resting condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (FU: n=54, age= 8.91 yr., BMI= 17.91 kg/m²; FR: n=43, age= 9.39 yr., BMI= 18.41 kg/m²) by MANOVA, and BMI as a covariate (see Table 5.4.1).

There is significant multivariate group mean difference between the two groups (Λ = 0.19, F(3,11)= 7.29, p<0.00, d= 0.97) on LF n.u. (FU: M= 28.10; SD= 14.10; FR: M= 36.75; SD= 13.91), HF n.u. (FU: M= 57.91; SD= 17.72; FR: M= 41.37; SD= 17.48), and SCL (FU: M= 1.64; SD= 0.54; FR: M= 1.73; SD= 0.51). Table 5.4.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at resting condition among children having similar ethnicities living in different physical environments (CHK and FU).

ii. Stimulation Condition

For the stimulation condition, LF n.u., HF n.u. and SCR were compared between the two groups of participants (FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²; FR: n= 43, age= 9.39 yr., BMI= 18.41 kg/m²) by MANOVA, and BMI as a covariate (see Table 5.4.1).

There is significant multivariate group mean difference between the two groups (Λ = 0.30, F(3,11)= 13.09, p<0.00, d= 1.31) on LF n.u. (FU: M= 26.98; SD= 12.74; FR: M= 41.00; SD= 14.32), HF n.u. (FU: M= 55.93; SD= 17.87; FR: M= 40.11; SD= 16.75), and SCR (FU: M= 0.43; SD= 0.57; FR: M= 0.82; SD= 0.60). Table 5.4.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCR at stimulation

condition among children having similar ethnicities living in different physical environments (CHK and FU).

iii. Recovery Condition

For the recovery condition, LF n.u., HF n.u. and SCL were compared between the two groups of participants (FU: n= 54, age= 8.91 yr., BMI= 17.91 kg/m²; FR: n= 43, age= 9.39 yr., BMI= 18.41 kg/m²) by MANOVA, and BMI as a covariate (see Table 5.4.1).

The multivariate group mean difference between the two groups approached traditional significance thresholds (Λ = 0.14, F(3,11)= 5.00, p<0.00, d= 0.81) on LF n.u. (FU: M= 33.23; SD= 11.97; FR: M= 40.59; SD= 13.42), HF n.u. (FU: M= 52.62; SD= 15.34; FR: M= 40.49; SD= 15.65), and SCL (FU: M= 1.78; SD= 0.67; FR: M= 1.83; SD= 0.52). Table 5.4.4 shows the summary of the MANOVA tests and follow-up ANOVA tests. There is significant difference in the regulation of response to sensory stimuli as represented by the levels of autonomic activity represented by LF n.u., HF n.u. and SCL at recovery condition among children having similar ethnicities living in different physical environments (CHK and FU).

Table 5.4.4. Summary of Differences in the Neurophysiological Response to Sensory Stimuli Across Conditions Between FU and FR Groups MANOVA Tests (n= 97).

	FU		FR		Total		MANOVA				ANOVA		
	n=54		n=43		n=97		Pillai's V	F	n	d	F	n	d
	M	SD	M	SD	M	SD	Fillat S V	ľ	p		<i>I</i> '	p	u
Resting- LF n.u.	28.1	14.1	36.75	13.91	31.93	14.6					9.51	<0.00*	0.64
Resting- HF n.u	57.91	17.72	41.37	17.48	50.58	19.37	0.19	7.29	<0.00*	0.97	21.21	<0.00*	0.95
Resting- SCL	1.64	0.54	1.73	0.48	1.68	0.51					2.01	0.16	0.29
Stimulation- LF n.u.	26.98	12.74	41	14.32	33.2	15.11					29.12	<0.00*	1.11
Stimulation- HF n.u.	55.93	17.87	40.11	16.75	48.92	19.01	0.30	13.09	<0.00*	1.31	23.76	<0.00*	1.01
Stimulation-SCR	0.17	0.10	0.14	0.08	0.16	0.09					13.46	<0.00*	0.76
Recovery- LF n.u.	33.23	11.97	40.59	13.42	36.49	13.09					7.80	<0.00*	0.58
Recovery- HF n.u.	52.62	15.34	40.49	15.65	47.24	16.55	0.14	5.00	<0.00*	0.81	13.82	<0.00*	0.77
Recovery- SCL	1.78	0.67	1.83	0.52	1.80	0.61					1.31	0.26	0.24

Note: Pillai's V refers to MANOVA test statistics; F refers to the F-statistics; p refers to the significance level set at p < 0.05; d refers to the effect size expressed in Cohen's d;*significant after Bonferroni Adjusted p = 0.017

E. Summary of Hypothesis Testing 4

The influence of physical environments on the adaptation to sensory stimuli using an auditory stimulus was examined by looking at the neurophysiological regulation of the ANS using HRV and EDA. There was no significant interaction between condition and participant grouping (FU and FR) across neurophysiological measures of regulation of response to sensory stimuli. The results of this thesis suggest that physical environments may not be sufficient to influence the direction of change in the neurophysiological measures in response to sensory stimuli among children.

Using the HF n.u. and SCR as representative measures of PNS and SNS activity respectively, there are interesting patterns of change in the neurophysiological regulation of response to sensory stimuli can be seen exhibited by each group. The significant increase in SCR activity with the maintenance of the HF n.u. activity from resting to stimulation conditions seen in the FU and FR groups.

Indexing the change in the activity of the HF n.u. and SCR when auditory stimulation is removed, from stimulation to recovery conditions, suggests another set of interesting results. The FU groups showed a significant decrease in their SCR while the HF n.u. is maintained. In contrast, the FR group's data showed maintenance of HF n.u. and SCR activity at baseline with no significant pattern of change.

To determine the overall autonomic activity of the HF n.u. and SCR, this thesis looks at the patterns of change from resting to recovery conditions. The FU group showed baseline control of HF n.u. and SCR, which suggests that at recovery conditions, autonomic activity was the same at resting condition. On the other hand, the FR group displays a significant change in the HF n.u. with a significant increase of SCR seen at recovery conditions compared to the resting conditions.

This section compared the ANS activity across conditions between FU and FR groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis found differences between the two groups having similar ethnicities and geographic environment but living in different physical environments on the levels of autonomic activity, but not on the autonomic patterns

of change from one condition to another. The physical environment significantly influences the neurophysiological regulation of response to sensory stimuli in children.

Chapter 6: Discussion

This research aims to identify the role of ethnicity and environments in the regulation of response to sensory stimuli in children, from a neurophysiological perspective by answering the question: "Do ethnicity and environment influence the regulation of response to sensory stimuli in children" using neurophysiological methods. In this thesis, to represent a sensory stimulus, an auditory stimulus was used to elicit ANS responses measured by PNS and SNS activity. Using groupwise (CHK, FHK, FU and FR) and pairwise (CHK and FHK, FHK and FU, CHK and FU, FU and FR) combinations of participant groups, several hypotheses were tested that lead to answering the research question. Below, the results of this thesis are summarized and discussed. Parts of this section of this thesis have already been published in Gomez et al., 2017 and Gomez et al., 2018.

I. Summary of Findings

Hypothesis Testing 1. In hypothesis 1, this thesis compared the ANS activity across conditions between CHK and FHK groups. This thesis found significant differences between these two groups that have different ethnicities but living in the same geographic environments in their SCL patterns of change from one condition to another. Furthermore, this thesis found no significant differences between CHK and FHK groups on the level of ANS activity in the three experimental conditions.

Hypothesis Testing 2. In hypothesis 2, this thesis compared FHK and FU groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis finds differences between the two groups having similar ethnicities but living in different geographic environments on their levels of autonomic activity, but not on the autonomic patterns of change from one condition to another.

Hypothesis Testing 3. In hypothesis 3, this thesis compared CHK and FU groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis found differences between the two groups having different ethnicities and geographic environments but living in similar physical environments.

Hypothesis Testing 4. In hypothesis 4, this thesis compared FU and FR groups on their level of ANS activity in the three experimental conditions. There is no significant difference in autonomic patterns of change from one condition to another. However, this thesis found differences between the two groups having similar ethnicities and geographic environment but living in different physical environments on the levels of autonomic activity, but not on the autonomic patterns of change from one condition to another.

II. Synthesis of Results

This thesis aimed to compare the regulation of physiological responses among 4 groups of children representing differences in ethnicity and environments. This thesis found that in response to sensory stimuli: 1) there are differences in the patterns of change in the SCL among CHK and FHK groups; and there are differences in the levels of autonomic activity between: a) FHK and FU groups; b) CHK and FU groups; and FU and FR group.

III. Discussion Proper

This section provides a discussion of the major findings in this thesis across hypotheses testing. Section A discusses salient the results of Hypotheses 1 to 4. Section B provides a discussion of the novel findings of this thesis.

A. Discussion of Salient Findings for Hypotheses 1 to 4

A. Hypothesis 1: Ethnicity Influence on the Regulation of Physiological Response to Sensory Stimuli

In hypothesis 1, this thesis compared the physiological (HRV and EDA) regulation of response towards auditory stimulus between children from different ethnicities living within the same geographic and physical environments across conditions between CHK and FHK groups. This thesis found significant differences between these two groups that have different ethnicities but living in the same geographic environments in their SCL patterns of change from one condition to another.

Ethnicity has biological origins related to genetics that may have an influence on the ANS (Ali-Khan et al., 2011; Jorde & Wooding, 2004); and this thesis found it specifically in the SCL, which is an EDA parameter reflecting sympathetic functions (Boucsein, 2012). Traditionally the SCL represents the

tonic phase of EDA and is measured in the absence of stimulation (i.e. resting or recovery conditions), while the SCR is measured during stimulus presentation (i.e. stimulation condition). SCR is measured to be the difference in skin conductance from the baseline SCL (Boucsein, 2012; Cacioppo, Tassinary & Berntson, 2007). However, this is an oversimplification of skin conductance activity. In this thesis, a different method of extracting the skin conductance drivers was used that enables continuous indexing both SCR and SCL across resting, stimulation and recovery conditions. Benedek and Kaernbach (2010) developed the CDA (continuous decomposition analysis) method which is able to retrieve the underlying sudomotor nerve activity responsible for the SCR and SCL. Decomposing EDA data thus produces separate but continuous tonic (SCL) and phasic (SCR) components. There is limited introspection in using this novel method. However, there is a theoretical reason to believe that both the SCR and SCL act in accordance to the sympathetic response (Benedek & Kaernbach, 2010; 2010). In this research, the changes in the SCL across conditions was found to be significantly different between the CHK and FHK groups. Furthermore, an analysis of the patterns of SCL changes across the sour groups reveals reciprocal response compared to the SCR. For example, when the child is presented with the auditory stimuli, sympathetic activity is initiated, and skin conductance is increased which can be seen in the magnitude of SCR. In contrast, the SCL is seen to significantly withdraw resulting in significantly lower values for all four groups. The reverse scenario is seen when the stimulus is removed. Previous studies found that genetic predispositions characterize individuals' regulation of sympathetic responses toward challenges (Bortoluzzi et al., 2015; McEwen, 2016; Pagliaccio et al., 2015; Roberts et al., 2015). It is possible that sympathetic differences in genetic origins have its specific patterns of response to external challenges, which may explain these findings.

The exact regulatory mechanisms that underlie SCL activity is yet to be fully understood. This thesis presents emerging evidence that may support the utility of the SCL as an important marker that may reflect the flexibility within the SNS. Furthermore, the importance of the SCL should be examined in relation to within SNS and between PNS regulation.

Furthermore, this thesis found no significant differences between CHK and FHK groups on the level of ANS activity in the three experimental conditions. This study's findings that ethnicity alone may not be able to influence the regulation of response to sensory stimuli replicates previous findings of Royeen and Mu (2003) which suggested that such ability is stable and seems to be universal. That is, regardless of ethnic origins, how a child responds to sensory stimuli would be similar across representative groups. However, the previous study primarily used behavioural measures among Caucasian children living in different geographic environments, which may make a comparison between this study's neurophysiological results dissimilar. Nevertheless, Chang et al. (2012) offered some supporting perspectives to the results of this research. In their study, the author suggested that Asian and American children living in the same geographic environments do not significantly differ in their autonomic activity in response to sensory stimuli. Differences between this research and Chang's group (2012) study exist and are worthy to be mentioned. Findings of the former group differed in their use of only skin conductance as their measure indexed during the SCP and lacked a further description of the specific ethnic origins of the Asian children sampled. In this research, a multiphysiological approach was adapted where autonomic measures of LF n.u., HF n.u. and EDA-SCL/SCR was indexed during a sensory paradigm using an auditory stimulus between Asian children of Chinese and Filipino ethnic origins.

The similarity in the level of ANS activity across conditions between the CHK and FHK group could be because stimuli are environment-dependent (Andreassi, 2013). According to the concept of allostasis, the exposure to similar stimuli, for example living in the same geographic environments, may shape parasympathetic and sympathetic responses, in order to adapt to challenges in a specific geographic environment (Danese & McEwen, 2012; McEwen 1998, 2000). The repeated exposure to an environment may shape people with different ethnicities to a similar set level of ANS activity in order to match the external demands, such as seen in the findings of this thesis (Danese & McEwen, 2012; McEwen, 2003, 2008).

While ethnicity may be a key variable that can profoundly modify the maturation of the neurophysiological systems responsible for adaptation to

external challenges (Rutter & Krepner, 2007), it remains unclear whether such differences are influenced by inherent ethnicity or the environment in context. Thus, the third hypothesis testing in this research inquires further on the influence of the geographic environments when ethnicity is held constant.

This research section concludes that there are significant differences between these two (CHK and FHK) groups that have different ethnicities but living in the same geographic environments in their SCL patterns of change from one condition to another.

B. Hypothesis 2: Geographic Environments Influence on the Regulation of Physiological Response to Sensory Stimuli

In hypothesis 2, this thesis compared the physiological (HRV and EDA) regulation of response towards auditory stimulus between children from similar ethnicities and physical environments living in different geographic environments between FHK and FU groups in the three experimental conditions. This thesis finds differences between the two groups having similar ethnicities but living in different geographic environments on their levels of autonomic activity.

This finding is similar to a previous behavioural study which found that responses to sensory stimuli differed among children living in distinct geographic environments (Caron et al., 2012; Gunn et al., 2009). However, it must be noted that while Gunn's (2009) group suggested the influence of geographic environments on the regulation of response to sensory stimuli, the study used primarily behavioural measures, which may complicate corroboration with the current research findings. Furthermore, it must be noted that there was limited literature that examined similar constructs as with the aims of this research. The available evidence discussed earlier in Chapter 2 of this thesis looked at the interacting effects of ethnicity and environmental variables (Caron et al., 2012). While this research shares similar aims at understanding the role of the environment on the autonomic activity in response to sensory stimuli, ethnicity of the children was controlled.

Since the concept of allostasis proposes that physiological functions such as the PNS and SNS underlie behavioural responses (McEwen, 2003, 2008), hence

similar differences are noted in this thesis. It may be that long-term dwelling in specific geographic environments led to the change in ANS activity, providing a possible explanation for the differences found in the level of ANS activity among the FHK and FU groups. The collective experiences within the child's environment form a life history that shapes our abilities to make sense of situations and instilled in us a set of codes that determines how we react or appraise a situation (Mims & Olden, 2012; Templer, 2008; Wolf et al., 2007). Thus, regardless of genetic predispositions, these life experiences may override such effects and allow individuals to adapt to their current environmental habitat (McEwen, 1999, 2008). The findings in this research suggesting an environment-dependent stimuli processing is supported by the concept of allostasis which explains why even with similarities in ethnic origins, children will eventually adapt their neurophysiological response to match the geographic environment where they live.

The specific dimension within a child's geographic environment that ultimately accounts for these differences remain unclear. Behaviours, beliefs, culture, parenting, physical landscapes, practices, socio-cultural and economic aspects have been previously suggested to explain these differences. Looking at the results of the study using Super and Harkness's (2002) developmental niche framework is one way to consider their findings. It is suggested in this framework that organization within the child's environment, culture is a factor that affects the course of the development of the child. Three operational subsystems comprise a child's developmental niche: (1) social and physical contexts, (2) historically ingrained behaviours (practices and customs) related to childcare and rearing, and (3) psychological makeup of the caregiver. A specific cultural niche is created from the interaction of these subsystems that organizes the daily environment of a child, thus possibly influencing the development of the ability to regulate responses to sensory stimuli.

Moreover, one difference noted in the FHK and FU groups was their sympathetic functions when responding to the challenge of auditory stimuli. The repeated activation of the SNS may have re-established a new baseline and magnitude of responsivity in order to adapt to the challenges in their distinct environments (McEwen & Wingfield, 2003). This is the possible reason why

there is a higher effect size in the group comparison of LF n.u. levels in the stimulation condition between the FHK and FU groups.

This research section concludes that there are significant differences between the two groups (FHK and FU) having similar ethnicities but living in different geographic environments on their levels of autonomic activity.

C. Hypothesis 3: Ethnicity and Geographic Environments Influence the Regulation of Physiological Response to Sensory Stimuli

In hypothesis 3, this thesis compared the physiological (HRV and EDA) regulation of response towards auditory stimulus between children from different ethnicities and geographic environments living in similar physical environments between CHK and FU groups in the three experimental conditions. This thesis found differences between the two groups having different ethnicities and geographic environments but living in similar physical environments on the levels of autonomic activity.

The previous hypotheses testing tried to isolate each of the independent variables by controlling in the pairwise combination; however, hypothesis 3 factors the interacting relationship between the two. This study's finding that environment and ethnicity may be able to influence the autonomic activity in response to sensory stimuli in children replicates previous findings of Caron et al. (2012) where sensory responses were found to be different between American and Israeli children when measured using behavioural checklists within the context of their own geographic environments. Limited evidence was likewise noted on research inquiries within the similar nature of the investigation. Nevertheless, methodological differences exist between this study and that of the previous one. The use of primarily behavioural measure is the most obvious. Behavioural measures may not be as objective, and the information may be confounded by parental factors (Hoyle, Harris & Judd, 2002; Ooi et al., 2016; Ozsivadjian, Knott & Magiati, 2012). Furthermore, the behavioural measure (i.e. Sensory Profile) used in Caron's (2012) study, has been challenged in its ability to reflect physiological symptoms related to the regulation of responses to sensory stimuli (Tirosh et al., 2003). Nevertheless, while behavioural measures have utility in measuring how daily activities are

affected by sensory symptoms, the latent constructs of the questionnaires used may not reflect a neurophysiological perspective related to autonomic functions. This study, on the other hand, employed neurophysiological methods which may have the ability to deeper insight into the mechanisms underlying the processing of sensory information and serve as a good option in research and clinical inquiries that aim to understand the regulation of responses to sensory information using laboratory paradigm experiments.

Specific to this subsection, the highest effect size was found in the LF n.u. at stimulation condition between the CHK and FU conditions. The LF n.u. is a measure of sympathetic activity. Responding to external challenges usually involves activation of the SNS. This physiological response has been suggested previously (Berntson et al., 1991; Berntson et al., 1994; Dawson & Schell, 2012; Dawson, Schell & Filion, 2007). The body is designed to protect itself. The SNS allows responding to the external threat to the body's homeostasis in various forms of autonomic reaction.

The normalised unit of the LF (LF n.u.) is an index of HRV. HRV refers to temporal fluctuations in between consecutive heartbeats (Berntson et al., 1997) that has been implicated in its ability to reflect cardiac autonomic activity (Cornforth, Jelinek & Tarvainen, 2015; von Borell et al. 2007). Analysing the fluctuations in the frequency domain, additional insight into the nature of HR fluctuations may be gained. Two major frequency domain measures have been suggested: the HF and LF. The HF has been consistently found to reflect PNS functions (Colombo et al., 2014, Malliani, et al., 1991; Montano et al., 2009; Pagani et al., 1986, Reyes del Paso, et al., 2013; Toninelli et al., 2012). However, the appropriateness of the LF in representing sympathetic functions has long been debated (Berntson et al., 1997; Hakim, Gozal & Gozal, 2012; Japundzic, Grichois, Zitoun, Laude, & Elghozi, 1990; Reyes del Paso et al., 2013; Randall, Brown, Raisch, Yingling, & Randall, 1991). While the results seem to suggest the influence of ethnicity and environment on the regulation of the SNS in response to sensory stimuli, this must be approached with caution. Nevertheless, while the exact representativeness of the LF measures (whether sympathetic or vagal), newer evidence suggests that the LF may represent a significant sympathetic function albeit not as a single measure to represent the

SNS (Thayer, Ahs, Frederikson, Sollers & Wager, 2012; Reyes del Paso, Langewitz, Mulder, Van Roon & Duschek, 2013).

Individual differences shape the adaptive abilities of children in responding to external challenges. The concept of allostasis (McEwen, 2003, 2008) is helpful in interpreting the results of this study by illustrating how individual differences related to environment-dependent stimuli processing and biological embedding in ethnicity influences the physiological response by way of autonomic activity in regulation mechanisms (Danese & McEwen, 2012). Furthermore, perturbations within multiphysiological mediators of allostasis enable this physiological response (McEwen, 1999, 2000; McEwen & Wingfield, 2003). In this study, meaningful results supporting the hypothesis were answered approaching such multiphysiological systems compared to individual indices seen in previous researches. Overall, this research demonstrates a meaningful and effective utilization of the concept of allostasis as applied to the inquiry on the regulation of response to children. The findings of this research further support the concept of allostasis as a versatile theoretical framework which can be applied in other areas of research.

The exemplified ethnicity and environment effects supported by this study reflect similar concepts offered by gene x environment interactions. Previously, it has been suggested that the interaction between ethnicity and environment has a moderating role in the activity of a child's autonomic activity that is further related to their ability to adapt to external stressors (Dieleman et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012). The regulation of such autonomic activity has likewise been proposed to be an underlying mechanism that supports childhood behaviour. The gene x environment role in autonomic activity influencing child behaviours has previously been suggested. The interaction between genetic and environmental variables influence the state of autonomic activity that in turn supports children's behaviour, specifically in the areas of stress regulation (Bortoluzzi et al., 2015; Kuhlman et al., 2015; Marsman et al., 2012; Roberts et al., 2015), emotion regulation (Dieleman et al., 2015), academic (Pagliaccio et al., 2015b) and social behaviours (Pagliaccio et al., 2015a). From the foregoing, it seems that environment experiencedependent information may mediate biologically-embedded genetic information

and influence its epigenetic expressions, measured from both behavioural and physiological perspectives. Such gene x environment interaction may be a stronger model to explain its role in regulating the autonomic states in children that may support underlying psychophysiological mechanisms of behaviours. The results of this study further expound the body knowledge that the interaction between genome and environment might have a stronger role in the regulation of autonomic activity that could support a psychophysiological mechanism that supports child behaviours. Nevertheless, experiences with sensory information in one physical environment could be different from another. The succeeding section, hypothesis testing 4, specifically looks at whether variations in the physical living environments may produce similar differences in the autonomic activity in response to sensory stimuli in children.

This research section concludes that there are significant differences between the two groups (CHK and FU) having different ethnicities and geographic environments but living in similar physical environments on the levels of autonomic activity.

 D. Hypothesis 4: Physical Environments Influence the Regulation of Physiological Response to Sensory Stimuli

In hypothesis 4, this thesis compared the physiological (HRV and EDA) regulation of response towards auditory stimulus between children from similar ethnicities and geographic environments living in different physical environments between FU and FR groups in the three experimental conditions. This thesis found differences between the two groups having similar ethnicities and geographic environment but living in different physical environments on the levels of autonomic activity.

This finding is similar to previous studies that found differences in the behavioural responses to sensory stimuli (Lin et al., 2004; Tirosh et al., 2003) and physiological activity among urban and rural-dwelling individuals (Laumann et al., 2003; Ulrich et al., 1999). The result of this study which supports variations in the regulation of response to sensory stimuli among a group of children living in different physical environments parallel the results of previous researches. For example, this study shares a similar proposal that urban

and rural children have different performances in the response to sensory stimuli such as seen among Taiwanese children Lin (2013). Tirosh et al. (2003) similarly found the same urban-rural differences and specifically suggested socio-cultural influences (i.e. cultural differences, maternal education) can moderate the regulation of response to sensory stimuli among children living in rural settings. However, findings from previous research showed varying methodologies in mainly adopting a behavioural perspective. The use of behavioural outcomes, such as parent/caregiver reports, is limited in its ability to provide objective and precise data related to the regulation of response towards sensory stimuli (Schoen et al., 2009). In this research, the results are supported by objective neurophysiological physiological outcomes indexing the regulation of PNS and SNS autonomic measures in response to sensory stimuli among children.

The influence of the physical environment was explored in this study by conceptualizing it as the objective characteristics of the physical context related to habituation and gradients of man-made or natural structures and components (McDonnell & Pickettt, 2000; Yung et al., 2017). Specifically, the dichotomous characteristics of urban and rural settings, further operationalized by Perloff (2015) were employed. The physical attribute of the living environment (i.e. noise level, pollution, infrastructures, topography, space, etc.), has been previously suggested to contribute to possible differences in the regulation of response to sensory stimuli. In comparison, urban environments offer more artificial sensory stimuli compared to nature rural settings. The sensory environment and the opportunities in the environment generally affect the brain and its functions (Kempermann, van Praag and Gage, 1999) that will be responsible for the responses to the stimuli in the environment. Constant exposure to such stimuli builds adaptation to these external challenges that set the physiologic responses. Thus, children growing up in different physical environments may have different capacities, reactivity and adaptability abilities in the face of stimulation. However, there is limited information available in the literature that tries to explore the role of the physical contexts of environments, specifically between different contexts such as urban and rural settings, on the physiological response behaviours of children to environmental challenges such as different sensory stimuli. This research offers novel findings and methods in exploring the influences of the dichotomous physical environmental exposures and its effects on the physiological responses of children.

Perhaps, features in the physical environment require different levels of autonomic activity in order to cope with the challenges in an urban or rural setting (Laumann et al., 2003; McEwen, 1998, 2003; van den Berg et al., 2015). For example, the FR group lived in mountainous regions, and previous studies found an increase in SNS functions among people in this kind of environment (Hainsworth et al., 2007).

In fact, the findings in this thesis are consistent with previous studies that found exposure to nature stimuli (i.e. vegetation) exhibit lower level of PNS activity in response to stressful challenges (Laumann et al., 2003; van den Berg et al., 2015). We can consider that in order to adapt to the challenges effectively and efficiently the mode of autonomic control, the readiness of the ANS, and the reactivity of the ANS play an important role (Berntson et al., 1991). For example, this thesis found the largest effect size in the comparison of the levels of LF n.u. and HF n.u. in the stimulation condition, which accounts for the reactivity towards stimulation. On the other hand, the effect size was found to be larger in the readiness or capacity to respond for both LF n.u. and HF n.u. levels because of the mode of autonomic control required to respond to the challenge of an auditory stimuli. This is further supported by the concept of allostasis which suggests that physiological responses mediated by the ANS are "set" according to the specific environments, explaining the differences found in this thesis (Danese & McEwen, 2012; McEwen, 2003, 2008).

This research section concludes that there are significant differences between the two groups (FU and FR) having similar ethnicities and geographic environment but living in different physical environments on the levels of autonomic activity.

B. Novel Finding in this Thesis: Migration Influences the Regulation of Physiological Response to Sensory Stimuli

While the main aim of this research is to elucidate findings that support the influence of children's ethnicity and environment on their ability to regulate responses to an

auditory stimuli, an interesting finding emerges. This deals with the phenomenon of migration. To recap, the results of hypothesis testing 3 of this research establishes that children from different ethnicities living in different geographic environments (but similar physical characteristics) have different regulation of response to sensory stimuli. Hypotheses testing 1 and 2 recruited a specific type of group of children that represents migrant children; these are children who are growing up in a geographic environment apart from where the biological life history originated. This research finds that their regulation of response to sensory stimuli is currently similar to those of their peers from their current host geographic environment (hypothesis testing 1) and significantly different from their peers from their country of origin. Moreover, when the effects of migration are accounted, this research finds that it significantly contributes to the variances in the regulation of response to response to sensory stimuli. Thus, migration needs to be further discussed.

Migration as a topic presents diversified points of view. In this thesis, migration is approached from a biological perspective. Migration is characterised with a movement (individual or a group) from one place to another, establishing a new dwelling (Dingle & Drake, 2007; Goldstein & Goldstein, 1981). Migration involves a change in the living environments from a place of origin to a new host environment.

The concept of allostasis can be used as a concept that unifies the underlying mechanism of adaptation related to migration and its probable effects on children's health and well-being. Supported by the earlier works of McEwen (1998, 2000), Sterling and Eyer (1988), and McEwen and Wingfield (2003), allostasis suggests that in order to adapt to external challenges there are consequent alterations in regulatory parameters resulting in new basal set points enabling optimum performance (Ganzel & Morris, 2011; Juster et al., 2011). Allostasis allows homeostasis, survival and short-term adaptation. However, it can likewise be involved in permanent changes after prolonged environmental exposures subsequently referred to as allostatic load (McEwen & Wingfield, 2003). Allostasis has been widely used to explain the adaptation of migrating animals (i.e. birds, fishes) to their new host environment measured using physiological parameters related to neurophysiologic mechanisms (Legagneux et al., 2013; Nardocci et al., 2014). Findings are consistently suggesting there is regulation of physiological parameters linked to survival in the host environment. In human migration studies, measures of allostatic load represent

multiphysiological system regulation (i.e. body composition, cardiovascular, metabolic) using several biomarkers (Doamekpor & Dinwiddie, 2015; Duong et al., 2017).

In the model of allostasis, environmentally contextualized interaction between individual differences, behavioural responses, and physiological responses suggests supporting mechanisms responsible for the adaptation of vulnerability among individuals (McEwen, 2008). The adaptive interaction among these components may represent the overall health and well-being of a child. Individual difference may include salient characteristics related to age, gender, environment, ethnicity, family dynamics, genetic predispositions, birthplace, site of migration, socio-cultural influences, socioeconomic status; all of which have been suggested to influence the allostatic load (Berry, 1997; McEwen, 2004). The function of the environment is underscored in its ability to shape behavioural and physiological responses. The physical environments (i.e. humidity, noise, physical landscapes and features, temperature, seasonality) succinctly account for physiological outcomes, which needs to be accounted for (Berntson et al., 1997; Boucsein, 2012; Roth, Dawson & Filion, 2012). Furthermore. there is promising evidence suggesting that one's nativity (birthplace) can affect the allostatic load (Salazar et al., 2016). The foregoing factors are similar to those suggested by Berry's (1997) classical paper on factors that can support migrants' adaptation. Nevertheless, the influence of these factors has been centred mainly on psychological adaptation to a new culture, notwithstanding relevant biological adaptation (i.e. physiology) associated with migration. Indeed, applying the concept of allostasis can provide a novel approach to understanding the effects of migration on the behavioural and physiological regulation of children. Moreover, it proposes a possible mechanism that can support neurophysiological conceptualizations of adaptation (or maladaptation) in children of migrant origins that can inform the provision of relevant and responsive health programs. Thus, this research seemingly fills in the need for providing insights on how migration influences children's regulation of physiological responses.

While findings in this research suggest that ethnicity can influence the development of behavioural and autonomic responses, there is stronger evidence implying the role of environments. The external environment which can comprise of socio-cultural and physical landscape settings can have the ability to shape genetic phenotypes responsible for consequent behavioural and autonomic responses. This theory of environmental

epigenetics seems to support the preliminary findings of this research (Cortessis et al., 2012; Ho, 2012). Epigenetics endorse the notions that DNA expression can be modified without alterations in the genetic code itself (Gräff & Mansuy, 2008). Inherited DNA from parents is embedded with information that directs how the body functions and influences its overall health state. This information is in the form of genes, and during gene expression, the DNA code is translated into proteins that carry out specific cell activities responsible for specific physiologic processes and mechanisms. The field of behavioural epigenetics has shown how experiences and environments can impact genetic expressions producing individual differences related to one's mental health (Mill & Petronis, 2008; Stuffrein-Roberts, Joyce & Kennedy, 2008), personality (Bagot & Meaney, 2010), cognition, (Powledge, 2011) and behaviour (Zhang & Meaney, 2010). Thus, while children can have genetic predispositions reflected in the behavioural and autonomic responses, some aspects and parameters of such can be modified through constant environmental exposures related to socio-cultural and physical landscape contexts and experiences. In this research, these modifications are seen as adaptive autonomic responses to adapt and maintain homeostasis in the face of external sensory challenges.

The life history theory is used herein to explain how organisms, in variable environmental contexts, adjust their developmental trajectories (Mims & Olden, 2012; Templer, 2008; Wolf et al., 2007). Such that, children develop bodily maintenance capacities that are in tune with the environment from which their life history is based on. This might be reflected in genetic predispositions that are manifested in ethnicbased behavioural responses. However, organisms have the capacity to adapt to novel external challenges of the environment through alteration of regulatory parameters (Ganzel & Morris, 2011; Juster et al., 2011). This is revealed through the continuous reevaluation and readjustments of allostatic mediators (PNS and SNS) that create new set points that maximize the organism's resources for adaptation (Danese & McEwen, 2012; Korte et al., 2005; Juster et al., 2011). Animal studies have likewise presented evidence of how an organism's measures of allostatic load are re-set in response to a new environment (i.e. migration) as part of their survival (Fryxell & Sinclair, 1988; Jachowski & Singh, 2015; Nathan et al., 2008). Animal allostatic responses were measured using behavioural and autonomic indices. It has been suggested earlier that long-term exposure to environmental demands and the concurrent allostatic responses

can shape the behavioural and autonomic responses of children (McEwen, 1998, 2000; McEwen & Wingfield, 2003; Sterling & Eyer, 1988). This research provides preliminary evidence to support the same mechanism in humans, specifically in the FHK group of children, suggesting that physiologic responses are re-shaped by their environments to similar reference points with their counterparts.

IV. Limitations of the Study

Although this research was carefully and thoughtfully planned and had reached its aims, there were some unavoidable limitations. This subsection discusses the limitations of the study methods and data analysis related to this thesis. For the purpose of organisation of presentation, this section is further subdivided into three areas: study design limitations, impact limitation, and statistical/data limitations.

A. Sample Size

While this research was able to recruit the necessary minimum number of samples for each group based on our previously published manuscript, there was an uneven sample size between the pairwise combinations of the groups. Though remediation was done statistically (i.e. use of Pillai's test, bootstrap method), this unevenness may likely have skewed some aspects of the data. Nevertheless, the researcher deemed it more meaningful to keep and use the data of the bigger group, rather than throw away the data to attain equality of sample. Moreover, despite efforts to control for theory-driven known demographic variables (i.e. age, gender, BMI, migration) that might have influenced the results, several variables related to individual differences among the participants were not regulated. These variables were carefully considered as covariates statistically, following the statistical assumptions testing and diagnostics. When appropriate, they have likewise been factored as covariates in the general linear model. However, heterogeneity in the other demographic variables (i.e. school type, no. of parents working, primary caregiver, educational attainment of caregiver, income classification, socioeconomic classification) may have likewise influenced the results. To account for the contributions in the variance, the entire set of demographic variables were statistically tested by way of regression and subsequently reported at the end of each study results. Nevertheless, this research suggests future efforts to control

further for these individual differences at the level of participant recruitment to lessen their effects on the overall data.

B. Representativeness of Samples

In this research, to represent differences in ethnicity, typically developing children from Chinese and Filipino ethnic origins were recruited. Geographic environments follow similar choice, specifically recruiting participants in Hong Kong and the Philippines. The physical environment was differentiated using groups coming from urban and rural settings in the Philippines. While the results of this research support its aims and hypothesis, the limited acumen in the representativeness of the sampled groups, their subsequent characteristics, regional focus, and population specificity may have limited the impact of the findings' generalisability. There will be a need to have future research to replicate the findings of this research as applied to different population group characteristics.

C. Measure of Change in the Environment

In this thesis, children from the FHK group represents a population which involved in a change of environment. Nevertheless, the actual change was not captured. In future studies, exploration on capturing events prior, during and several time points after the change in geographic environments using similar neurophysiological methods is likewise recommended.

D. Statistical/Data Limitations

In the analysis of the data coming from multiphysiological system, this research used separate physiological instruments with their own data acquisition systems. Data processing needed aligning epochs to specifically represent the conditions in the laboratory paradigm of this research. While this research has carefully followed the standards and guidelines in the processing and analysis of physiological data, there might have been some unavoidable human error on the realignment of these separate data. Although the nature of the research calls for a block-design (Gomez et al., 2018) and negligible errors may ensue, it cannot be underestimated that these could be further lessened using high-grade physiological laboratory equipment systems. Certain factors related to financial resource constraints and mobility needs (the research must be carried in several locations) restricted this research. It would

be interesting if future research can use such high-grade physiological laboratory equipment systems to replicate the findings of this research and examine whether similar results are gathered.

V. Future Research and Recommendations

This subsection addresses the limitations of this research by providing recommendations to improve future research related to the inquiry on the regulation of response to sensory stimuli in children

The published preliminary findings of this research recommended a sample size of n=28 to reach moderate effect sizes, has been realised in this research. However, there were some issues on the equality of sample sizes. It is further suggested by this research to recruit a similar number of participants in each group to avoid skewing of data or statistical remediation.

This research has demonstrated that individual differences (related to ethnicity and environment of children) have an influence on the autonomic activity in response to sensory stimuli in children. Thus, it is endorsed that other demographic data be robustly controlled to minimize their contributions to variance. This research suggests that future research apply multilevel cluster sampling procedures, considering sample demographic characteristics factored in the recruitment of participants. Age, gender, BMI, migration status, school type, no. of parents working, primary caregiver, educational attainment of caregiver, income classification, socio-economic classification needs to be succinctly controlled to improve similarity of the sample characteristics at baseline.

The behavioural outcome to measure behavioural responses related to the regulation of responses to sensory stimuli used in this research may not be from the same perspective from which the research questions was conceived. Thus, construct differences may have likely occurred. It is suggested by the researcher to use similar behavioural measures that specifically address physiologic symptoms related to autonomic functions reflected in daily life activities. It may be interesting to use sensory behaviour questionnaires reflecting underlying neurophysiologic functions of the ANS such as the one conceptualised in the Sensory Processing and Self-Regulation Checklist (Lai & Chiu, 2013). SPSRC is a parent/caregiver checklist that incorporates aspects of self-regulation from a neurophysiological perspective to provide a clearer picture of how children regulate sensory responses. SPSRC is a reliable and valid parent-reported single checklist that can

provide a summary of a child's sensory processing and self-regulation performance in daily life activities (Lai & Chiu, 2013).

While this research utilised physiological outcomes that conform to industry and research standards and guidelines for measurement, it is further suggested by the researcher that future research consider high-end multiphysiological systems in the data acquisition and data processing. The robustness of such systems eliminates unavoidable human-error factors that may confound the results and addresses related to the acquisition and processing of physiological data. Furthermore, it must be considered that future physiological instrument should likewise be mobile so that it can be transported to several locations where specific participants can be tested.

This research was able to meet its aim and prove the hypothesis related to examining the influence of ethnicity and environment on the regulation of response to sensory stimuli in children. However, the impact of the results may be limited to the specific types of children's ethnicity and environment recruited for this study. Thus, it is recommended that future researchers replicate the findings in other ethnicities, geographic and physical environments so that generalizability of the findings is strengthened and proven. Research in the future may consider recruiting participant from different racial profiles (i.e. Asian, Caucasian, African), ethnicities within and across continents (i.e. Asia, North America, South America, Africa, Europe, etc.), and physical environments (i.e. regional, citywide, provincial, nationwide).

Lastly, one of the novel and interesting findings of this research reported on the influence of migration on the regulation of response to sensory stimuli in children. While findings of this research suggest migration can account for variances in the ability to regulate responses to sensory stimuli children, further inquiry on the topic is advised. Controlling or differentiation for migration status (i.e. native-born, foreign-born), years of migration and even longitudinal follow-ups may provide fascinating prospects. Furthermore, other than following-up on the development of the ability to regulate response to sensory stimuli in children, its correlates with health well-being should likewise be noted. Doing so provides more depth and clinical relevance on this topic.

VI. Implications of this Study

This research has reached its aim in supporting the hypothesis that ethnicity and environment can influence how children regulate responses to sensory stimuli. Considering

the findings in this thesis, this section outlines the following significant implications that can inform both research and practice.

Previous studies have suggested how a child's ethnicity and environment may influence behavioural responses to sensory stimuli. This thesis finds the influence of ethnicity and environment on the regulation of response to sensory stimuli among children. Given the findings in this thesis, there are several implications.

First is on understanding how a child's ethnic background and environment can influence physiological functions that may support health and participation behaviours. The idea that such individual differences can impact the response to sensory stimuli will provide a salient rationale for considering these factors in assessing children from different backgrounds.

Second, is on the use of client-specific assessment and intervention programs tailor-fitted to individuals within a specific context (i.e. country, urban, rural) that may affect a child's health and participation behaviours. Careful consideration should be integrated into intervention planning for an ethnic-environment diverse clinical population.

Third, the use of physiological function-based measures, evaluation tools and interventions in the measurement of sensory responses. The findings of this research have implications on the opportunity to use objective neurophysiological measures in adjunct to conventional behavioural measures to better help in assessing client needs and outcome performance. While many performance measurements used by clinicians in the clinics considers cultural sensitivity and specificity, objective neurophysiological measures such as demonstrated in this research (i.e. HRV and EDA) that has been suggested to support behaviours and performance, can provide a more in-depth picture of the clients' health and performance. While existing behavioural measures might have psychometric evidence on its utility, the findings of this research challenge existing items on such by exploring the inclusion of specific neurophysiological functions in the context of activity performance and occupation.

Fourth, this thesis informs the development of interventions(i.e. mindfulness) related to adaptation to external challenges (i.e. stress) specific to migrant (i.e. domestic workers, migrant workers and their families, foreign students) ethnic groups. Since evidence suggest that the ANS is the first neurophysiologic system to respond to external environmental stimuli, intervention techniques aimed at preparing, enhancing or managing the body to be optimally available to respond could be considered (i.e. biofeedback, Neurofeedback,

stress-management and relaxation techniques, music therapy, etc.) in adjunct to conventional interventions. Considering the ethnic background will provide tailor-fitted programs responsive to the innate physiological responses specific to the target ethnic group.

Lastly, is on further understanding the influence of migration on the physiological functions of individuals. this research has an implication on understanding the effects of migration on children's adaptation. In the case of migrant children whose regulation of response to sensory stimuli has been found to have adapted to their current host environment, it is still unclear whether such adaptation is beneficial to their health, or certain trade-offs may ensue in the future. Therefore, this research has further implications in suggesting future research to examine the health and well-being of migrant children in all their variants (i.e. native-born, foreign-born, internal migrants, forced migrants, etc.).

Chapter 7: Conclusion

This research aims to identify the role of ethnicity and environments in the regulation of response to sensory stimuli in children, from a neurophysiological perspective by answering the question: "Do ethnicity and environment influence the regulation of response to sensory stimuli in children" using neurophysiological methods. In this thesis, to represent a sensory stimulus, an auditory stimulus was used to elicit ANS responses measured by PNS and SNS activity. This last chapter summarises the findings of the different studies of this thesis to answer the research questions. Lastly, the future work directions of the author are explored.

I. Summary of Conclusions

This thesis proposed to apply allostasis as a concept to approach the influence of ethnicity and environment on the neurophysiologic regulation of response to sensory stimuli among children. There are four main hypotheses tested in this thesis. Each hypothesis testing is summarised in the preceding paragraphs considering the specific research question it aims to answer.

- Conclusion for Hypothesis Testing 1. Hypothesis testing 1 aimed to determine the influence of ethnicity on the regulation of response to sensory stimuli by comparing two groups of typically developing children with different ethnicities but lives in the same geographic environments (Hong Kong) and environment landscapes (urban setting) were recruited. The results of this study found significant differences between CHK and FHK in their SCL patterns of change from one condition to another. Furthermore, this thesis found no significant differences between CHK and FHK groups on the level of ANS activity across the three experimental conditions.
- Conclusion for Hypothesis Testing 2. Hypothesis testing 2 sought to determine whether geographic environments influence the regulation of response to sensory stimuli by comparing two groups of typically developing children with the same ethnicity (Filipino) but lives in different geographic environments (Hong Kong and Philippines) and similar environment landscapes (urban settings). This study found significant differences between FHK and FU groups on their levels of autonomic activity across the three experimental conditions.
- Conclusion for Hypothesis Testing 3. Hypothesis testing 3 investigated the role of ethnicity and geographic environment on the regulation of response to sensory

stimuli by comparing two groups of typically developing children with different ethnicities living in geographic environments (Hong Kong and Philippines) but similar environment landscapes (urban setting) were recruited. This thesis found differences between CHK and FU groups on the levels of autonomic activity across the three experimental conditions.

• Conclusion for Hypothesis Testing 4. Hypothesis testing 4 aimed to explore the influence of the physical environment on the regulation of response to sensory stimuli by comparing two groups of children from similar ethnicities (Filipinos) and geographic environments (Philippines) living in different physical environments (urban and rural setting). This thesis found differences between FU and FR groups on the levels of autonomic activity across the three experimental conditions.

Future studies need to carefully consider the limitations of this thesis. First, this research suggests future efforts to control further for these individual differences at the level of participant recruitment to lessen their effects on the overall data. Second, future research needs to consider the use of behavioural outcome measures that may reflect similar underlying theoretical frameworks. Third, future research may explore more robust and high-end laboratory equipment as instruments of choice. Lastly, there will be a need to have future researches to replicate the findings of this research as applied to different population group characteristics.

II. Future Work Directions

The evidence produced in this thesis presents an interesting springboard for future work directions. First, the conceptual model used in this thesis can be applied in other areas of inquiry on the topic of adaptation. Sensory stimuli are a mere embodiment of external challenges. Other forms of external challenges such as stress and cognitive tasks can be explored further. It is recommended to include these constructs concurrently with sensory responses to explore their interactions and relationships. Second, the discrete use of multiphysiological autonomic measures presents a novel explanation of the regulatory mechanisms of adaptation. Future research undertakings may look into including CNS measures (i.e. EEG, ERP) to further explore central and peripheral neurophysiologic systems mechanisms that support the regulation of responses leading to adaptive abilities in children. Third, replication of the findings of this study among other ethnicities is essential for generalisation. Employing similar methods, the author of this thesis intends to partner

with other researchers across the international scene to gather data from children across the world to test whether the hypotheses substantiated in this research holds robust. Lastly, children of migrant origins raise an important query that needs to be further explored. With the growing rate of globalisation comes consequent migration of people. Although migration has economic benefits, its health impacts are yet to be fully understood. Thus, it is worthwhile to employ longitudinal research that aims to track and monitor the life history of migrant children as they set forth on a journey to the adaptation to a foreign land. The findings of these future work directions are anticipated to provide not only salient neuroscientific evidence on the neural mechanism of adaptation among children but likewise in informing global and local policies that aim to facilitate the successful adaptation of children of migrant origins.

References:

- Ali-Khan, S. E., Krakowski, T., Tahir, R., & Daar, A. S. (2011). The use of race, ethnicity and ancestry in human genetic research. *The HUGO Journal*, *5*(1-4), 47-63. doi:10.1007/s11568-011-9154-5
- American Psychiatric Association. *Diagnostic and statistical manual of mental disorders* (DSM-5), American Psychiatric Association, Lake St. Louis, Mo, USA, 2013.
- Andreassi, J. L. (2013). *Psychophysiology: Human behavior & physiological response* (4th ed.). Psychology Press.
- Appelhans, B. M., & Luecken, L. J. (2006). Heart rate variability as an index of regulated emotional responding. *Review of General Psychology*, *10*(3), 229–240. doi:10.1037/1089-2680.10.3.229
- Aron, A., Ketay, S., Hedden, T., Aron, E. N., Markus, H. R., & Gabrieli, J. D. (2010). Temperament trait of sensory processing sensitivity moderates cultural differences in neural response. *Social Cognitive and Affective Neuroscience*, 5(2-3), 219-226. doi: 10.1093/scan/nsq028
- Aston-Jones, G., Rajkowski, J., Kubiak, P., Valentino, R.J., & Shipley, M.T. (1996). Role of locus coeruleus in emotional activation. *Progress in Brain Research*, 107, 379-402. doi: 10.1016/S0079-6123(08)61877-4
- Aymerich-Franch, L., Petit, D., Ganesh, G., & Kheddar, A. (2017). Non-human looking robot arms induce illusion of embodiment. *International Journal of Social Robotics*, 9(4), 479-490. doi: 10.1007/s12369-017-0397-8
- Bach, D. R. (2014). A head-to-head comparison of SCRalyze and Ledalab, two model-based methods for skin conductance analysis. *Biological Psychology*, *103*, 63-68. doi: 10.1016/j.biopsycho.2014.08.006
- Bagot, R. C., & Meaney, M. J. (2010). Epigenetics and the biological basis of gene× environment interactions. *Journal of the American Academy of Child & Adolescent Psychiatry*, 49(8), 752-771. doi: 10.1016/j.jaac.2010.06.001
- Bakermans-Kranenburg, M. J., & van Ijzendoorn, M. H. (2011). Differential susceptibility to rearing environment depending on dopamine-related genes: New evidence and a

- meta-analysis. *Development and Psychopathology*, *23*(1), 39–52. http://doi.org/10.1017/S0954579410000635
- Baranek, G. T., Boyd, B. A., Poe, M. D., David, F. J., & Watson, L. R. (2007).

 Hyperresponsive sensory patterns in young children with autism, developmental delay, and typical development. *American Journal on Mental Retardation*, 112(4), 233-245. doi: 10.1352/0895-8017(2007)112[233:HSPIYC]2.0.CO;2
- Barbosa, R., da Costa, M. P., Silva, N. T. D., Azevedo, F. M., Pastre, C. M., & Vanderlei, L. C. M. (2016). Comparison of Polar® RS800G3TM heart rate monitor with Polar® S810iTM and electrocardiogram to obtain the series of RR intervals and analysis of heart rate variability at rest. *Clinical Physiology and Functional Imaging*, *36*(2), 112-117. doi: 10.1111/cpf.12203
- Baxter, M. G., & Murray, E. A. (2002). The amygdala and reward. *Nature Reviews Neuroscience*, *3*(7), 563. doi: 10.1038/nrn875
- Benedek, M., & Kaernbach, C. (2010). A continuous measure of phasic electrodermal activity. *Journal of Neuroscience Methods*, *190*(1), 80–91. doi:10.1016/j.jneumeth.2010.04.028
- Benevides, T. W., & Lane, S. J. (2015). A review of cardiac autonomic measures: considerations for examination of physiological response in children with autism spectrum disorder. *Journal of Autism and Developmental Disorders*, 45(2), 560-575. doi:10.1007/s10803-013-1971-z
- Berntson, G. G., Bigger, J. T., Eckberg, D. L., Grossman, P., Kaufmann, P. G., Malik, M., ... & Van Der Molen, M. W. (1997). Heart rate variability: origins, methods, and interpretive caveats. *Psychophysiology*, *34*(6), 623-648. doi: 10.1111/j.1469-8986.1997.tb02140.x
- Berntson, G. G., Cacioppo, J. T., & Quigley, K. S. (1991). Autonomic determinism: the modes of autonomic control, the doctrine of autonomic space, and the laws of autonomic constraint. *Psychological Review*, *98*(4), 459–487. doi:10.1037/0033-295X.98.4.459
- Berntson, G. G., Cacioppo, J. T., Binkley, P. F., Uchino, B. N., Quigley, K. S., & Fieldstone, A. (1994). Autonomic cardiac control. III. Psychological stress and

- cardiac response in autonomic space as revealed by pharmacological blockades. *Psychophysiology*, *31*(6), 599-608. doi:10.1111/j.1469-8986.1994.tb02352.x
- Berntson, G. G., Cacioppo, J. T., & Fieldstone, A. (1996). Illusions, arithmetic, and the bidirectional modulation of vagal control of the heart. *Biological Psychology*, *44*(1), 1-17. doi:10.1016/S0301-0511(96)05197-6
- Berntson, G. G., Quigley, K. S., Jang, J. F., & Boysen, S. T. (1990). An approach to artifact identification: Application to heart period data. *Psychophysiology*, 27(5), 586-598. doi: 10.1111/j.1469-8986.1990.tb01982.x
- Berthold, P., Gwinner, E., & Sonnenschein, E. (Eds.). (2013). *Avian migration*. Springer Science & Business Media.
- Bock, W. J. (1980). The definition and recognition of biological adaptation. *American Zoologist*, 20(1), 217-227. doi: 10.1093/icb/20.1.217
- Bortoluzzi, A., Blaya, C., Rosa, E. D. da, Paim, M., Rosa, V., Leistner-Segal, S., & Manfro, G. G. (2015). What can HPA axis-linked genes tell us about anxiety disorders in adolescents? *Trends in Psychiatry and Psychotherapy*, *37*(4), 232–7. doi:10.1590/2237-6089-2015-0035
- Boucsein W. (2012). *Electrodermal Activity*. Springer Science & Business Media, Berlin, Germany.
- Boyce, W. T., & Ellis, B. J. (2005). Biological sensitivity to context: I. An evolutionary—developmental theory of the origins and functions of stress reactivity. *Development and Psychopathology*, *17*(2), 271-301. doi:10.1017/S0954579405050145
- Brett-Green, B. A., Miller, L. J., Gavin, W. J., & Davies, P. L. (2008). Multisensory integration in children: A preliminary ERP study. *Brain Research*, *1242*, 283-290. doi: 10.1016/j.brainres.2008.03.090
- Brown, N. B., & Dunn, W. (2010). Relationship between context and sensory processing in children with autism. *American Journal of Occupational Therapy*, 64(3), 474-483. doi:10.5014/ajot.2010.09077
- Bundy, A. C., Shia, S., Long, Q., & Miller, L. J. (2007). How does sensory processing dysfunction affect play?. *American Journal of Occupational Therapy*, 61(2), 201. doi: 10.5014/ajot.61.2.201

- Cabiddu, R., Cerutti, S., Viardot, G., Werner, S., & Bianchi, A. M. (2012). Modulation of the sympatho-vagal balance during sleep: frequency domain study of heart rate variability and respiration. *Frontiers in Physiology*, *3*(45). doi:10.3389/fphys.2012.00045
- Cacioppo, J. T., Tassinary, L. G., & Berntson, G. (Eds.). (2007). *Handbook of psychophysiology*. Cambridge University Press.
- Cacioppo, J. T., Tassinary, L. G., & Berntson, G. G. (2000). *Psychophysiological science*. *Handbook of psychophysiology*, 2, 3-23.
- Cacioppo, J.T., Berntson, G.G., & Klein, D.J. (1992). What is an emotion? The role of somatovisceral 'illusions'. *Review of Personality and Social Psychology*, *14*, 63-98.
- Caprio, S., Daniels, S. R., Drewnowski, A., Kaufman, F. R., Palinkas, L. A., Rosenbloom, A. L., & Schwimmer, J. B. (2008). Influence of race, ethnicity, and culture on childhood obesity: implications for prevention and treatment: a consensus statement of Shaping America's Health and the Obesity Society. *Diabetes Care*, 31(11), 2211–2221. doi:10.2337/dc08-9024
- Caron, K. G., Schaaf, R. C., Benevides, T. W., & Gal, E. (2012). Cross-cultural comparison of sensory behaviors in children with autism. *American Journal of Occupational Therapy*, 66(5), e77-e80. doi:10.5014/ajot.2012.004226
- Castiglioni, P., Parati, G., Di Rienzo, M., Carabalona, R., Cividjian, A., & Quintin, L. (2011). Scale exponents of blood pressure and heart rate during autonomic blockade as assessed by detrended fluctuation analysis. *The Journal of Physiology*, 589(2), 355-369. doi:10.1113/jphysiol.2010.196428
- Champagne, F. A. (2013). Early environments, glucocorticoid receptors, and behavioral epigenetics. *Behavioral Neuroscience*, *127*(5), 628. doi: 10.1037/a0034186
- Chang, M. C., Parham, L. D., Blanche, E. I., Schell, A., Chou, C.-P., Dawson, M., & Clark, F. (2012). Autonomic and behavioral responses of children with autism to auditory stimuli. *The American Journal of Occupational Therapy*, 66(5), 567–576. doi:10.5014/ajot.2012.004242
- Chatterjee, D., Sinha, A., Sinha, M., & Saha, S. K. (2016). A Probabilistic Approach for Detection and Analysis of Cognitive Flow. *BMAW* 2016.

- Cheung, P. P. P., & Siu, A. M. H. (2010). Chinese Sensory Profile: users' manual.
- Christou, C., Herakleous, K., Tzanavari, A., & Poullis, C. (2015, September).

 Psychophysiological responses to virtual crowds: Implications for wearable computing. *In Affective Computing and Intelligent Interaction (ACII)*, 2015

 International Conference on (pp. 35-41). IEEE. doi: 10.1109/ACII.2015.7344548
- Cicchetti, D. (2010). Resilience under conditions of extreme stress: a multilevel perspective. *World Psychiatry*, *9*(3), 145-154. doi: 10.1002/j.2051-5545.2010.tb00297.x
- Cloninger, C. R., Sigvardsson, S., Bohman, M., & Von Knorring, A. L. (1982).

 Predisposition to petty criminality in Swedish adoptees: II. Cross-fostering analysis of gene-environment interaction. *Archives of General Psychiatry*, *39*(11), 1242-1247. doi: 10.1001/archpsyc.1982.04290110010002
- Cohen, J. (1992). A power primer. *Psychological Bulletin*, *112*(1), 155-159. doi:10.1037/0033-2909.112.1.155
- Collet, C., Di Rienzo, F., Hoyek, N., & Guillot, A. (2013). Autonomic nervous system correlates in movement observation and motor imagery. *Frontiers in Human Neuroscience*, 7(415). doi:10.3389/fnhum.2013.00415
- Colombo, J., Arora, R., DePace, N. L., & Ball, C. (2014). *Clinical Autonomic Dysfunction: Measurement, Indications, Therapies, and Outcomes*. Switzerland: Springer International Publishing.
- Cornforth, D., Jelinek, H. F., & Tarvainen, M. (2015). A Comparison of Nonlinear Measures for the Detection of Cardiac Autonomic Neuropathy from Heart Rate Variability. *Entropy*, *17*(3), 1425-1440. doi:10.3390/e17031425
- Cortessis, V. K., Thomas, D. C., Levine, A. J., Breton, C. V., Mack, T. M., Siegmund, K. D., ... & Laird, P. W. (2012). Environmental epigenetics: prospects for studying epigenetic mediation of exposure–response relationships. *Human Genetics*, *131*(10), 1565-1589.
- Critchley, H. D. (2002). Book review: electrodermal responses: what happens in the brain. *The Neuroscientist*, 8(2), 132-142. doi:10.1177/107385840200800209

- Cui, L., Morris, A. S., Harrist, A. W., Larzelere, R. E., Criss, M. M., & Houltberg, B. J. (2015). Adolescent RSA responses during an anger discussion task: Relations to emotion regulation and adjustment. *Emotion*, *15*(3), 360. doi:10.1037/emo0000040
- Cysarz, D., Van Leeuwen, P., Edelhäuser, F., Montano, N., & Porta, A. (2012). Binary symbolic dynamics classifies heart rate variability patterns linked to autonomic modulations. *Computers in Biology and Medicine*, *42*(3), 313-318. doi:10.1016/j.compbiomed.2011.04.013
- Daluwatte, C., Miles, J. H., Sun, J., & Yao, G. (2015). Association between pupillary light reflex and sensory behaviors in children with autism spectrum disorders. *Research in Developmental Disabilities*, *37*, 209–215. doi:10.1016/j.ridd.2014.11.019
- Danese, A., & McEwen, B. S. (2012). Adverse childhood experiences, allostasis, allostatic load, and age-related disease. *Physiology & Behavior*, *106*(1), 29-39. doi:10.1016/j.physbeh.2011.08.019
- Davis, M., & Whalen, P. J. (2001). The amygdala: vigilance and emotion. *Molecular Psychiatry*, 6(1), 13.
- Dawson, M. E., & Schell, A. M. (2002). What does electrodermal activity tell us about prognosis in the schizophrenia spectrum? *Schizophrenia Research*, *54*(1), 87-93. doi:10.1016/S0920-9964(01)00355-3
- Dawson, M. E., Schell, A. M., & Filion, D. L. (2007). *The electrodermal system. Handbook of Psychophysiology*, 2, 200-223.
- De Rooij, S. R., van Eijsden, M., Roseboom, T. J., & Vrijkotte, T. G. (2013). Ethnic differences in childhood autonomic nervous system regulation. *International Journal of Cardiology*, *168*(5), 5064-5066. doi:10.1016/j.ijcard.2013.07.202
- DeRijk, R. H. (2009). Single nucleotide polymorphisms related to HPA axis reactivity. *Neuroimmunomodulation*, *16*(5), 340-352. http://doi.org/10.1159/000216192
- Dibbets, P., Poort, H., & Arntz, A. (2012). Adding imagery rescripting during extinction leads to less ABA renewal. *Journal of Behavior Therapy and Experimental Psychiatry*, 43(1), 614-624. doi:10.1016/j.jbtep.2011.08.006
- Dieleman, G. C., Huizink, A. C., Tulen, J. H., Utens, E. M., Creemers, H. E., van der Ende, J., & Verhulst, F. C. (2015). Alterations in HPA-axis and autonomic nervous

- system functioning in childhood anxiety disorders point to a chronic stress hypothesis. *Psychoneuroendocrinology*, 51, 135-150. doi:10.1016/j.psyneuen.2014.09.002
- Dingle, H., & Drake, V. A. (2007). What is migration?. *Bioscience*, 57(2), 113-121.
- Doamekpor, L. A., & Dinwiddie, G. Y. (2015). Allostatic load in foreign-born and US-born blacks: evidence from the 2001–2010 National Health and Nutrition Examination Survey. *American Journal of Public Health*, 105(3), 591-597. doi: 10.2105/AJPH.2014.302285
- Doyle-Portillo, S., & Pastorino, E. (2015). What is Psychology?. South-Western.
- Dunn, W. (1999). *The sensory profile manual*. San Antonio, TX: Psychological Corporation.
- Dunn, W. (2007). Supporting children to participate successfully in everyday life by using sensory processing knowledge. *Infants & Young Children*, 20(2), 84-101. doi: 10.1097/01.IYC.0000264477.05076.5d
- Duong, M. T., Bingham, B. A., Aldana, P. C., Chung, S. T., & Sumner, A. E. (2017). Variation in the calculation of allostatic load score: 21 examples from NHANES. *Journal of Racial and Ethnic Health Disparities*, 4(3), 455-461. doi: 10.1007/s40615-016-0246-8
- Durrant, J. D., & Lovrinic, J. H. (1984). *Bases of hearing science*. Baltimore: Williams & Wilkins.
- Dworkin, B.R. (2000). Interoception. In J.T. Cacioppo, L.G. Tassinary, & G.G. Berntson (Eds.), *Handbook of Psychophysiology* (pp. 482-505). Cambridge, UK: Cambridge University Press.
- Eckberg, D. L. (1997). Sympathovagal balance a critical appraisal. *Circulation*, *96*(9), 3224-3232. doi:10.1161/01.CIR.96.9.3224
- Edelberg, R. (1983). The effects of initial levels of sweat duct filling and skin hydration on electrodermal response amplitude. *Psychophysiology*, 20(5), 550-557. doi: 10.1111/j.1469-8986.1983.tb03012.x

- Edelberg, R. (1993). *Electrodermal mechanisms: A critique of the two-effector hypothesis and a proposed replacement*. In Progress in electrodermal research (pp. 7-29). Springer, Boston, MA.
- Efron, B. (1987). Better bootstrap confidence intervals. *Journal of the American Statistical Association*, 82(397), 171-185. doi:10.1080/01621459.1987.10478410
- Efron, Bradley. (1994). "Missing data, imputation, and the bootstrap." *Journal of the American Statistical Association* 89(426), 463-475. doi:10.1080/01621459.1994.10476768
- Engel-Yeger, B. (2010). The applicability of the short sensory profile for screening sensory processing disorders among Israeli children. *International Journal of Rehabilitation Research*, *33*(4), 311-318. doi:10.1097/MRR.0b013e32833abe59
- Evans, G.W., & English, K. (2002). The Environment of Poverty: Multiple Stressor Exposure, Psychophysiological Stress, and Socioemotional Adjustment. *Child Development*, 73(4), 1238-1248. doi:10.1111/1467-8624.00469
- Everly Jr, G. S., & Lating, J. M. (2012). A clinical guide to the treatment of the human stress response. Springer Science & Business Media.
- Fabian, D. & Flatt, T. (2012) Life History Evolution. *Nature Education Knowledge* 3(10):24. doi:
- Fazel, M., Reed, R. V., Panter-Brick, C., & Stein, A. (2012). Mental health of displaced and refugee children resettled in high-income countries: Risk and protective factors. *The Lancet*. doi:10.1016/S0140-6736(11)60051-2
- Fowles, D. C., Christie, M. J., Edelberg, R., Grings, W. W., Lykken, D. T., & Venables, P. H. (1981). Publication recommendations for electrodermal measurements.

 Psychophysiology, 18(3), 232-239. doi:10.1111/j.1469-8986.1981.tb03024.x
- Fredrikson, M., Furmark, T., Olsson, M. T., Fischer, H., Andersson, J., & Långström, B. (1998). Functional neuroanatomical correlates of electrodermal activity: a positron emission tomographic study. *Psychophysiology*, *35*(2), 179–185. doi:10.1111/1469-8986.3520179

- Fryxell, J. M., & Sinclair, A. R. E. (1988). Causes and consequences of migration by large herbivores. *Trends in Ecology & Evolution*, *3*(9), 237-241. doi: 10.1016/0169-5347(88)90166-8
- Futuyma, D. J. (2009). *Natural selection and adaptation*. Evolution. Massachusetts, USA: Sinauer Associates, Inc. Sunderland, 279-301.
- Gamer, M. (2011). Detecting concealed information using autonomic measures. *Memory Detection: Theory and Application of the Concealed Information Test*, 27-45.
- Ganger, S., Hahn, A., Küblböck, M., Kranz, G. S., Spies, M., Vanicek, T., ...

 Lanzenberger, R. (2015). Comparison of continuously acquired resting state and extracted analogues from active tasks. *Human Brain Mapping*, *36*(10), 4053–4063. doi:10.1002/hbm.22897
- Ganzel, B. L., & Morris, P. A. (2011). Allostasis and the developing human brain: Explicit consideration of implicit models. *Development and Psychopathology*, 23(4), 955-974. doi:10.1017/S0954579411000447
- Ganzel, B. L., Morris, P. A., & Wethington, E. (2010). Allostasis and the human brain: Integrating models of stress from the social and life sciences. *Psychological Review*, *117*(1), 134. doi:10.1037/a0017773
- Garrido, M. I., Barnes, G. R., Sahani, M., & Dolan, R. J. (2012). Functional evidence for a dual route to amygdala. *Current Biology*, 22(2), 129-134. doi:10.1016/j.cub.2011.11.056
- Gelfand, S. A., & Levitt, H. (2004). *Hearing—an introduction to psychological and physiological acoustics*. Marcel Dekker. New York.
- Ghanizadeh, A. (2011). Sensory processing problems in children with ADHD, a systematic review. *Psychiatry Investigation*, 8(2), 89-94. doi:10.4306/pi.2011.8.2.89
- Goldstein, D. S., Bentho, O., Park, M. Y., & Sharabi, Y. (2011). Low-frequency power of heart rate variability is not a measure of cardiac sympathetic tone but may be a measure of modulation of cardiac autonomic outflows by baroreflexes.

 Experimental Physiology, 96(12), 1255-1261. doi:10.1113/expphysiol.2010.056259
- Goldstein, D. S., Bentho, O., Park, M. Y., & Sharabi, Y. (2011). Low-frequency power of heart rate variability is not a measure of cardiac sympathetic tone but may be a

- measure of modulation of cardiac autonomic outflows by baroreflexes. Experimental Physiology, 96(12), 1255-1261. doi: 10.1113/expphysiol.2010.056259
- Goldstein, S., & Goldstein, A. (1981). The impact of migration on fertility: an 'own children' analysis for Thailand. *Population Studies*, *35*(2), 265-284. doi:10.1080/00324728.1981.10404967
- Goldstein, S., Naglieri, J. A., Princiotta, D., & Otero, T. M. (2014). *Introduction: a history of executive functioning as a theoretical and clinical construct. In Handbook of executive functioning* (pp. 3-12). Springer, New York, NY.
- Gomez, I. N., Lai, C. Y., Chan, C. C., & Tsang, H. W. (2018). The Role of Ethnicity and Environment in the Regulation of Response to Sensory Stimulus in Children: Protocol and Pilot Findings of a Neurophysiological Study. *JMIR Research Protocols*, 7(1). doi:10.2196/resprot.8157
- Gomez, I. N., Lai, C. Y., Morato-Espino, P. G., Chan, C. C., & Tsang, H. W. (2017).

 Behavioural and Autonomic Regulation of Response to Sensory Stimuli among
 Children: A Systematic Review of Relationship and Methodology. BioMed
 Research International, 2017. doi:10.1155/2017/2629310
- Gomez, I. N., Lai, C. Y., Yung, T. W., Chan, C. C., & Tsang, H. W. (2018). Migration Influences on the Allostatic Load of Children: Systematic Review Protocol. JMIR Research Protocols, 7(1). doi:10.2196/resprot.8332
- Goodwin, C. J. (2015). A History of Modern Psychology. John Wiley & Sons.
- Gräff, J., & Mansuy, I. M. (2008). Epigenetic codes in cognition and behaviour. *Behavioural Brain Research*, 192(1), 70-87. doi:10.1016/j.bbr.2008.01.021
- Grassi-Oliveira, R., Ashy, M., & Stein, L. M. (2008). Psychobiology of childhood maltreatment: effects of allostatic load? *Revista Brasileira de Psiquiatria*, 30(1), 60-68. doi:10.1590/S1516-44462008000100012
- Grossman, P., & Taylor, E.W. (2007). Toward understanding respiratory sinus arrhythmia: Relations to cardiac vagal tone, evolution and biobehavioral functions. *Biological Psychology*, 74(2), 263-285. doi:10.1016/j.biopsycho.2005.11.014

- Gunn, T. E., Tavegia, B. D., Houskamp, B. M., McDonald, L. B., Bustrum, J. M., Welsh, R. K., & Mok, D. S. (2009). Relationship between sensory deficits and externalizing behaviors in an urban, Latino preschool population. *Journal of Child and Family Studies*, 18(6), 653–661. doi:10.1007/s10826-009-9266-x
- Hainsworth, R., Drinkhill, M. J., & Rivera-Chira, M. (2007). The autonomic nervous system at high altitude. *Clinical Autonomic Research*, 17(1), 13-19. doi:10.1007/s10286-006-0395-7
- Hakim, F., Gozal, D., & Gozal, L. K. (2012). Sympathetic and catecholaminergic alterations in sleep apnea with particular emphasis on children. *Frontiers in Neurology*, *3*(7). doi:10.3389/fneur.2012.00007
- Hardaway, C. R., Wilson, M. N., Shaw, D. S., & Dishion, T. J. (2012). Family functioning and externalizing behaviour among low-income children: Self-regulation as a mediator. *Infant and Child Development*, 21(1), 67-84. doi:10.1002/icd.765
- Harms, N. J., Legagneux, P., Gilchrist, H. G., Bêty, J., Love, O. P., Forbes, M. R., ... & Soos, C. (2015). Feather corticosterone reveals effect of moulting conditions in the autumn on subsequent reproductive output and survival in an Arctic migratory bird. Proceedings of the Royal Society of London B: Biological Sciences, 282(1800), 20142085. doi:10.1098/rspb.2014.2085
- Hertzman, C. (2012). Putting the concept of biological embedding in historical perspective. *Proceedings of the National Academy of Sciences*, 109(Supplement 2), 17160-17167. doi:10.1073/pnas.1202203109
- Hesterberg, T. C. (2015). What teachers should know about the bootstrap: Resampling in the undergraduate statistics curriculum. *The American Statistician*, 69(4), 371-386. doi:10.1080/00031305.2015.1089789
- Ho, S. M., Johnson, A., Tarapore, P., Janakiram, V., Zhang, X., & Leung, Y. K. (2012). Environmental epigenetics and its implication on disease risk and health outcomes. *ILAR Journal*, *53*(3-4), 289-305. doi:10.1093/ilar.53.3-4.289
- Hörmann, T., Hesse, M., Christ, P., Adams, M., Menßen, C., & Rückert, U. (2016). Fine-grained prediction of cognitive workload in a modern working environment by utilizing short-term physiological parameters. *In Proceedings of the 9th*

- International Joint Conference on Biomedical Engineering Systems and Technologies (Vol. 4). doi:10.5220/0005665000420051
- Hoyle, R. H., Harris, M. J., & Judd, C. M. (2002). *Research methods in social relations*. Thomson Learning.
- Huitt, W. (2003). The information processing approach to cognition. *Educational Psychology Interactive*, *3*(2), 53-67.
- Jachowski, D. S., & Singh, N. J. (2015). Toward a mechanistic understanding of animal migration: incorporating physiological measurements in the study of animal movement. *Conservation Physiology*, *3*(1), cov035. doi:10.1093/conphys/cov035
- Jackson, F. L. (1992). Race and ethnicity as biological constructs. *Ethnicity and Disease*, 2(2), 120-125.
- Japundzic, N., Grichois, M. L., Zitoun, P., Laude, D., & Elghozi, J. L. (1990). Spectral analysis of blood pressure and heart rate in conscious rats: effects of autonomic blockers. *Journal of the Autonomic Nervous System*, 30(2), 91-100. doi:10.1016/0165-1838(90)90132-3
- Järvinen, A., Ng, R., Crivelli, D., Neumann, D., Arnold, A. J., Woo-VonHoogenstyn, N., ... & Bellugi, U. (2016). Social functioning and autonomic nervous system sensitivity across vocal and musical emotion in Williams syndrome and autism spectrum disorder. *Developmental Psychobiology*, 58(1), 17-26. doi:10.1002/dev.21335
- Johnston-Brooks, C. H., Lewis, M. A., Evans, G. W., & Whalen, C. K. (1998). Chronic stress and illness in children: the role of allostatic load. *Psychosomatic Medicine*, 60(5), 597-603. doi:10.1.1.534.1257
- Jorde, L. B., & Wooding, S. P. (2004). Genetic variation, classification and 'race'. *Nature Genetics*, *36*, S28-S33. doi10.1038/ng1435
- Juster, R. P., Sindi, S., Marin, M. F., Perna, A., Hashemi, A., Pruessner, J. C., & Lupien, S. J. (2011). A clinical allostatic load index is associated with burnout symptoms and hypocortisolemic profiles in healthy workers. *Psychoneuroendocrinology*, 36(6), 797-805. doi:10.1016/j.psyneuen.2010.11.001

- Kandel, E. R., & Jessell, T. (1995). Early experience and the fine tuning of synaptic connections. In E.R. Kandel, J.H. Schwartz, & T. Jessell (Eds.), *Essentials of Neural Science and Behavior*, (pp. 1247-1279). Norwalk, CT: Appleton & Lange.
- Karatsoreos, I. N., & McEwen, B. S. (2013). Annual research review: the neurobiology and physiology of resilience and adaptation across the life course. *Journal of Child Psychology and Psychiatry*, *54*(4), 337-347. doi:10.1111/jcpp.12054
- Karlamangla, A. S., Singer, B. H., McEwen, B. S., Rowe, J. W., & Seeman, T. E. (2002). Allostatic load as a predictor of functional decline: MacArthur studies of successful aging. *Journal of Clinical Epidemiology*, 55(7), 696-710. doi:10.1016/S0895-4356(02)00399-2
- Kempermann, G., van Praag, H., & Gage, F. H. (1999). Activity-dependent regulation of neuronal plasticity and self-repair. *Progress in Brain Research*, 127, 35-48. doi:10.1016/S0079-6123(00)27004-0
- Keuroghlian, A. S., & Knudsen, E. I. (2007). Adaptive auditory plasticity in developing and adult animals. *Progress in Neurobiology*, 82(3), 109-121. doi:10.1016/j.pneurobio.2007.03.005
- Klahr, A. M., & Burt, S. A. (2014). Elucidating the etiology of individual differences in parenting: A meta-analysis of behavioral genetic research. *Psychological Bulletin*, 140(2), 544–586. doi:10.1037/a0034205
- Kleiger, R. E., Stein, P. K., & Bigger, J. T. (2005). Heart rate variability: measurement and clinical utility. *Annals of Noninvasive Electrocardiology*, *10*(1), 88-101. doi:10.1111/j.1542-474X.2005.10101.x
- Knafo, A., Israel, S., & Ebstein, R. P. (2011). Heritability of children's prosocial behavior and differential susceptibility to parenting by variation in the dopamine receptor D4 gene. *Development and Psychopathology*, 23(1), 53-67. doi:10.1017/S0954579410000647
- Kohrs, C., Hrabal, D., Angenstein, N., & Brechmann, A. (2014). Delayed system response times affect immediate physiology and the dynamics of subsequent button press behavior. *Psychophysiology*, *51*(11), 1178-1184. doi:10.1111/psyp.12253

- Kopp, C. B. (1982). Antecedents of self-regulation: a developmental perspective. *Developmental Psychology*, 18(2), 199. doi:10.1037/0012-1649.18.2.199
- Korte, S. M., Koolhaas, J. M., Wingfield, J. C., & McEwen, B. S. (2005). The Darwinian concept of stress: benefits of allostasis and costs of allostatic load and the trade-offs in health and disease. *Neuroscience & Biobehavioral Reviews*, 29(1), 3-38. doi:10.1016/j.neubiorev.2004.08.009
- Krabs, R. U., Enk, R., Teich, N., & Koelsch, S. (2015). Autonomic effects of music in health and Crohn's disease: the impact of isochronicity, emotional valence, and tempo. *Plos One*, *10*(5), e0126224. doi:10.1371/journal.pone.0126224
- Kraus, K. S., & Canlon, B. (2012). Neuronal connectivity and interactions between the auditory and limbic systems. Effects of noise and tinnitus. *Hearing Research*, 288(1-2), 34-46. doi: 10.1016/j.heares.2012.02.009
- Kreibig, S. D. (2010). Autonomic nervous system activity in emotion: A review. *Biological Psychology*, 84(3), 394-421. doi:10.1016/j.biopsycho.2010.03.010
- Kuhlman, K. R., Geiss, E. G., Vargas, I., & Lopez-Duran, N. L. (2015). Differential associations between childhood trauma subtypes and adolescent HPA-axis functioning. *Psychoneuroendocrinology*, *54*, 103–114. doi:10.1016/j.psyneuen.2015.01.020
- Kuo, T. B., & Chan, S. H. (1993). Continuous, on-line, real-time spectral analysis of systemic arterial pressure signals. *American Journal of Physiology-Heart and Circulatory Physiology*, 264(6), H2208-H2213. doi:10.1152/ajpheart.1993.264.6.H2208
- Lai C. Y. Y. & Chiu A. S. M. (2013) Sensory Processing and Self-Regulation Checklist, Heep Hong Society, Hong Kong.
- Lai, Y. Y. C. (2013). A study of behavioral and autonomic responses in autistic children with sensory processing difficulty (Doctoral dissertation, The Hong Kong Polytechnic University).
- Lajante, M., & Droulers, O. (2012). Measuring arousal in consumer research: a new EDA signal processing method. *NA-Advances in Consumer Research*, 40.

- Lane, S. J., & Schaaf, R. C. (2010). Examining the Neuroscience Evidence for Sensory-Driven Neuroplasticity: Implications for Sensory-Based Occupational Therapy for Children and Adolescents. *American Journal of Occupational Therapy*, 64(3), 375–390. doi:10.5014/ajot.2010.09069
- Lane, S. J., Reynolds, S., & Thacker, L. (2010). Sensory over-responsivity and ADHD: Differentiating using electrodermal responses, cortisol, and anxiety. *Frontiers in Integrative Neuroscience*, 4(8). doi:10.3389/fnint.2010.00008
- Lapate, R. C., Rokers, B., Li, T., & Davidson, R. J. (2014). Nonconscious emotional activation colors first impressions: a regulatory role for conscious awareness. *Psychological Science*, 25(2), 349–357. doi:10.1177/0956797613503175
- Laumann, K., Gärling, T., & Stormark, K. M. (2003). Selective attention and heart rate responses to natural and urban environments. *Journal of Environmental Psychology*, 23(2), 125–134. doi:10.1016/S0272-4944(02)00110-X
- LeDoux, J. (2007). The amygdala. *Current Biology*, *17*(20), R868-R874. doi:10.1016/j.cub.2007.08.005
- Lee, D. H., & Minard, D. (Eds.). (2013). Physiology, Environment, and Man: Based on a Symposium Conducted by the National Academy of Sciences–National Research Council, August, 1966. Elsevier.
- Leekam, S. R., Nieto, C., Libby, S. J., Wing, L., & Gould, J. (2007). Describing the sensory abnormalities of children and adults with autism. *Journal of Autism and Developmental Disorders*, *37*(5), 894-910. doi:10.1007/s10803-006-0218-7
- Liapis, A., Katsanos, C., Sotiropoulos, D. G., Karousos, N., & Xenos, M. (2017). Stress in interactive applications: analysis of the valence-arousal space based on physiological signals and self-reported data. *Multimedia Tools and Applications*, 76(4), 5051-5071. doi: 10.1007/s11042-016-3637-2
- Lin, C.-K., Wu, H.-M., Wang, H.-Y., Tseng, M.-H., & Lin, C.-H. (2013). Age as a factor in sensory integration function in Taiwanese children. *Neuropsychiatric Disease and Treatment*, *9*, 995–1001. doi:10.2147/NDT.S49514

- Logan, J. G., & Barksdale, D. J. (2008). Allostasis and allostatic load: expanding the discourse on stress and cardiovascular disease. *Journal of Clinical Nursing*, 17(7b), 201-208.
- Lole, L., Gonsalvez, C. J., Barry, R. J., & Blaszczynski, A. (2014). Problem gamblers are hyposensitive to wins: An analysis of skin conductance responses during actual gambling on electronic gaming machines. *Psychophysiology*, *51*(6), 556-564. doi:10.1111/psyp.12198
- Lopizzo, N., Bocchio Chiavetto, L., Cattane, N., Plazzotta, G., Tarazi, F. I., Pariante, C. M., ... & Cattaneo, A. (2015). Gene–environment interaction in major depression: focus on experience-dependent biological systems. *Frontiers in Psychiatry*, 6(68). doi:10.3389/fpsyt.2015.00068
- Losos, J. B. (2011). Convergence, adaptation, and constraint. *Evolution*, 65(7), 1827-1840. doi:10.1111/j.1558-5646.2011.01289.x
- Lovallo, W. R., & Sollers, J. J. (2007). Autonomic nervous system.
- Malik, M. & Camm, A. J. (2004). *Dynamic Electrocardiography*. Elmsford, New York, USA: Blackwell/Futura.
- Malik, M., & Camm, A. J. (Eds.). (1995). *Heart rate variability*. Futura publishing company.
- Malliani, A., Lombardi, F., & Pagani, M. (1994). Power spectrum analysis of heart rate variability: a tool to explore neural regulatory mechanisms. *British Heart Journal*, 71(1), 1.
- Malliani, A., Pagani, M., Lombardi, F., & Cerutti, S. (1991). Cardiovascular neural regulation explored in the frequency domain. *Circulation*, 84, 482–492. doi:10.1161/01.CIR.84.2.482
- Malpas, S. C. (2002). Neural influences on cardiovascular variability: possibilities and pitfalls. *American Journal of Physiology-Heart and Circulatory Physiology*, 282(1), H6-H20. doi:10.1152/ajpheart.2002.282.1.H6
- Mann, C. J. (2003). Observational research methods. Research design II: cohort, cross sectional, and case-control studies. *Emergency Medicine Journal*, 20(1), 54-60. doi:10.1136/emj.20.1.54

- Margolin, G., & Vickerman, K. A. (2007). Post-traumatic Stress in Children and Adolescents Exposed to Family Violence: I. Overview and Issues. *Professional Psychology, Research and Practice*, *38*(6), 613–619. doi:10.1037/0735-7028.38.6.613
- Marieb, E. N., & Hoehn, K. (2007). Human anatomy & physiology. Pearson Education.
- Marsh, R. A., Fuzessery, Z. M., Grose, C. D., & Wenstrup, J. J. (2002). Projection to the inferior colliculus from the basal nucleus of the amygdala. *Journal of Neuroscience*, 22(23), 10449-10460. doi:10.1523/JNEUROSCI.22-23-10449.2002
- Marsman, R., Nederhof, E., Rosmalen, J. G. M., Oldehinkel, A. J., Ormel, J., & Buitelaar, J. K. (2012). Family environment is associated with HPA-axis activity in adolescents. The TRAILS study. *Biological Psychology*, 89(2), 460–466. doi:10.1016/j.biopsycho.2011.12.013
- Martini, F. H., Nath, J. L., & Bartholomew, E. F. (2010). *Blood vessels and circulation*. *Foundamentals of Anatomy and Physiology*, 9th ed., Benjamin Cummings, Boston, MA, 707-763.
- Masterton, B. (1993). Central auditory system. *Otorhinolaryngology*, 55(3), 159-163.
- Matsubara, T., Funato, H., Kobayashi, A., Nobumoto, M., & Watanabe, Y. (2006).

 Reduced Glucocorticoid Receptor alpha Expression in Mood Disorder Patients and First-Degree Relatives. *Biological Psychiatry*, *59*(8), 689–695.

 doi:10.1016/j.biopsych.2005.09.026
- Matsushima, K., Matsubayashi, J., Toichi, M., Funabiki, Y., Kato, T., Awaya, T., & Kato, T. (2016). Unusual sensory features are related to resting-state cardiac vagus nerve activity in autism spectrum disorders. *Research in Autism Spectrum Disorders*, 25, 37–46. doi:10.1016/j.rasd.2015.12.006
- Matthews, K. A. (1986). *Handbook of stress, reactivity, and cardiovascular disease* (Vol. 6). Wiley-Interscience.
- McCormick, C., Hessl, D., Macari, S. L., Ozonoff, S., Green, C., & Rogers, S. J. (2014). Electrodermal and behavioral responses of children with autism spectrum disorders to sensory and repetitive stimuli. *Autism Research*, 7(4), 468-480. doi:10.1002/aur.1382

- McDonnell, M. J., & Pickett, S. T. (1990). Ecosystem structure and function along urbanrural gradients: an unexploited opportunity for ecology. *Ecology*, 71(4), 1232-1237. doi:10.2307/1938259
- McEwen, B S. (1998). Protective and damaging effects of stress mediators. *The New England Journal of Medicine*, *338*(3), 171–179. doi:10.1056/NEJM199801153380307
- McEwen, B S. (2000). The neurobiology of stress: from serendipity to clinical relevance. *Brain Research*, 886(1-2), 172–189. doi:10.1016/S0006-8993(00)02950-4
- McEwen, B. S. (1998). Stress, adaptation, and disease. Allostasis and allostatic load. *Annals of the New York Academy of Sciences*, 840, 33–44. doi:10.1111/j.1749-6632.1998.tb09546.x
- McEwen, B. S. (1999). Stress and hippocampal plasticity. *Annual Review of Neuroscience*, 22, 105–122. doi:10.1146/annurev.neuro.22.1.105
- McEwen, B. S. (2000). Allostasis and allostatic load: implications for neuropsychopharmacology. *Neuropsychopharmacology*, 22(2), 108-124. doi:10.1016/S0893-133X(99)00129-3
- McEwen, B. S. (2003). Interacting mediators of allostasis and allostatic load: Towards an understanding of resilience in aging. *Metabolism: Clinical and Experimental*. doi:10.1016/S0026-0495(03)00295-6
- McEwen, B. S. (2004). Protection and damage from acute and chronic stress: allostasis and allostatic overload and relevance to the pathophysiology of psychiatric disorders.

 Annals of the New York Academy of Sciences, 1032(1), 1-7.

 doi:10.1196/annals.1314.001
- McEwen, B. S. (2007). Physiology and neurobiology of stress and adaptation: central role of the brain. *Physiological Reviews*, 87(3), 873-904. doi:10.1152/physrev.00041.2006
- McEwen, B. S. (2008). Central effects of stress hormones in health and disease:

 Understanding the protective and damaging effects of stress and stress mediators.

 European Journal of Pharmacology, 583(2-3), 174-185.

 doi:10.1016/j.ejphar.2007.11.071

- McEwen, B. S. (2012). Brain on stress: how the social environment gets under the skin. *Proceedings of the National Academy of Sciences*, 109(Supplement 2), 17180-17185. doi:10.1073/pnas.1121254109
- McEwen, Bruce S, & Wingfield, J. C. (2003). The concept of allostasis in biology and biomedicine. *Hormones and Behavior*, 43(1), 2–15. doi:10.1016/S0018-506X(02)00024-7
- McEwen, Bruce S, & Wingfield, J. C. (2010). What is in a name? Integrating homeostasis, allostasis and stress. *Hormones and Behavior*, *57*(2), 105–111. doi:10.1016/j.yhbeh.2009.09.011
- McGaugh, J.L., Roozendall, B., & Cahill, L. (2000). Modulation of memory storage by stress hormones and the amygdala complex. In M.S. Gazanniga (Ed.), *The New Cognitive Neurosciences* (2nd ed., pp. 1081-1098). Cambridge, MA: MIT Press.
- McIntosh, D. N., Miller, L. J., McGrath, J., Shyu, V., Lampe, M., Taylor, A. K., ... & Hagerman, R. J. (1999b). Electrodermal responses to sensory stimuli in individuals with fragile X syndrome. *Am J Med Genet*, 83, 268-279. doi:10.1002/(SICI)1096-8628(19990402)83:4<268::AID-AJMG7>3.0.CO;2-K
- McIntosh, D. N., Miller, L. J., Shyu, V., & Dunn, W. (1999). Overview of the short sensory profile (SSP). *The sensory profile: Examiner's manual*, 59-73.
- McIntosh, D. N., Miller, L. J., Shyu, V., & Hagerman, R. J. (1999a). Sensory-modulation disruption, electrodermal responses, and functional behaviors. *Developmental Medicine & Child Neurology*, 41(09), 608-615.
- McKay, K. A. C., Buen, J. E., Bohan, K. J., & Maye, J. P. (2010). Determining the Relationship of Acute Stress, Anxiety, and Salivary α-Amylase Level With Performance of Student Nurse Anesthetists During Human-Based Anesthesia Simulator Training. *AANA Journal*, 78(4).
- Méndez-Bértolo, C., Moratti, S., Toledano, R., Lopez-Sosa, F., Martínez-Alvarez, R., Mah, Y. H., ... & Strange, B. A. (2016). A fast pathway for fear in human amygdala. *Nature neuroscience*, 19(8), 1041. doi:10.1038/nn.4324
- Mill, J., & Petronis, A. (2008). Pre- and peri-natal environmental risks for attention-deficit hyperactivity disorder (ADHD): the potential role of epigenetic processes in

- mediating susceptibility. *Journal of Child Psychology and Psychiatry*, 49(10), 1020-1030. doi:10.1111/j.1469-7610.2008.01909.x
- Miller, L. J., McIntosh, D. N., McGrath, J., Shyu, V., Lampe, M., Taylor, A. K., ... & Hagerman, R. J. (1999). Electrodermal responses to sensory stimuli in individuals with fragile X syndrome. *American Journal of Medical Genetics*, 83(4), 268-279. doi:10.1002/(SICI)1096-8628(19990402)83:4<268::AID-AJMG7>3.0.CO;2-K
- Miller, L. J., Nielsen, D. M., & Schoen, S. A. (2012). Attention deficit hyperactivity disorder and sensory modulation disorder: a comparison of behavior and physiology. *Research in Developmental Disabilities*, *33*(3), 804-818. doi:10.1016/j.ridd.2011.12.005
- Miller, L. J., Nielsen, D. M., Schoen, S. A., & Brett-Green, B. A. (2009). Perspectives on sensory processing disorder: a call for translational research. *Frontiers in Integrative Neuroscience*, *3*, 22. doi:10.3389/neuro.07.022.2009
- Miller, Lucy Jane, Coll, J. R., & Schoen, S. A. (2007). A randomized controlled pilot study of the effectiveness of occupational therapy for children with sensory modulation disorder. *The American Journal of Occupational Therapy*, 61(2), 228–238. doi:10.5014/ajot.61.2.228
- Miller, Lucy Jane, Nielsen, D. M., & Schoen, S. A. (2012). Attention deficit hyperactivity disorder and sensory modulation disorder: a comparison of behavior and physiology. *Research in Developmental Disabilities*, *33*(3), 804–818. doi:10.1016/j.ridd.2011.12.005
- Mims, M. C., & Olden, J. D. (2012). Life history theory predicts fish assemblage response to hydrologic regimes. *Ecology*, *93*(1), 35-45. doi:10.1890/11-0370.1
- Montano, N., Porta, A., Cogliati, C., Costantino, G., Tobaldini, E., Casali, K. R., & Iellamo, F. (2009). Heart rate variability explored in the frequency domain: a tool to investigate the link between heart and behavior. *Neuroscience & Biobehavioral Reviews*, 33(2), 71-80. doi:10.1016/j.neubiorev.2008.07.006
- Munoz-Lopez, M., Mohedano-Moriano, A., & Insausti, R. (2010). Anatomical pathways for auditory memory in primates. *Frontiers in Neuroanatomy*, *4*, 129. doi:10.3389/fnana.2010.00129

- Nagai, Y., Critchley, H. D., Featherstone, E., Trimble, M. R., & Dolan, R. J. (2004).

 Activity in ventromedial prefrontal cortex covaries with sympathetic skin conductance level: a physiological account of a "default mode" of brain function.

 Neurolmage, 22, 243-251. doi:10.1016/j.neuroimage.2004.01.019
- Nardocci, G., Navarro, C., Cortés, P. P., Imarai, M., Montoya, M., Valenzuela, B., ... & Fernández, R. (2014). Neuroendocrine mechanisms for immune system regulation during stress in fish. *Fish and Shellfish Immunology*, 40(2), 531-538. doi:10.1016/j.fsi.2014.08.001
- Nathan, R., Getz, W. M., Revilla, E., Holyoak, M., Kadmon, R., Saltz, D., & Smouse, P. E. (2008). A movement ecology paradigm for unifying organismal movement research. *Proceedings of the National Academy of Sciences*, *105*(49), 19052-19059. doi:10.1073/pnas.0800375105
- National Institute for Occupational Safety and Health (NIOSH). (1998). Criteria for a Recommended Standard: Occupational Noise Exposure. Revised Criteria 1998. http://www.cdc.gov/niosh/98-126.html. National Institute for Occupational Safety and Health, Cincinnati.
- Nelson, A., & Mooney, R. (2016). The basal forebrain and motor cortex provide convergent yet distinct movement-related inputs to the auditory cortex. *Neuron*, 90(3), 635-648. doi:10.1016/j.neuron.2016.03.031
- Nemenman, I. (2012). 4. Information theory and adaptation. *Quantitative Biology: From Molecular to Cellular Systems*, 73.
- Neto, E. S., Custaud, M. A., Cejka, J. C., Abry, P., Frutoso, J., Gharib, D., & Flandrin, P. (2004). Assessment of cardiovascular autonomic control by the empirical mode decomposition. *Methods of Information in Medicine*, *43*(1), 60-65.
- Nicholls, J. G., Martin, A. R., Wallace, B. G., & Fuchs, P. A. (2001). From neuron to brain (Vol. 271). Sunderland, MA: Sinauer Associates.
- O'Donnell, A., & Glasgow, B. (2011). The autonomic nervous system. *New Zealand Medical Student Journal*, (13).
- Oji-Mmuo, C. N., Gardner, F. C., & Doheny, K. K. (2018). Heightened sympathetic arousal is demonstrated by skin conductance responsivity to auditory stimuli in a

- small cohort of neonates with opiate withdrawal. *Brain Research Bulletin*, *138*, 106-111. doi:10.1016/j.brainresbull.2017.06.007
- Ooi, Y. P., Weng, S. J., Magiati, I., Ang, R. P., Goh, T. J., Fung, D. S., & Sung, M. (2016). Factors influencing agreement between parent and child reports of anxiety symptoms among children with high-functioning autism spectrum disorders.

 Journal of Developmental and Physical Disabilities, 28(3), 407-424.

 doi:10.1007/s10882-016-9481-5
- Orekhova, E. V., & Stroganova, T. A. (2014). Arousal and attention re-orienting in autism spectrum disorders: evidence from auditory event-related potentials. *Frontiers in Human Neuroscience*, 8, 34. doi:10.3389/fnhum.2014.00034
- Overfield, T. (2017). Biological variation in health and illness: race, age, and sex differences. CRC Press.
- Ozsivadjian, A., Knott, F., & Magiati, I. (2012). Parent and child perspectives on the nature of anxiety in children and young people with autism spectrum disorders: a focus group study. *Autism*, *16*(2), 107-121. doi:10.1177/1362361311431703
- Pagani, M., Lombardi, F., Guzzetti, S., Rimoldi, O., Furlan, R., Pizzinelli, P., . . . Malliani, A. (1986). Power spectral analysis of heart rate and arterial pressure variabilities as a marker of sympatho-vagal interaction in man and conscious dog. *Circulation Research*, 59, 178–193. doi:10.1161/01.RES.59.2.178
- Pagliaccio, D., Luby, J. L., Bogdan, R., Agrawal, A., Gaffrey, M. S., Belden, A. C., . . . Barch, D. M. (2015). HPA axis genetic variation, pubertal status, and sex interact to predict amygdala and hippocampus responses to negative emotional faces in school-age children. *NeuroImage*, 109, 1-11. doi:10.1016/j.neuroimage.2015.01.017
- Pagliaccio, D., Luby, J. L., Bogdan, R., Agrawal, A., Gaffrey, M. S., Belden, A. C., ...
 Barch, D. M. (2015). Amygdala Functional Connectivity, HPA Axis Genetic
 Variation, and Life Stress in Children and Relations to Anxiety and Emotion
 Regulation. *Journal of Abnormal Psychology*, 124(4), 817–833.
 doi:10.1037/abn0000094

- Parati, G., Mancia, G., Di Rienzo, M., & Castiglioni, P. (2006). Point: Counterpoint: Cardiovascular variability is/is not an index of autonomic control of circulation. *Journal of Applied Physiology*, 101(2), 676-682. doi:10.1152/japplphysiol.00446.2006
- Parush, S., Sharoni, C., Hahn-Markowitz, J., & Katz, N. (2000). Perceptual, motor and cognitive performance components of Bedouin children in Israel. *Occupational Therapy International*, 7(4), 216-231. doi:10.1002/oti.124
- Perakakis, P., Joffily, M., Taylor, M., Guerra, P., & Vila, J. (2010). KARDIA: A Matlab software for the analysis of cardiac interbeat intervals. *Computer Methods and Programs in Biomedicine*, *98*(1), 83-89. doi:10.1016/j.cmpb.2009.10.002
- Perdue, K. L., Edwards, L. A., Tager-Flusberg, H., & Nelson, C. A. (2017). Differing developmental trajectories in heart rate responses to speech stimuli in infants at high and low risk for autism spectrum disorder. *Journal of Autism and Developmental Disorders*, 47(8), 2434-2442. doi:10.1007/s10803-017-3167-4
- Perloff, H. S. (2015). The quality of the urban environment: essays on" new resources" in an urban age. New York, NY: Routledge.
- Perry, B. D., Pollard, R. A., Blakley, T. L., Baker, W. L., & Vigilante, D. (1995). Childhood trauma, the neurobiology of adaptation, and? use? dependent? development of the brain: How? states? become? traits?. *Infant Mental Health Journal*, 16(4), 271-291. doi:10.1002/1097-0355(199524)16:4<271::AID-IMHJ2280160404>3.0.CO;2-B
- Phelps, E. A., & LeDoux, J. E. (2005). Contributions of the amygdala to emotion processing: from animal models to human behavior. *Neuron*, 48(2), 175-187. doi:10.1016/j.neuron.2005.09.025
- Phillips, D. P., Hall, S. E., & Boehnke, S. E. (2002). Central auditory onset responses, and temporal asymmetries in auditory perception. *Hearing Research*, 167(1-2), 192-205. doi:10.1016/S0378-5955(02)00393-3
- Pichon, A., Roulaud, M., Antoine-Jonville, S., de Bisschop, C., & Denjean, A. (2006).

 Spectral analysis of heart rate variability: interchangeability between autoregressive

- analysis and fast Fourier transform. *Journal of Electrocardiology*, *39*(1), 31-37. doi:10.1016/j.jelectrocard.2005.08.001
- Pillai, K. C. S. (2004). Multivariate analysis of variance (MANOVA). *Encyclopedia of Statistical Sciences*, 8.
- Plews, D. J., Scott, B., Altini, M., Wood, M., Kilding, A. E., & Laursen, P. B. (2017). Comparison of Heart-Rate-Variability Recording With Smartphone Photoplethysmography, Polar H7 Chest Strap, and Electrocardiography. *International Journal of Sports Physiology and Performance*, 12(10), 1324-1328. doi:10.1123/ijspp.2016-0668
- Poliakova, N., Dionne, G., Dubreuil, E., Ditto, B., Pihl, R. O., Pérusse, D., ... & Boivin, M. (2014). A methodological comparison of the Porges algorithm, fast Fourier transform, and autoregressive spectral analysis for the estimation of heart rate variability in 5-month-old infants. *Psychophysiology*, *51*(6), 579-583. doi:10.1111/psyp.12194
- Porges, S. W. (1995). Cardiac vagal tone: A physiological index of stress. Cardiac vagal tone: A physiological index of stress. *Neuroscience & Biobehavioral Reviews*, 19(2), 225-233. doi:10.1016/0149-7634(94)00066-A
- Porges, S. W. (1995). Orienting in a defensive world: Mammalian modifications of our evolutionary heritage. A Polyvagal Theory. *Psychophysiology*, *32*(4), 301–318. doi:10.1111/j.1469-8986.1995.tb01213.x
- Porges, S. W., & Byrne, E. A. (1992). Research methods for measurement of heart rate and respiration. *Biological Psychology*, *34*(2), 93-130. doi:10.1016/0301-0511(92)90012-J
- Porges, S. W., & Furman, S. A. (2011). The early development of the autonomic nervous system provides a neural platform for social behaviour: A polyvagal perspective. *Infant and Child Development*, 20(1), 106–118. doi:10.1002/icd.688
- Porter, M. B. (2011). *The bellhop manual and user's guide: Preliminary draft*. Heat, Light, and Sound Research, Inc., La Jolla, CA, USA, Tech. Rep.
- Powledge, T. M. (2011). Behavioral epigenetics: how nurture shapes nature. *BioScience*, 61(8), 588-592. doi:10.1525/bio.2011.61.8.4

- Purves, D., Cabeza, R., Huettel, S. A., LaBar, K. S., Platt, M. L., Woldorff, M. G., & Brannon, E. M. (2008). *Cognitive Neuroscience*. Sunderland: Sinauer Associates, Inc.
- Randall, D. C., Brown, D. R., Raisch, R. M., Yingling, J. D., & Randall, W. C. (1991). SA nodal parasympathectomy delineates autonomic control of heart rate power spectrum. *American Journal of Physiology-Heart and Circulatory Physiology*, 260(3), H985-H988. doi:10.1152/ajpheart.1991.260.3.H985
- Rasia-Filho, A. A., Londero, R. G., & Achaval, M. (2000). Functional activities of the amygdala: an overview. *Journal of Psychiatry and Neuroscience*, 25(1), 14.
- Read, S., & Grundy, E. (2012). Allostatic load—a challenge to measure multisystem physiological dysregulation. *Pathways Node at NCRM*. Available at http://eprints.ncrm.ac.uk/2879/
- Reyes del Paso, G. A., Langewitz, W., Mulder, L. J., Roon, A., & Duschek, S. (2013). The utility of low frequency heart rate variability as an index of sympathetic cardiac tone: a review with emphasis on a reanalysis of previous studies. *Psychophysiology*, 50(5), 477-487. doi:10.1111/psyp.12027
- Roberts, S., Keers, R., Lester, K. J., Coleman, J. R. I., Breen, G., Arendt, K., ... Wong, C.
 C. Y. (2015). HPA axis related genes and response to psychological therapies:
 genetics and epigenetics. *Depression and Anxiety*, 32(12), 861–870.
 doi:10.1002/da.22430
- Robinson-Wood, T. (2016). *The convergence of race, ethnicity, and gender: Multiple identities in counseling*. SAGE Publications.
- Roth, W. T., Dawson, M. E., & Filion, D. L. (2012). Publication recommendations for electrodermal measurements. *Psychophysiology*, *49*(8), 1017-1034. doi:10.1111/j.1469-8986.2012.01384.x
- Royeen, C., & Mu, K. (2003). Stability of tactile defensiveness across cultures: European and American children's responses to the Touch Inventory for Elementary School Aged Children (TIE). *Occupational Therapy International*, *10*(3), 165-174. doi:10.1002/oti.183

- Rutkowski, R. G., & Weinberger, N. M. (2005). Encoding of learned importance of sound by magnitude of representational area in primary auditory cortex. *Proceedings of the National Academy of Sciences of the United States of America*, 102(38), 13664-13669. doi:10.1073/pnas.0506838102
- Rutter, M., Pickles, A., Murray, R., & Eaves, L. (2001). Testing hypotheses on specific environmental causal effects on behavior. *Psychological Bulletin*, *127*(3), 291–324. doi:10.1037//0033-2909.127.3.291
- Salazar, C. R., Strizich, G., Seeman, T. E., Isasi, C. R., Gallo, L. C., Avilés-Santa, L. M., ... & Lipton, R. B. (2016). Nativity differences in allostatic load by age, sex, and Hispanic background from the Hispanic Community Health Study/Study of Latinos. SSM-Population Health, 2, 416-424. doi:10.1016/j.ssmph.2016.05.003
- Salomon, K., Matthews, K. A., & Allen, M. T. (2000). Patterns of sympathetic and parasympathetic reactivity in a sample of children and adolescents. *Psychophysiology*, *37*(6), 842-849.
- Sammito, S., Thielmann, B., Seibt, R., Klussmann, A., Weippert, M., & Böckelmann, I. (2015). Guideline for the application of heart rate and heart rate variability in occupational medicine and occupational science. *ASU Int*. doi:10.17147/ASUI.2015-06-09-03
- Schaaf, R. C., Benevides, T. W., Blanche, E., Brett-Green, B. A., Burke, J., Cohn, E., ... & Parham, D. (2010). Parasympathetic functions in children with sensory processing disorder. *Frontiers in Integrative Neuroscience*, 4, 4. doi:10.3389/fnint.2010.00004
- Schaaf, R. C., Benevides, T. W., Leiby, B. E., & Sendecki, J. A. (2015). Autonomic dysregulation during sensory stimulation in children with autism spectrum disorder. *Journal of Autism and Developmental Disorders*, 45(2), 461-472. doi: 10.1007/s10803-013-1924-6
- Schaaf, R. C., Miller, L. J., Seawell, D., & O'Keefe, S. (2003). Children with disturbances in sensory processing: A pilot study examining the role of the parasympathetic nervous system. American Journal of Occupational Therapy, *57*(4), 442-449. doi:10.5014/ajot.57.4.442

- Schauder, K. B., & Bennetto, L. (2016). Toward an Interdisciplinary understanding of sensory dysfunction in autism spectrum disorder: An integration of the neural and symptom literatures. *Frontiers in Neuroscience*, 10. doi:10.3389/fnins.2016.00268
- Schlichting, C. D., & Wund, M. A. (2014). Phenotypic plasticity and epigenetic marking: an assessment of evidence for genetic accommodation. *Evolution*, 68(3), 656-672. doi:10.1111/evo.12348
- Schlotz, W., Kumsta, R., Layes, I., Entringer, S., Jones, A., & Wüst, S. (2008). Covariance between psychological and endocrine responses to pharmacological challenge and psychosocial stress: a question of timing. *Psychosomatic Medicine*, 70(7), 787-796. doi:10.1097/PSY.0b013e3181810658
- Schmitz, J., Krämer, M., Tuschen-Caffier, B., Heinrichs, N., & Blechert, J. (2011).

 Restricted autonomic flexibility in children with social phobia. *Journal of Child Psychology and Psychiatry*, 52(11), 1203-1211. doi:10.1111/j.1469-7610.2011.02417.x
- Schoen, S. A., Miller, L. J., Brett-Green, B. A., & Nielsen, D. M. (2009). Physiological and behavioral differences in sensory processing: a comparison of children with autism spectrum disorder and sensory modulation disorder. *Frontiers in Integrative Neuroscience*, *3*, 29. doi:10.3389/neuro.07.029.2009
- Schoen, S. A., Miller, L. J., Brett-Green, B., Reynolds, S., & Lane, S. J. (2008). Arousal and reactivity in children with sensory processing disorder and autism specturm disorder. *Psychophysiology*, 45, pp. S102-S102.
- Seidel, E.-M., Pfabigan, D. M., Keckeis, K., Wucherer, A. M., Jahn, T., Lamm, C., & Derntl, B. (2013). Empathic competencies in violent offenders. *Psychiatry Research*, 210(3), 1168–1175. doi:10.1016/j.psychres.2013.08.027
- Self, D. (2012). Audio engineering explained. Focal Press.
- Shields, S. A., MacDowell, K. A., Fairchild, S. B., & Campbell, M. L. (1987). Is mediation of sweating cholinergic, adrenergic, or both? A comment on the literature.

 Psychophysiology, 24(3), 312-319. doi:10.1111/j.1469-8986.1987.tb00301.x*
- Shim, S. H., Park, S. Y., Moon, S. N., Oh, J. H., Lee, J. Y., Kim, H. H., ... & Lee, S. J. (2014). Baseline heart rate variability in children and adolescents with vasovagal

- syncope. *Korean Journal of Pediatrics*, *57*(4), 193-198. doi:10.3345/kjp.2014.57.4.193
- Shimada, M., Ishii, Y., & Shibao, H. (2010). Rapid adaptation: a new dimension for evolutionary perspectives in ecology. *Population Ecology*, *52*(1), 5-14. doi:10.1007/s10144-009-0187-8
- Silva, L. M., & Schalock, M. (2012). Sense and self-regulation checklist, a measure of comorbid autism symptoms: initial psychometric evidence. *American Journal of Occupational Therapy*, 66(2), 177-186. doi:10.5014/ajot.2012.001578
- Sokhadze, E. M., Casanova, M. F., Tasman, A., & Brockett, S. (2016).

 Electrophysiological and behavioral outcomes of Berard Auditory Integration

 Training (AIT) in children with autism spectrum disorder. *Applied Psychophysiology and Biofeedback, 41*(4), 405-420. doi:10.1007/s10484-016-9343-
- Staib, M., Castegnetti, G., & Bach, D. R. (2015). Optimising a model-based approach to inferring fear learning from skin conductance responses. *Journal of Neuroscience Methods*, 255, 131–138. doi:10.1016/j.jneumeth.2015.08.009
- Sterling, P. (2004). Principles of allostasis: optimal design, predictive regulation, pathophysiology and rational therapeutics. In: Schulkin, J. (Ed.), Allostasis, Homeostasis, and the Costs of Adaptation. Cambridge University Press, Cambridge.
- Sterling, P., & Eyer, J. (1988). Allostasis: a new paradigm to explain arousal pathology. In S. Fisher & J. Reason (Eds.), *Handbook of life stress, cognition and health* (pp. 629-649). Oxford, England: John Wiley & Sons.
- Strickland, B. R. (2001). The Gale encyclopedia of psychology. Gale.
- Stuffrein-Roberts, S., Joyce, P. R., & Kennedy, M. A. (2008). Role of epigenetics in mental disorders. *Australian & New Zealand Journal of Psychiatry*, 42(2), 97-107. doi:10.1080/00048670701787495
- Super, C. M., & Harkness, S. (2002). Culture structures the environment for development. *Human Development*, 45(4), 270-274. doi:10.1159/000064988
- Task Force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology. (1996). Heart rate variability. Standards of

- measurement, physiological interpretation, and clinical use. *European Heart Journal*, 17(3), 354–81. doi:10.1161/01.CIR.93.5.1043
- Taylor, J. A., & Studinger, P. (2006). Counterpoint: cardiovascular variability is not an index of autonomic control of the circulation. *Journal of Applied Physiology*, 101(2), 678.
- Templer, D. I. (2008). Correlational and factor analytic support for Rushton's differential K life history theory. *Personality and Individual Differences*, *45*(6), 440-444. doi:10.1016/j.paid.2008.05.010
- Thayer, J. F., Åhs, F., Fredrikson, M., Sollers III, J. J., & Wager, T. D. (2012). A meta-analysis of heart rate variability and neuroimaging studies: implications for heart rate variability as a marker of stress and health. *Neuroscience & Biobehavioral Reviews*, 36(2), 747-756. doi: 10.1016/j.neubiorev.2011.11.009
- Thayer, J. F., & Lane, R. D. (2007). The role of vagal function in the risk for cardiovascular disease and mortality. *Biological Psychology*, 74(2), 224-242. doi:10.1016/j.biopsycho.2005.11.013
- Tirosh, E., Bettesh Bendrian, S., Golan, G., Tamir, A., & Cohen Dar, M. (2003).

 Regulatory disorders in Israeli infants: epidemiologic perspective. *Journal of Child Neurology*, *18*(11), 748–754. doi:10.1177/08830738030180110901
- Tobaldini, E., Nobili, L., Strada, S., Casali, K. R., Braghiroli, A., & Montano, N. (2013). Heart rate variability in normal and pathological sleep. *Frontiers in Physiology*, *4* (294). doi:10.3389/fphys.2013.00294
- Tomchek, S. D., & Dunn, W. (2007). Sensory processing in children with and without autism: a comparative study using the short sensory profile. *American Journal of Occupational Therapy*, 61(2), 190-200. doi: 10.5014/ajot.61.2.190
- Toninelli, G., Vigo, C., Vaglio, M., Porta, A., Lucini, D., Badilini, F., ... & CTNV, L. (2012). DynaScope: a software tool for the analysis of heart rate variability during exercise. *Comput Cardiol*, *39*, 181-184.
- Turpin, G. (1986). Effects of stimulus intensity on autonomic responding: The problem of differentiating orienting and defense reflexes. *Psychophysiology*, 23(1), 1-14. doi:10.1111/j.1469-8986.1986.tb00583.x

- Ulrich, R. S., Simons, R. F., Losito, B. D., Fiorito, E., Miles, M. A., & Zelson, M. (1991). Stress recovery during exposure to natural and urban environments. *Journal of Environmental Psychology*, 11(3), 201-230. doi:10.1016/S0272-4944(05)80184-7
- van den Berg, M., Wendel-Vos, W., van Poppel, M., Kemper, H., van Mechelen, W., & Maas, J. (2015). Health benefits of green spaces in the living environment: A systematic review of epidemiological studies. Urban Forestry & Urban Greening, 14(4), 806-816. doi:10.1016/j.ufug.2015.07.008
- Von Borell, E., Langbein, J., Després, G., Hansen, S., Leterrier, C., Marchant-Forde, J., ... & Veissier, I. (2007). Heart rate variability as a measure of autonomic regulation of cardiac activity for assessing stress and welfare in farm animals—a review. *Physiology & Behavior*, 92(3), 293-316. doi:10.1016/j.physbeh.2007.01.007
- W. Dunn, Sensory Profile, Psychological Corporation, San Antonio, Tex, USA, 1999.
- Weaver, L. C., & Polosa, C. (Eds.). (2006). *Autonomic dysfunction after spinal cord injury* (Vol. 152). Amsterdam: Elsevier.
- Weinberger, N. M. (2015). New perspectives on the auditory cortex: learning and memory. In Handbook of clinical neurology (Vol. 129, pp. 117-147). Elsevier.
- White, D. W., & Raven, P. B. (2014). Autonomic neural control of heart rate during dynamic exercise: revisited. *The Journal of Physiology*, 592(12), 2491-2500. doi:10.1113/jphysiol.2014.271858
- Widmann, A., Schröger, E., & Wetzel, N. (2018). Emotion lies in the eye of the listener: emotional arousal to novel sounds is reflected in the sympathetic contribution to the pupil dilation response and the P3. *Biological Psychology*, *133*, 10-17. doi:10.1016/j.biopsycho.2018.01.010
- Williams, D. P., Jarczok, M. N., Ellis, R. J., Hillecke, T. K., Thayer, J. F., & Koenig, J. (2017). Two-week test–retest reliability of the Polar® RS800CX[™] to record heart rate variability. *Clinical Physiology and Functional Imaging*, *37*(6), 776-781. doi:doi.org/10.1111/cpf.12321
- Winer, J. A., & Lee, C. C. (2007). The distributed auditory cortex. *Hearing Research*, 229(1-2), 3-13. doi:10.1016/j.heares.2007.01.017

- Withagen, R., & van Wermeskerken, M. (2010). The role of affordances in the evolutionary process reconsidered: A niche construction perspective. *Theory & Psychology*, 20(4), 489-510. doi:10.1177/0959354310361405
- Wolf, M., Van Doorn, G. S., Leimar, O., & Weissing, F. J. (2007). Life-history trade-offs favour the evolution of animal personalities. *Nature*, *447*(7144), 581-584. doi:10.1038/nature05835
- Woodard, C. R., Goodwin, M. S., Zelazo, P. R., Aube, D., Scrimgeour, M., Ostholthoff, T., & Brickley, M. (2012). A comparison of autonomic, behavioral, and parent-report measures of sensory sensitivity in young children with autism. *Research in Autism Spectrum Disorders*, 6(3), 1234–1246. doi:10.1016/j.rasd.2012.03.012
- Yung, T. W. K., Lai, C. Y., Gomez, I. N., Loh, V., Wang, J., Chan, J. Y., ... & Ng, S. S. (2017). Parents' perceptions of children's executive functions across different cities. Child Neuropsychology, 1-10. doi:10.1080/09297049.2017.1406075
- Zhang, T. Y., & Meaney, M. J. (2010). Epigenetics and the environmental regulation of the genome and its function. *Annual Review of Psychology*, *61*, 439-466. doi:10.1146/annurev.psych.60.110707.163625
- Zimmerman, E., & Woolf, S. H. (2014). *Understanding the relationship between education and health*. National Academy of Sciences.