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DESIGN OF LANTHANIDE SUPRAMOLECULAR

ARCHITECTURES

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PhD

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Design of Lanthanide Supramolecular Architectures

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A thesis submitted in partial fulfilment of the requirements for

the degree of Doctor of Philosophy

August 2021

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Abstract

Self-assembly of supramolecular architectures have been widely used in catalysis and luminescent sensing. By contrast to transition metals, lanthanide selfassembled supramolecular complexes exhibit unique chemical and physical properties which have shown promising application in luminescent sensing and diagnostic applications. In Chapter 1, the background of supramolecular chemistry and lanthanides are briefly introduced. Since the applications of these lanthanide self assembled systems are all structure-dependent, this thesis aims to develop different lanthanide supramolecular compounds from small assembled units by targeting changes in ligand design and reaction conditions. The other goal is to create chiral functional materials to explore and develop the area of chiroptic by looking at the effect of chirality in structure and correlating to its optical properties and function. In this aspect, such assemblies require the control of stereoselectivity of the lanthanide supramolecular architectures. However, controlling the stereoselectivity of lanthanide complexes is rather challenging due to the kinetically labile properties and poor stereochemical preferences. In Chapter 2- point chirality as well as the linker/spacer effect will be explored.

In Chapter 2a, two pairs of linear biphenyl-linked chiral bis-tridentate ligands were prepared and the point chirality effect of four resulting lanthanide supramolecular bimetallic helicates were studied. By extending the point chirality from the two metal centers, the ability of chirality transfer was shown to be greatly affected and diastereoselective and nondiastereoselective formation of bimetallic triple helicates were observed.

In Chapter 2b, two *C*₂-symmetric bis-tridentate ligands were designed based on symmetry principles where the effect of the length of the spacer towards selfassembly process was studied. Results showed that the length of the spacer greatly affected the supramolecular formation even if the offset distance of two metal chelating units remained unchanged. Higher-ordered supramolecular tetrahedron was then prepared based on the small helicate units, which resulted in an helicate-tocage transformation. In this system, it was found that the stability of the tetrahedron was dependent on the size of the ionic radii. Hence, this study showed the importance of ionic radii and ligand design towards supramolecular transformation. Lanthanide heterometallic complexes are important for engineering optical material and is promising for creating more versatile multifunctional properties as well as improving up-conversion efficiency and enhancing optical properties. However, preparation of lanthanide heterometallic complexes is extremely difficult owing to the similar ionic radii causing issues in controlled selectively.

In Chapter 3, attempt to prepare lanthanide heterometallic tetrahedron by a simple crystallization method was reported. In this work, it was found that upon crystallization with a mixture of two different lanthanide metal helicates with very similar ionic radii, e.g. one helicate strand of Eu and Ln (Ln = Sm /Gd), a mixture of five different lanthanide tetrahedron was formed as evidenced by ESI-HRMS. The percentage amount of each tetrahedron was estimated by MS deconvolution. It was

evidenced, that the self-assembly process of the tetrahedron deviated from the statistical result. The helicate-to-tetrahedron mechanism was also investigated in this chapter.

In Chapter 4, focus here is channeled to looking at point chirality and the chiral domino effect in a lanthanide system. Hence different types of small building blocks based on amino acids were used to investigate and explore the area. The construction of enantiomeric pure lanthanide supramolecular complexes strongly relies on point chirality induced by a chiral group with the strongest point chirality effect. As a result, the ligand design is restricted by the use of a specific chiral group and the distance between the point chirality and metal centers. A series of ligands which was coupled to different types of amino acid were designed and synthesized. The design principal here is based on a chiral group being attached to the C-terminal of a short peptide which is expected to transfer chiral information to the metal center via the chiral domino effect.

Publications

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- Wong, H.-Y.; Lo, W.-S.; <u>Yim, K.-H.</u>; Law, G.-L., Chirality and Chiroptics of Lanthanide Molecular and Supramolecular Assemblies. *Chem* **2019**, *5*, 3058-3095.
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List of Abbreviations

a.u.	Arbitrary unit		
Вос	Tert-butoxycarbonyl protecting group		
CHCl₃	Chloroform		
CPL	Circularly polarized luminescence		
DCM	Dichloromethane		
DMAP	4-Dimethylaminopyridine		
DMF	Dimethylformamide		
DMSO	Dimethyl sulfoxide		
DNA	Deoxyribonucleic Acid		
EA	Ethyl Acetate		
EDCI	1-Ethyl-3-(3-dimethylaminopropyl)-carbodiimide		
em	Emission		
Equiv.	Equivalent		
ESI	Electrospray ionization		
ex	Excitation		
HPLC	High Performance Liquid Chromatography		
HRMS	High-resolution mass spectroscopy		
Ln	Lanthanide		
m/z	Mass-to-charge ratio		
MeCN	Acetonitrile		

MeOH	Methanol		
mM	Millimolar (10 ⁻³ M)		
ms	Millisecond (10 ⁻³ s)		
NMR	Nuclear magnetic resonance		
THF	Tetrahydrofuran		
TMS	Trimethylsilyl		
UV/Vis	Ultraviolet/Visible		
μΜ	Micromolar (10 ⁻⁶ M)		

Chapter 1: Introduction to lanthanide and supramolecular coordination chemistry

1.1 Lanthanide

1.1.1 Background

Lanthanide, also known as "rare earth metal", are a group of elements located between the third-row elements barium and hafnium. The series of lanthanide consist of 15 elements from lanthanum to lutetium with atomic numbers from 57 to 71. The history of lanthanide chemistry started in 1794. A Finnish scientist called Johann Gadolin successfully isolated an 'earth' oxide from a black mineral known as gadolinite. Lanthanide ions exhibit a number of interesting characteristics that are different from those of d-block transition metals. For example, lanthanide ions show a very wide range of coordination numbers, small crystal field splitting as well as unique photophysical and magnetic properties.¹ Therefore, lanthanide ions are considered to be a promising candidate for developing more efficient magnetic and luminescent materials.

1.1.2 Electronic properties

The unique photophysical and magnetic properties of lanthanide ions can be attributed to their electronic configurations, in which the 4f orbitals are gradually filled across the series. Their electronic configurations can be described as [Xe]4fⁿ, where n =0-14 (table 1-1).^{2,3} Compared with outer filled 5s²5p⁶ orbitals, 4f sub-shell penetrates

the xenon core appreciably.⁴ Therefore, the 4f orbital cannot overlap with ligand orbitals effectively and they do not involve significantly in bonding. When lanthanide ions coordinate with a ligand, perturbation by the ligand to the lanthanide ions will be insignificant.³ As a result, the optical spectra as well as the magnetic properties of the lanthanide complexes are generally not affected by the coordination environment.

	Atom	Ln ²⁺	Ln ³⁺	Ln ⁴⁺
La	[Xe]5d ¹ 6s ²		[Xe]	
Ce	[Xe]4f ¹ 5d ¹ 6s ²		[Xe]4f ¹	[Xe]
Pr	[Xe]4f ³ 6s ²		[Xe]4f²	[Xe]4f ¹
Nd	[Xe]4f ⁴ 6s ²	[Xe]4f ⁴	[Xe]4f ³	[Xe]4f ²
Pm	[Xe]4f ⁵ 6s ²		[Xe]4f ⁴	
Sm	[Xe]4f ⁶ 6s ²	[Xe]4f ⁶	[Xe]4f ⁵	
Eu	[Xe]4f ⁷ 6s ²	[Xe]4f ⁷	[Xe]4f ⁶	
Gd	[Xe]4f ⁷ 5d ¹ 6s ²		[Xe]4f ⁷	
Tb	[Xe]4f ⁹ 6s ²		[Xe]4f ⁸	[Xe]4f ⁷
Dy	[Xe]4f ¹⁰ 6s ²	[Xe]4f ¹⁰	[Xe]4f ⁹	[Xe]4f ⁸
Но	[Xe]4f ¹¹ 6s ²		[Xe]4f ¹⁰	
Er	[Xe]4f ¹² 6s ²		[Xe]4f ¹¹	
Tm	[Xe]4f ¹³ 6s ²	[Xe]4f ¹³	[Xe]4f ¹²	
Yb	[Xe]4f ¹⁴ 6s ²	[Xe]4f ¹⁴	[Xe]4f ¹³	
Lu	[Xe]4f ¹⁴ 5d ¹ 6s ²		[Xe]4f ¹⁴	

 Table 1-1. Electronic configurations of lanthanides and their common ions.³

4f sub-shell is located closely to the nuclear core which leads to lanthanide contraction. The term "contraction" describes a greater-then-expect decrease in ionic radius of the elements.⁵ The effect of lanthanide contraction increases with the increasing atomic number due to the positively charged nucleus would have a stronger attraction to the 4f electrons across the period (figure 1-1).

	La	Ce	Pr	Nd	Pm	Sm	Eu	Gd	Tb	Dy	Ho	Er	Tm	Уb	Lu
Atomic radii (Å)	1.877	1.825	1.828	1.821	1.810	1.802	2.042	1.802	1.782	1.773	1.766	1.757	1.746	1.940	1.734
	La ³⁺	Ce ^{3*}	Pr ³⁺	Nd ³⁺	Pm ³⁺	5m ³⁺	Eu ³⁺	Gd ³⁺	Tb ³⁺	Dy³⁺	Ho ³⁺	Er ³⁺	Tm ³⁺	Yb³⁺	Lu ³⁺
Ionic radii (Å, CN = 9)	1.216	1.196	1.179	1.163	1.144	1.132	1.120	1.107	1.095	1.083	1.072	1.062	1.052	1.042	1.032

Figure 1-1. Atomic radii and ionic radii of lanthanide at +3 oxidation state (coordination number = 9).⁶

<u>1.1.3 Lanthanide coordination chemistry</u>

Traditionally, the coordination chemistry of lanthanide ions is considered to be purely electrostatic interaction. However, recent research has demonstrated that lanthanide ions also participate in covalent bonding with ligand.⁷ With the use of a combination of ligand (Cl) K-edge X-ray absorption spectroscopy (XAS) and timedependent density functional theory (TDDFT), the covalency in $LnCl_6^{3-}$ (Ln = Ce, Nd, Sm, Eu and Gd) was investigated. Result showed that there was an insignificant amount of mixing of 3p orbital with 4f orbital.

Even if lanthanide ions do participate in covalent bonding, lanthanide ions are still considered to be hard Lewis acids because of their large ionic radii as well as high charge density, with the most stable +3 oxidation state.^{8, 9} Lanthanide ions can accommodate a large coordination number in the range of 6 to 12, with a predominance for coordination number 9 for earlier lanthanide ions (La³⁺ to Eu³⁺), and 8 for later lanthanide ions (Dy³⁺ to Lu³⁺).^{4, 10} The coordination number of a lanthanide ion is determined by either first order- or second order effect.³ For first order effect, the saturation of lanthanide ion is determined by the number of ligand that pack around the metal centre. For second order effect, the use of bulky ligands may cause crowding around metal centre and the coordination number is governed by the steric effects generated by the ligands. Due to the second order effect, the coordination number of some exceptional lanthanide system can be as small as 3 to 4 when bulky ligands, such as bis(isopropyl)amide, are used.¹¹ The coordination number also affect the coordination geometry (table 1-2). Lanthanide ions preferably interact with hard Lewis bases in the order of O>N>S.^{12, 13} Typical ligands for lanthanide complexation include aminocaboxylates and β -diketonates, which are anionic ligands that can maximize the strength of interaction with lanthanide ions.

Coordination number	Geometry	Examples
2	Bent	[Yb{C(SiMe ₃)}]
2	Pyramidal (solid)	La[N(SiMe)]
3	Planar (gas and solution)	La[14(5)141C ₃ / ₂] ₃ ,
4	Tetrahedral	Yb[(N(SiMe ₃) ₂] ₃ (Ph ₃ PO)
5	Trigonal bipyramidal	$[Nd{P(SiMe_3)_2}_3(THF)_2]$
6	Trigonal prismatic	$[Ln{S_2P(cyclohexyl)_2}_3].$
7	Capped trigonal prismatic	Ba ₂ EuCl ₇
7	Pentagonal bipyramidal	[Yb(C ₆ F ₅) ₂ (THF) ₅].
0	Dodecahedral	Et ₄ N [Eu(S ₂ CNEt ₂) ₄],
8	Square antiprismatic	Ph ₄ P [Pr(S ₂ PMe ₂) ₄].
9	Tricapped trigonal prismatic	[Ln(H ₂ O) ₉] ³⁺

Table 1-2. Examples of coordination numbers and geometries of lanthanide complexes.³

In view of complexation, lanthanide ions are labile in solution state and the ligands can be exchanged easily in the presence of water and methanol. Lanthanide ions highly prefer a saturation of the inner coordination sphere. Therefore, careful designing of ligand should be taken into consideration. Generally, a ligand should consist of an adequate number of donor atoms as well as an appropriate size of binding site because lanthanide tends to be saturated with large coordination number. Otherwise, competition for lanthanide ions by solvent molecules and ligands will affect the photophysical properties of the complexes.^{14, 15}

1.2 Lanthanide photophysical properties

1.2.1 Luminescence of lanthanide ions

One of the most attractive properties of lanthanide is their unique luminescence properties.^{14, 16} Molecular luminescence can be classified into either fluorescence or phosphorescence according to the IUPAC rule.¹⁷ However, for lanthanide systems, some transitions involve fluorescence, which is a process that occurs without changing the spin state of the excited electron (e.g. Yb³⁺, ${}^{2}F_{5/2} \rightarrow {}^{2}F_{7/2}$), while some transitions involve phosphorescence (e.g. Eu, ${}^{5}D_{0} \rightarrow {}^{7}F_{J}$).¹⁸ Therefore, emission of lanthanide ions is commonly referred to luminescence to avoid any confusion.

The photophysical properties of lanthanide ions can be attributed to their electronic configuration. As the 4*f* electrons are well shielded by 5*s* and 5*p* orbitals, ligand perturbations are minimized and the 4*f* orbitals are thus well hidden from the external environment. Upon excitation by electromagnetic radiation, lanthanide ions give rise to characteristic needle-like emission spectra (figure 1-2 and 1-3).^{19, 20} Theoretically, most lanthanide (III) ions are luminescent but some of them are more emissive than others.

5



Figure 1-2. Energy diagram of the lanthanide ions. Main luminescent levels are shown in red.¹⁹



Figure 1-3. Luminescence spectra of lanthanide complexes with narrow-like emission bands.²⁰

Lanthanide ions exhibit three different types of electronic transitions, which are 4*f*-4*f* transitions, 4*f*-5*f* transitions and charge transfer transitions. The 4*f*-4*f* transitions are the easiest to recognize as they are quite narrow and do not depend much on the chemical environment of the lanthanide ions. However, 4*f*-4*f* transitions are forbidden by the Laporte rule, which states that ΔI must be ± 1. 4*f*-4*f* transitions can only be observed when there is a change in 4*f* wavefunction by vibronic coupling or mixing of parity wavefunction with 5*d* orbitals.²¹ The Laporte forbidden 4*f*-4*f* transitions lead to small extinction coefficients (smaller than 10 M^{-1} cm⁻¹).^{10, 11, 22}

For 4*d*-5*f* transitions, they are allowed and has a relatively larger emission intensity compared with those of 4*f*-4*f* transitions. However, 4*f*-5*d* transitions are highly dependent on ligand field effect on the d-orbitals as the 5*d* orbitals interact directly with the ligands. The 4*f*-5*d* transitions generally require high energy and only Ce³⁺, Pr³⁺ and Tb³⁺ can be observed.²³ Similar to 4*f*-5*d* transitions, charge transfer transitions are also Laporte allowed.²⁴ However, they requires high energy which can be only observed for Sm³⁺, Eu³⁺, Tm³⁺ and Yb³⁺.

1.2.2 Antenna effect

Since 4f-4f transitions are Laporte forbidden and 4f-5d transitions are uncommon, excitation of lanthanide ions to 4f-4f excited state remains a challenge. This problem can be solved by both direct and indirect method. In the direct method, lanthanide ions are excited using laser, but this method is not practical for biological application. The indirect method, which is called sensitization or antenna effect,²⁵ is a more common practice for sensitizing lanthanide ions. This method involves the coordination of a chromophore to the lanthanide ions. The chromophore will first absorb the electromagnetic radiation and then transfer the energy to the excited state of the lanthanide ions and decay through radiative 4f-4f transitions. The energy can be transferred from the ligand triplet excited state to the metal excited state, or from the ligand singlet excited state to the metal excited state (figure 1-4).^{21, 26, 27}



Figure 1-4. A modified Jablonski diagram showing the energy transfer from ligand to lanthanide ion (ISC = intersystem crossing; ET = energy transfer, IC = internal conversion).¹⁹

1.2.3 Dexter and Förster mechanism

Dexter and Förster mechanisms are two mechanisms that are commonly accepted to explain the antenna effect from the ligand triplet state to the excited state of lanthanide ions. Dexter mechanism describes the antenna effect as a double electron exchange between donor and acceptor through physical contact (figure 1-5a).²⁸ An electron is transferred from the ligand triple state to the excited state of lanthanide ions, while another electron is transferred from the highest occupied energy orbital of the lanthanide ions to the ligand. This process requires a short distance to facilitate effective overlapping of wavefunction between the ligand donor and the metal acceptor.

Förster mechanism states that energy is transferred from a ligand donor to a metal acceptor through diploe-diploe coupling (figure 1-5b).^{28, 29} The donor is first

excited and then relaxes to release energy. The energy is then absorbed by the lanthanide ions.



Figure 1-5. Scheme of (a) Dexter mechanism and (b) Förster mechanism (D = energy donor; A = energy acceptor).²⁸

1.3 Spectroscopic techniques for characterization of lanthanide

complexes

<u>1.3.1 UV-visible spectroscopy</u>

The interaction between photons and a chemical compound can be investigated by UV-visible spectroscopy. When light passes through a cuvette containing a sample, the compound will absorb the photon at a specific wavelength of light. Energy will be transferred from the photon to an electron and the electron may be promoted to a higher energy level. The wavelength of light that is required for such a promotion depends on the energy difference between two states. The relationship between the energy difference and the wavelength of light follows Planck's equation:

$$E = hv = hc/\lambda$$
 (Eq. 1)

The extinction coefficient provides an indication of the intensity of an absorption transition, which can be determined by Beer's law:

$$log(I_0/I) = A = \varepsilon cl$$
 (Eq. 2)

Where A is absorbance, I_0 is the incident light intensity, I is the transmitted light intensity, ε is the extinction coefficient, c is the concentration of the sample and l is the path length of the cuvette.

1.3.2 Emission and quantum yield

Emission happens when a molecule absorbs a photon and then emits the light at a longer wavelength. The important parameters for characterizing the efficiency of emission are the quantum yield Q. The quantum yield can be expressed as

$$Q = \frac{number of emitted photons}{number of absorbed photons} (Eq. 3)$$

There are two common methods for determining the quantum yield experimentally which are the comparative method and the absolute method. The comparative method involves the comparison between a sample and a standard with a known quantum yield.³⁰ For the comparative method, the quantum yield of a sample can be calculated by the following equation,

$$Q_L^{Ln} = Q_S \times \frac{E_X}{E_S} \times \frac{A_S(\lambda_S)}{A_X(\lambda_X)} \times \frac{I_S(\lambda_S)}{I_X(\lambda_X)} \times \frac{n_S^2}{n_X^2}$$
(Eq. 4)

where *E* is the corrected emission spectrum, *I* is the intensity of the excitation source and *n* is the refractive index. In order to achieve a linearity between the intensity of the emission and the concentration of the sample, the absorbance of the sample should be less than 0.1 AU to minimize re-absorption effect. For the choice of standard, the best option will be the standard compound that has the same excitation wavelength with the sample. Then measurement can be safely performed up to 0.5 AU without the problem of dissociation of the sample.

The quantum yield can also be determined by the absolute method.^{31, 32} The absolute method involves the use of an integrating sphere. The integrating sphere is made up of a material with >99% reflectance and a lens that can focus the excitation light beam into the sample. The emission and the scatter can be monitored by a detector. Using integrating sphere to determine the quantum yield is more accurate since no standard is required.

1.3.4 Circular dichroism spectroscopy

Different from UV-visible spectroscopy which measures the absorption of unpolarized light, circular dichroism measures the differential absorption of left- and right-handed circularly polarized light.³³ A chiral molecule has different left- and right-handed light absorption coefficients. In circular dichroism spectroscopy, polarized light will pass through a sample and be absorbed by the sample in different proportion. The unit for circular dichroism is ellipticity ([θ]), which can be determined by the equation

$$\theta = 32.98\Delta A$$
 (Eq. 5)

where ΔA is the differential absorption of left- and right-handed circularly polarized light. A chiral compound will exhibit a positive circular dichroism signal when the left-handed circularly polarized light is absorbed in a greater extent than the righted handed circularly polarized light. Circular dichroism spectroscopy is useful for studying the chiroptical information of the ground state of the sample as well as the optical activity of biological secondary structure such as protein.³⁴ Inherently optically active system exhibits active CD signals. Moreover, for achiral system, CD may be induced by an external magnetic field.³⁵

<u>1.3.5 Circular polarized luminescence spectroscopy</u>

Circularly polarized luminescence spectroscopy is another technique that can characterize and give chiroptical information of chiral compounds. CPL is a more sensitive technique as it shows the emission of chiral compounds. CPL spectroscopy measures the difference in emission intensities of the left- and right-handed CPL and the result is reported in a parameter called dissymmetry factor (g_{lum}) ,³³ which is defined as

$$g_{lum} = 2(I_L - I_R)/(I_L + I_R)$$
 (Eq. 6)

where I_L and I_R are the emission intensity of left- and right-handed CPL. The maximum of g_{lum} value is ±2.

The nature of g_{lum} can be considered as among electric dipole forbidden and magnetic dipole-allowed transitions. The luminescence dissymmetry factor can be expressed as,

$$g_{lum} = \frac{4m(\cos\theta)}{\mu}$$
 (Eq. 7)

Where μ and m are the electric and magnetic transition dipole vectors respectively, and θ is the angle between them.

It is expected that large g_{lum} value can be obtained when the transition involved is weak. From Eq. 7, it is expected that large g_{lum} value can be observed when electric dipole transition is forbidden but magnetic dipole transition is allowed.³⁶ Due to the unique electronic properties of lanthanide, circularly polarized luminescence spectroscopy is commonly used for characterizing lanthanide systems because of the magnetically allowed intra-configuration f-f transition, which can give high g_{lum} value.³⁷ The highest g_{lum} value that has been reported is 1.38.³³ Although the magnetically allowed f-f transition can give high g_{lum} value, the transition generally has very low intensity and thus CPL of lanthanide systems are difficult to measure.

Richardson has developed selection rules for lanthanide CPL.³⁸ The selection rules state that the dissymmetry factor of lanthanide transitions can be classified into three classes (*DI*, *DII* and *DIII*) based on angular momentum quantum number. The magnitude of the dissymmetry factor decreases from *DI* to *DIII*. For example, table 1-3 shows some examples of Eu³⁺ and Tb³⁺ transition. For ⁵D₀ to ⁷F₁ emission, it is predicted that this transition will exhibit the largest g_{lum} value since it is classified as DI, which is also evidenced by experimental observation.

lon	Transition Classification			
Eu ³⁺	${}^{5}D_{0} \rightarrow {}^{7}F_{0}$	DII		
	${}^{5}D_{0} \rightarrow {}^{7}F_{1}$	DI		
	${}^{5}D_{0} \rightarrow {}^{7}F_{2}$	DIII		
	$^{5}D_{0} \rightarrow ^{7}F_{3}$	DII		
	$^{5}D_{0} \rightarrow ^{7}F_{4}$	DIII		
Tb ³⁺	$^{5}D_{4} \rightarrow ^{7}F_{6}$	DIII		
	$^{5}D_{4} \rightarrow ^{7}F_{5}$	DII		
	${}^{5}D_{4} \rightarrow {}^{7}F_{4}$	DII		
	$^{5}D_{4} \rightarrow ^{7}F_{3}$	DII		
	${}^{5}D_{4} \rightarrow {}^{7}F_{2}$	DIII		
	${}^{5}D_{4} \rightarrow {}^{7}F_{1}$	DIII		
	${}^{5}D_{4} \rightarrow {}^{7}F_{0}$	DIII		

 Table 1-3.
 Selection rule for CPL of lanthanide complexes proposed by Richardson's group.³⁸

1.4 Supramolecular coordination compounds and their applications

1.4.1 General background

The origin of supramolecular chemistry can be traced back to biology. In biological systems, there are different types of biomacromolecules that can be found in the human body such as proteins and DNAs. With a few building blocks with simple organic structure, protein can achieve specific biological functions which allow information to be stored, retrieved and processed. Supramolecular chemistry applies this concept to the molecular level by constructing well-defined structure through intermolecular noncovalent interaction. The information can then be stored in the artificial molecular level and processed at the supramolecular level. To data, numerous artificial supramolecular systems have been reported. These systems included not only monometallic complexes, but also the higher ordered supramolecular complexes such as helicates, tetrahedra, cubes, metal-organic frameworks as well as interlocked structures.¹²

<u>1.4.2 Assembling strategies: pre-organization</u>

The classical method for preparing supramolecular coordination compounds involves the use of a ligand that can pre-organize a binding cavity for metal ion.³⁹⁻⁴¹ Generally, the stability of complexes that are made by pre-organization is high due to the de-solvation of water molecules. Taking lanthanides as an example, numerous lanthanide supramolecules have been built based on macrocyclic ligands such as cyclen⁴²⁻⁴⁴ and calixarences.⁴⁵ These macrocycle ligands are well designed before complexation in order to contain an inner cavity with appropriate size that can encapsulate the target lanthanide ion of choice to maximize the binding between the lanthanide ions and ligand. Upon complexation, the reorganization energy of the ligand is therefore minimized.¹⁸ A large number of macrocycle-based ligands with functional side arms have been prepared to investigate the potential application of these complexes.

1.4.3 Assembling strategies: self-assembly

Many supramolecules that are made based on pre-organization process have been reported and scientists would like to simplify the synthetic procedures. Taking advantage of the high electric field of metal ion, the small coordination building blocks can be self-assembled around the metal ions through weak intermolecular interactions.^{46, 47} Different from pre-organization which has a well-defined structure and binding site for metal ion, the formation of supramolecular architectures through self-assembly have received intense attention since it gives access to a wide range of supramolecular complexes based on multiple hydrogen bonding and donor-acceptor interactions.

Self-assembly process is governed by both internal factors and external factors.⁴⁶ Internal factors are concerned about the interaction between metal ions and ligands, such as ligand design, secondary metal coordination, hydrogen bond and van der Waals forces. Proper designing of ligand can enhance the interaction between metal ions and ligands and successful controlled formation of desired supramolecular structure with high stability will result (figure 1-6). External factors involve the concentration and stoichiometries of the reactant. The presence of foreign species is also considered an external factor since they may affect the self-assembly process through interaction with either metal ions or ligands. Moreover, since the formation of higher order of supramolecular architecture may require a progressive buildup of small units, reaction time should be taken into consideration.⁴⁶ The above factors increase the diversity of artificial supramolecules in terms of structure and function.⁴⁸



Figure 1-6. Parallel formation of Cu^+ double helicate and Ni^{2+} triple helicate via as self-assembly process of a mixture of two different ligands and metal ions.⁴⁶

To overcome the challenges mentioned above, Raymond et al. suggested three important criteria for ligand design.⁴⁹ First, the self-assembled supramolecular complexes can be constructed through multibranched chelating ligands to maximize the metal-ligand interaction. Second, any unwanted cluster and geometries can be removed by fixing the multiple chelating group within the ligand rigidly. Finally, labile metal ions are highly preferred so that mistakes can be corrected by the breaking and re-forming of metal-ligand bond.

<u>1.4.4 Ligand design for self-assembled helicate</u>

Helicate is one of the secondary structures that can be found in nature. Helicate can exist in either right- or left-handed structure.⁵⁰ Natural examples include amylose and DNA. In supramolecular chemistry, numerous artificial metal-based helicates have been reported and helicity can be introduced by conformation restriction of bulky side arms,⁵¹ inter- or intramolecular hydrogen bond⁵² as well as coordination geometry of metal ions.^{53, 54}



Figure 1-7. Different types of supramolecular helicate.⁵⁵

Many types of helicate have been reported and all these structures can be classified in terms of the number of metal ion, the number of coordination and the presence of helical twist (figure 1-7).^{55, 56} For example, (B) and (C) are coordinated tetrahedrally and octahedrally, respectively. For (E), (F) and (G), these three types do not contain any helical twist, while (H), (I), (J) and (K) are typical double- or triple-stranded that possess a helical twist and are commonly observed for self-assembled supramolecules.

The construction of helicate can be done by utilizing C_2 -symmetrical ligand. For example, the D_3 -symmetrical triple helicate can be formed by coordinating two metal ions with C_2 -symmetrical bis(bidentate) ligand (figure 1-8).⁴⁹ To ensure the formation

of desired helical structure, the two metal-chelating planes must be parallel to each other, given that they share the same C_3 axis. The central bridging linker between two metal chelating group can be flexible or rigid but a rigid linker is highly recommended since the increased flexibility of ligand may increase the amount of unwanted cluster.



Figure 1-8. Graphical representation of formation of D₃-symmetrical triple helicate by C₂-symmetrical ligands. Two metal-chelating planes are parallel along C₃ axis.⁴⁹

Different from metal-organic cage, which will be discussed in the following part, helicate does not contain any inner cavity which hinders its the potential application. However, such a simple system is extremely useful in studying the mechanism of supramolecular chemistry.⁵⁷ For example, during the formation of artificial supramolecular helicates, reactants may undergo a cooperative self-assembly process to form oligonuclear complexes.⁵⁸ This problem can be solved by adding a template molecule to direct to formation of M₂L₃ helicate.^{59, 60} Such a transformation depends on the number of coordinating sites of the ligand, the distance between two binding sites as well as the geometric constraints of the resulting complex, which is also crucial when preparing other supramolecular structures.⁵⁵

<u>1.4.5 Ligand design for self-assembled metal-organic tetrahedra</u>

Metal-organic tetrahedra, also known as metal-organic container, are a class of coordination driven metal complexes.⁶¹ Compared with helicates, metal-organic cages generally exhibit well-defined shape with a specific size of an inner cavity.⁶²⁻⁶⁴ The geometry of metal-organic tetrahedra can be described as polyhedron. There are two different types of polyhedron, Platonic solids and Archimedean solids (figure 1-9).⁶⁵ Platonic solids consist of multiple symmetry axes and mirror planes, while Archimedean solids are made up of unidentical polygonal planes. Most of the metalorganic cages are achiral because of the presence of inversion center and mirror plane symmetries.



Figure 1-9. Graphical representation of Platonic solids and Archimedean solids.⁶⁶

Generally, most of the metal-organic tetrahedra are constructed by two different metal-to-ligand ratios, M₄L₆ and M₄L₄.^{12, 65, 66} For M₄L₆ tetrahedron, four metal ions serve as vertices and six ligands serve as edges.⁶⁷⁻⁶⁹ In this case, C_2 -symmetrical bis(bidentate) ligands with rigid backbone is used to coordinated with metal ions. Raymond et al. suggested that the angle between two chelating planes should be 70.6° so that a tetrahedral cage would be formed instead of helicate (figure 1-10a).⁷⁰ For M₄L₃ tetrahedron, a C_3 -symmetrical ligand with a rigid central bridging linker should be used to avoid the coordination of two chelating groups with the same metal ion.⁴⁹ If the ligand is planar (for example, the use of a benzene as the central bridging linker), the ideal approach angle should be 19.4° (figure 1-10b).



Figure 1-10. Design principle of metal-organic tetrahedron proposed by Raymond's group. (a) Graphical representation showing the angle between two chelating planes is 70.6°. (b) Graphical representation showing the ideal approaching angle of 19.4°.⁴⁹

1.4.6 Examples of chiral lanthanide supramolecular complexes

Based on the ligand design principle, a number of chiral lanthanide supramolecular complexes have been reported. For example, Prof. Gunnlaugsson and
co-workers have reported the preparation of chiral lanthanide supramolecular helicates with use of bis-tridentate ligands (figure 1-11a).⁷¹ The ligand was designed and synthesized by incorporating the pcam chelating units with a 4,4'- methylenediphenyl central bridging linker. Upon coordination with Eu(OTf)₃, enantiomerically pure triple-stranded helicates were prepared and exhibited CPL signal with a large g_{lum} value = -0.23 at 593 nm. Prof. Gunnlaugsson further studied the supramolecular formation of lanthanide bimetallic helicates by varying the size of linker (figure 1-11b).^{72, 73} The supramolecular helicates displayed CPL signals and result showed that smaller linker could increase the stability of the helicates.



Figure 1-11. Structure of ligand designed by Prof. Gunnlaugsson and co-workers. (a) Reaction scheme of preparation of chiral lanthanide bimetallic helicates.⁷¹ (b) Ligands designed for studying the effect of linker effect.^{72, 73}

Apart from lanthanide bimetallic helicates, Ln_4L_6 tetrahedra are another structure that can be prepared by C_2 -bistridentate ligands. The first example of stereoselective chiral luminescent Eu³⁺ tetrahedron was reported by Prof. Sun and coworkers in 2015. Based on the ligand design principle that proposed by Prof. Raymond, the ligand was carefully designed to avoid the formation of helicate by using a naphthalene moiety as the central bridging linker. Upon coordination with Eu³⁺, the ligand and europium ions self-assembled to form a tetrahedron topology and the structure was confirmed by X-ray crystallography.⁷⁴ The chirality from the chiral moiety was found to transferred during the self-assembly process to give enantiomeric pure product as evidenced by mirror image in CD spectra. Based on this result, numerous tetrahedra structures were reported and their ligands structures were shown in figure 1-12.^{75, 76}



Figure 1-12. Structure of ligand prepared for forming tetrahedron. (a) Reaction scheme of preparation of chiral lanthanide supramolecular tetrahedron reported by Prof. Sun and co-workers.⁷⁴ (b) Ligands designed for preparation of tetrahedron.^{75, 76}

1.5 Scope of thesis

This thesis demonstrates lanthanide supramolecular complexes with various topologies. This thesis aims at investigating the design principle as well as the factors that governing the structural formation of lanthanide self-assembled supramolecules. Chapter 2a is a study of preparing chiral lanthanide supramolecular helicate by point chirality effect. Chapter 2b investigates the helicate-to-tetrahedron transformation as well as the effect of ionic radii towards supramolecular formation. Chapter 3 focuses on the preparation of challenging lanthanide heterometallic tetrahedron with the use of C_2 -symmetrical bis(tridentate) ligands. Chapter 4 examines the remote control of chirality in supramolecular complexes induced by chiral peptide.

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Chapter 2a: Lanthanide supramolecular helical diastereoselective breaking induced by point chirality

2a.1 Introduction

2a.1.1 Introduction to chiroptical lanthanide complexes

Lanthanide self-assembly supramolecular complexes have been proven to be an excellent candidate for molecular recognition due to their unique physical properties.¹ In order to apply these fascinating systems into biological research, numerous chiroptical lanthanide systems have been developed.²⁻⁷ Chirality is essential for biological research, especially for medicinal chemistry. For example, the change in chiral centre may change the effect of drugs.⁸ Therefore, development of chiroptical recognition is important for detecting chiral biomolecule such as amino acid.⁹⁻¹¹ To date, most of the chiral recognition by lanthanide supramolecules strongly rely on induced CPL.¹ The chiral induction was achieved by the interaction between chiral molecular and achiral lanthanide complexes. However, careful ligand design has to be taken into consideration since vacant metal site is required for induced iCPL and most of the reported iCPL lanthanide sensors are mononuclear complex with well-defined ligands.¹²⁻¹⁵

2a.1.2 Chirality and preparation of chiral lanthanide complexes

In order to develop chiroptical lanthanide system, control of stereoselectivity of lanthanide system is important.¹⁶⁻¹⁸ Due to the labile nature and poor stereochemical preferences of lanthanide ions, precise control of stereoselective formation of desired lanthanide supramolecular complexes is challenging.¹⁹ One of the most straightforward approach is point chirality.²⁰ By introduction of a predisposed point chiral group, the chiral information can be transferred from the chiral moiety to the metal centre, forcing the coordination orientation of the metal centre to be either Δ (right-handed twist) or Λ (left-handed twist). However, the control of point chirality is difficult and only few examples were reported.^{2, 3, 21}

2a.1.3 Scope of study



Figure 2a-1. Stereoselective formation of L1 and L2 upon coordination with Eu(OTf)3.

This study focuses on the development of new chiral lanthanide supramolecular complexes for potential application including chiroptical sensing of biomolecules. Two pairs of chiral ligands, **L1** and **L2**, were synthesized by introducing chiral moiety next to the metal chelating group. It is expected that the chiral complexes can be form via point chirality. Compared with that of **L1**, the point chirality of **L2** is one carbon unit farther apart from the chelating unit. This slight difference in distance between the point chirality and the metal chelating group towards the stereoselective formation is investigated.

2a.2 Result and discussion

2a.2.1 Design and synthesis of L1 and L2

L1 and L2 were designed with the use of pyridine-2,6-dicarboxamide (pcam) moieties. Each pcam moiety contains a pyridine ring and two carbonyl groups which is able form stable nine-coordinated metal complexes with lanthanide ion.²² Each metal chelating group was allowed to connect with a chiral moiety. A rod-like biphenyl linker was used as the linker to connect two metal chelating moieties. Although the rod-like linker is linear, recent research have demonstrated that bimetallic transition metal triple helicate can be prepared with the use of biphenyl linker.

L1 and L2 were synthesized by using generic HATU amide coupling reactions in 2 steps. All the intermediates and final ligand products were well characterized by NMR and ESI-HRMS. The structure of L2 was further characterized by X-ray crystallography. In the X-ray structure of L2, the pyridine ring and the carbonyl oxygens adopted a transoid conformations about the interannular C-C bonds (figure 2a-2).

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Figure 2a-2. X-ray crystallography of L2^{RR}.

2a.2.2 ¹HNMR and UV-CD titration

Previous research has shown that the use of transition metal with similar ligand design can lead to the formation bimetallic M₂L₃ triple helicate. Therefore, it was hypothesized that the metal-to-ligand ratio is 2 to 3. This hypothesis was first tested by ¹HNMR titration. The ¹HNMR titration of L1^{RR} was done in a mixture of CDCl₃/CD₃OD/CD₃CN in a ratio of 75:5:20 due to poor ligand solubility (figure 2a-3). Upon addition of Eu(OTf)₃ to the ligand solution, the ¹HNMR signal at 1.72 ppm, which corresponds to the CH₃ group of L1^{RR}, decreased gradually. A new ¹HNMR signal gradually increased at 1.79 ppm. The new signal at 1.79 ppm was broad which corresponds to the europium complex due to the paramagnetic nature of europium ions. When 0.66 eq. of Eu(OTf)₃ was added, all the ligand signal disappeared. The consumption of ligand at 0.66 eq. of Eu(OTf)₃ indicated the metal-to-ligand ratio is 2 to 3.



Figure 2a-3. Variation in ¹H NMR spectra upon titrating L1^{RR} (5.11 × 10⁻³ M in 75:5:20, v/v/v, of CDCl₃/CD₃OD/CD₃CN) with Eu(OTf)₃ (0.271 M in CD₃OD) at 296 K. (Peaks that are marked as a, b, c and d are from the residual solvents of CHCl₃, MeOH, H₂O and MeCN, respectively.)

The metal-to-ligand ratio was further examined by UV-Vis titration. UV-Vis titration was performed by slow addition of Eu(OTf)₃ to the ligand solution and the UV absorption of the solution was monitored (figure 2a-4). Upon addition of Eu(OTf)₃, the signal of ligand absorption decreased progressively at 326 nm. Simultaneously, an evolution of the absorption peak corresponding to the europium complex at 346 nm was observed. A single sharp end point was observed when 0.65 eq. Eu(OTf)₃ was added which conformed the hypothesis of metal-to-ligand 2 to 3.



Figure 2a-4. (a) Variation in UV-Vis absorption spectra of titrating $L1^{RR}$ (1.72 × 10⁻⁴ M, in 73:3:24, v/v/v, of CHCl₃/MeOH/MeCN) with Eu(OTf)₃ (0.038M in MeCN) at 298 K (Eu:L1^{RR} = 0.0–2.0). (b) Variation of molar extinction coefficients at four different wavelengths upon titrating L1^{RR} with Eu(OTf)₃.

¹HNMR and UV-Vis titration were also performed for **L2** to investigate the metal-to-ligand ratio. Since the ligand design of **L2** is similar with that of **L1** except for the terminal chiral group, it was expected that **L2** will also form Ln₂**L2**₃ species upon coordination with lanthanide ions. Indeed, similar result was observed for both UV-Vis titration. An end point of 0.66 eq. of Eu(OTf)₃ was observed for **L2** (figure 2a-5).



Figure 2a-5. (a) Variation in UV-Vis absorption spectra of titrating $L2^{RR}$ (1.65 × 10⁻⁴ M, in 73:3:24, v/v/v, of CHCl₃/MeOH/MeCN) with Eu(OTf)₃ (0.038M in MeCN) at 298 K (Eu: $L2^{RR}$ = 0.0–2.0). (b)

Eu(OTf)₃.

However, the ¹HNMR titration of **L2** showed a different result compared with that of **L1**. Although the disappearance of ligand signal was also observed when 0.68 eq. of Eu(OTf)₃ was added, two sets of signal appeared simultaneously at 0.97 and 1.40 ppm in a ratio of 1:1.1. This ¹HNMR titration confirmed that the metal-to-ligand ratio of **L2** was 2:3 and two species were formed upon coordination of Eu(OTf)₃ (figure 2a-6).



Figure 2a-6. Variation in ¹H NMR (400 MHz, 295 K) spectra of titrating $L2^{SS}$ (4.64 × 10⁻³ M in 47:6:47, v/v/v, of CDCl₃/CD₃OD/CD₃CN) with Eu(OTf)₃ (0.271 M in CD₃OD) at 296K. (Peaks that are marked as a, b, c, d are from the residual solvents.)

2a.2.3 Synthesis and X-ray crystal structure of [Eu₂L1₃], [Tb₂L1₃], and [Eu₂L2₃]

The corresponding europium complexes with L1 and L2 was synthesized following the experimental condition of ¹HNMR titration. [Eu₂L1₃] and [Eu₂L2₃] were synthesized by reacting two equivalents of $Eu(OTf)_3$ with three equivalents of ligand. Mixture of solvent was used due to the poor solubility of L1 and L2. Single crystal of both $[Eu_2L1_3]$ and $[Eu_2L2_3]$ can be obtained by slow evaporation of the complex in MeCN. In the crystal structure of $[Eu_2L1_3]$, the overall P helicate was formed by the successive interacnnular C-N and C-C bond rotation between two metal centres and the overall helicate structure was stabilized by a total of five strong and one weak CH- π interaction (figure 2a-7a).²³ Each Eu was coordinated with three pcam moieties and exhibited the same Δ -configuration. The Eu metal centre can be best described as distorted tricapped trigonal prismatic geometry. Eu-N distances was found to be 2.532(5) to 2.566(5) Å and Eu-O distances range from 2.380(5) to 2.441(5) Å. [Tb₂L1₃] was also prepared by slow evaporation of complex solution in MeCN. Similar with $[Eu_2L1_3]$, each terbium centre can be described as tricapped trigonal prismatic geometry. Tb-N distances range from 2.500(5) to 2.566(5) Å and Tb-O distances range from 2.377(4) to 2.432(4) Å (figure 2a-7b). A single crystal of $[Eu_2L2_3]$ was also obtained for X-ray crystallography. Although the quality of the crystal was not suitable for analysis, it showed the overall helical structure of [Eu₂L2₃] (figure 2a-7c).



Figure 2a-7. X-ray crystal structure of (a) [Eu₂L1₃], (b) [Tb₂L1₃].and (c) [Eu₂L2₃].

2a.2.4 NMR analysis of [Eu₂L1₃] and [Eu₂L2₃]

After confirming the topologies of $[Eu_2L1_3]$ and $[Eu_2L2_3]$, $[Eu_2L1_3]$ and $[Eu_2L2_3]$ were then characterized by NMR. ¹HNMR was first done for $[Eu_2L1_3]$. In the ¹HNMR spectrum of $[Eu_2L1_3]$, all signals are broad due to the paramagnetic nature of europium ions. The number of integrations also matched the *C*₂-symmetric nature of L1 (figure 2a-8). For ¹³CNMR, only single set of signals was observed which matched the *C*₂-symmetric nature of L1 (figure 2a-9). The corresponding $[La_2L1_3]$ complex was also synthesized for comparing with $[Eu_2L1_3]$ (figure 2a-10). The ¹HNMR of $[La_2L1_3]$ also showed single set of signals which indicated the presence of single and highly symmetric species.



Figure 2a-8. ¹HNMR spectrum of [Eu₂(L1^{RR})₃](CF₃SO₃)₆ (CD₃CN). The insets show the expansion of the

corresponding region as indicated.



Figure 2a-9. ¹³CNMR spectrum of [Eu₂(L1^{RR})₃](CF₃SO₃)₆ (CD₃CN). The insets show the expansion of the

corresponding region as indicated.



Figure 2a-10. ¹H NMR spectrum of **[La₂(L1^{ss})₃](CF₃SO₃)**₆ (CD₃CN). The insets show the expansion of the corresponding region as indicated.

Similar NMR studies were also performed for L2. However, both ¹HNMR (figure 2a-11) and ¹³CNMR (figure 2a-12) of [Eu₂L2₃] showed a very different result compared with that of [Eu₂L1₃]. In the ¹HNMR and ¹³CNMR spectrum of [Eu₂L2₃], two sets of signals were observed. Two CH₃ signals at aliphatic region (0.84 and 1.26 ppm) were observed which indicated the presence of two species. The integration of CH₃ signal showed the ratio of two species was around 1:1.1. Increasing temperature tends to alter the dynamic exchange process by shifting one to another (figure 2a-13). However, preparation of pure single species is not possible. Diamagnetic [La₂L2₃] was also prepared for further NMR studies (figure 2a-14). The NMR spectrum of [La₂L2₃] also



Figure 2a-11. ¹HNMR spectrum of **[Eu2(L2^{RR})3](CF3SO3)**₆ (CD₃CN). The insets show the expansion of the corresponding region as indicated. Set 1 and set 2 are preliminary assigned that based on their



integration.

Figure 2a-12. ¹³CNMR spectrum of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (CD₃CN). The insets show the expansion of the corresponding region as indicated. Set 1 and set 2 are preliminary assigned that based on their

integration.



Figure 2a-13. ¹HNMR spectrum of [Eu2(L2^{RR})₃](CF₃SO₃)₆ (CD₃CN). a) At 295 K. b) At 345 K, c) Overlaying

of the two spectra from 295 K and 345 K.



Figure 2a-14. ¹HNMR spectrum of [La₂(L2^{SS})₃](CF₃SO₃)₆ (CD₃CN). The insets show the expansion of the corresponding region as indicated. Set 1 and set 2 are preliminary assigned that based on their integration.

2a.2.5 Photophysical measurement

Solution circular dichroism (CD) measurement was performed for both $[Eu_2L1_3]$ and $[Eu_2L2_3]$. In the CD spectrum of $[Eu_2L1_3]$, five strong cotton effects from π to π^* was observed at 379, 333, 283, 252 and 216 nm. Mirror image of CD spectrum was observed for *R*- and *S*-isomer which indicated the successful transfer of chiral information from the chiral moiety to metal centre (figure 2a-15a). Similar mirror image was also observed for the CD spectrum of $[La_2L1_3]$.

For $[Eu_2L2_3]$, different result was observed in CD spectrum. Although mirror image of CD spectrum was still observed for both *R*- and *S*- isomer, a significant decrease in CD intensity was observed (figure 2a-15b). Compared with that of $[Eu_2L1_3]$, the percentage of decrease in CD intensity of $[Eu_2L2_3]$ was found to be 96.8% (379 nm), 99% (333 nm), 77% (283 nm), 69% (252 nm) and 84% (216 nm) respectively. Decrease in CD intensity was also observed for $[La_2L2_3]$. Both NMR and CD result revealed that the diastereomeric *P* and *M* helicate were formed with the use of L2. The decrease in CD signal can be attributed by the presence of two isomers (*P* and *M* helicate). *P* and *M* helicate twisted into two different directions, which exhibited opposite CD signal at the same wavelength and thus the overall CD signal was decreased. ESI-HRMS showed no higher-ordered supramolecules (e.g. $[Eu_4L2_6]$) which confirmed the CD signals are arised from helicates only.



Figure 2a-15. CD spectra of (a) $[Eu_2(L1)_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M) and $[Eu_2(L2)_3](CF_3SO_3)_6$ (3.52 × 10⁻⁵ M) in MeCN and (b) $[La_2(L1)_3](CF_3SO_3)_6$ (3.58 × 10⁻⁵ M) and $[La_2(L2)_3](CF_3SO_3)_6$ (4.23 × 10⁻⁵ M) in MeCN.

The luminescent properties of $[Eu_2L1_3]$, and $[Eu_2L2_3]$ were examined in both solid state and solution state. Generally, both $[Eu_2L1_3]$, and $[Eu_2L2_3]$ showed characteristic narrow Eu emission at 594, 615 and 619, 687, 697 and 704 nm, corresponding to the transition from the first excited state ⁵D₀ to the multiple ground states (⁷F_J, *J* = 1, 2 and 4 respectively). Lifetime measurement was also performed for $[Gd_2L1_3]$ at 77K and the lifetime as determined as around 0.2 to 0.4 ms, which are shorter than typical europium complex.

2a.3 Conclusion

In conclusion, this study demonstrated the strategies of preparing chiral lanthanide supramolecular complexes. With the use of linear central bridging linker, lanthanide triple stranded helicate can be prepared. Moreover, it was found that the diastereoselective and non-diastereoselective supramolecular formation was highly sensitive to the position of chiral moiety. Point-to-helical chirality transfer was efficient when the metal centre was close to the chiral group, while the nondiastereoselective formation of lanthanide helicate was observed by just simply extending the point chirality from the metal centre. This study was important for design and preparation of chiral lanthanide supramolecular complexes.

2a.4 Experimental section

2a.4.1 Synthesis

(S)-6-((1-phenylethyl)carbamoyl)picolinic acid and (R)-6-((1phenylethyl)carbamoyl)picolinic acid



This synthesis is reported by our group previously. To a stirred solution of 2,6pyridinedicarboxylic acid (5 g, 29.90 mmol, 2.50 equiv.) in anhydrous DMF (70 ml) at room temperature, HATU (4.55 g, 11.90 mmol, 1 equiv.) was added in portions over 10 min under nitrogen. After allowing it to stir for 20 min, (S)-(-)-1-phenylethylamine (1.53 ml, 11.97 mmol, 1 equiv.) was added dropwise and the reaction mixture was allowed to stir for 20 min. DIPEA (5.50 ml, 31.60 mmol, 2.60 equiv.) was then added to the reaction mixture over 5 min and the resulting solution was stirred at room temperature for 14 h. The reaction mixture was then diluted with H₂O (100 ml), and extracted with DCM (30 ml x 3), dried with MgSO₄, filtered, and concentrated in vacuo. Then dissolved it in water and extracted with EA to remove excess DMF. The resulting residue was purified by recrystallization in EtOAc solvent to give a white solid. **(S)-6-((1-phenylethyl)carbamoyl)picolinic acid**: (1.46g, 5.42 mmol, 45.39% yield), mp 110.2 - 117.3 °C. ¹H NMR (400 MHz, CDCl₃, 298K, δ): 1.65 (d, 7.0Hz, 3H), 5.37 (p, J=7.1Hz, 1H), 7.20-7.44 (m, 5H), 8.01 (d, J=8.2Hz, 1H), 8.11 (t, J=7.8Hz, 1H), 8.36 (dd, J=7.7, 1.2Hz, 1H), 8.48 (dd, J=7.8, 1.2Hz, 1H).¹³C NMR (400MHz, CDCl₃, 300K, δ): 21.50, 49.14, 126.25, 126.72, 126.95, 127.48, 128.67, 139.50, 142.63, 145.37, 149.56, 162.48, 164.67. HRMS (ESI) calcd. for $C_{15}H_{14}N_2O_3Na$ [(S)-6-((1phenylethyl)carbamoyl)picolinic acid+Na]⁺: 293.0897, found 293.0895. The enantiomeric purity was determined with HPLC using AS-H column (Hexane/ipropanol: 80/20; flow rate 1.0 ml/min) and compared with a racemic mixture according to the elution orders with the retention times, $t_s = 7.15$ mins and $t_{R} = 9.62$ mins) to be > 99.9%ee. (R)-6-((1-phenylethyl)carbamoyl)picolinic acid was isolated, following the procedure for (S)-6-((1-phenylethyl)carbamoyl)picolinic acid with the use of (R)-(+)-1-phenylethylamine instead, in 55.10% yield (1.78g, 6.59 mmol): mp 110.3 – 113.9 °C. ¹H NMR (400 MHz, CDCl₃, 298K): δ 1.60 (d, J=6.9Hz, 3H), 5.34 (p, J=7.1Hz, 1H), 7.18-7.41 (m, 5H), 8.06 (t, J=7.8Hz, 1H), 8.29-8.38 (m, 2H), 8.44 (dd, J=7.8, 1.2Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, 300K, δ): 21.50, 30.94, 49.12, 126.23, 126.69, 126.95, 127.44, 128.63, 139.48, 142.63, 145.41, 149.52, 162.58, 164.80. HRMS (ESI) calcd. for C₁₅H₁₄N₂O₃Na [(R)- 6-((1-phenylethyl)carbamoyl)picolinic acid +Na]⁺: 293.0897, found 293.0896. The enantiomeric purity was determined to be 97.0% ee.

N,*N*[']-(biphenyl-4,4'-diyl)bis[6-(*R*)-(1-phenylethylcarbamoyl)-pyridine-2-

dicarboxamide] $(L1^{RR})$ and N,N'-(biphenyl-4,4'-diyl)bis[6-(S)-(1phenylethylcarbamoyl)- pyridine-2-dicarboxamide] (L1^{SS})



To a stirred solution of (R)-6-((1-phenylethyl)carbamoyl)picolinic acid (2.50 g, 9.26 mmol, 2.2 equiv.) in anhydrous DMF (40 mL) at room temperature, HATU (7.70 g, 20.3 mmol, 4.8 equiv.) was added under nitrogen. After allowing it to stir for 20 min, a benzidine (0.79 g, 4.27 mmol, 1.0 equiv.) was added and the reaction mixture was allowed to stir for 20 min in dark. DIPEA (9.24 mL, 53.03 mmol, 12.5 equiv.) was then added to the reaction mixture and the resulting solution was stirred at room temperature for 14 h. The reaction mixture was then diluted with H₂O (100 mL) and extracted with DCM (5 × 30 mL). After removing all of the organic volatile under reduced pressure, the residue was diluted with ethyl acetate (50 mL) and then washed with H_2O (5 × 30 mL) to remove the remained DMF. The organic layer was separated and then concentrated directly under reduced pressure. The residue was then diluted with MeCN (20 mL), and fine powder was progressively precipitated out. Then the solid was collected by filtration and the desired compound was isolated. (L1^{RR}): (2.53 g, 3.67 mmol, 86% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 299 K, δ): 1.62 (d, J = 8 Hz, 6H), 5.25–5.31 (m, 2H), 7.23 (t, J = 8 Hz, 2H), 7.34 (t, J = 8 Hz, 4H), 7.45 (d, J = 8 Hz, 4H), 7.79 (d, J = 8 Hz, 4H), 7.93 (d, J = 8 Hz, 4H), 8.20-8.26 (m, 4H), 8.34 (d, J = 8 Hz, 2H), 9.60 (d, J = 8 Hz, 2H), 10.93 (s, 2H). ¹³C NMR (100.6 MHz, CD₃)₂SO, 300 K, δ): 22.67, 49.08, 122.54, 125.79, 126.05, 127.02, 127.51, 127.69, 129.23, 136.38, 138.02, 140.62, 145.00, 149.58, 150.02, 162.62, 163.53. HRMS (ESI) calcd. for C₄₂H₃₆N₆O₄Na [M+Na]⁺: 711.2690, found 711.2681. (L1^{SS}) was synthesized, following the procedure for (L1^{RR}) with the use of (S)-6-((1-phenylethyl)carbamoyl)picolinic acid instead, in 80% yield (2.35 g, 3.42 mmol): ¹H NMR (400 MHz, (CD₃)₂SO, 299 K, δ): 1.61 (d, J = 8 Hz, 6H), 5.24– 5.28 (m, 2H), 7.19 (t, J = 8 Hz, 2H), 7.30 (t, J = 8 Hz, 4H), 7.42 (d, J = 8 Hz, 4H), 7.75 (d, J = 8 Hz, 4H), 7.90 (d, J = 8 Hz, 4H), 8.18–8.21 (m, 4H), 8.31 (d, J = 8 Hz, 2H), 9.58 (d, J = 8 Hz, 2H), 10.91 (s, 2H). ¹³C NMR (100.6 MHz, CD₃)₂SO, 300 K, δ): 22.74, 49.17, 122.62, 125.86, 126.12, 127.10, 127.58, 127.76, 129.30, 136.46, 138.10, 140.68, 145.08, 149.66, 150.10, 162.70, 163.62. HRMS (ESI) calcd. for C₄₂H₃₆N₆O₄Na [M+Na]⁺: 711.2690, found 711.2682.

(*R*)-6-(2-phenylpropylcarbamoyl)picolinic acid or (*S*)-6-(2-phenylpropyl carbamoyl)picolinic acid



To a stirred solution of 2,6-pyridinedicarboxylic acid (9.89 g, 59.1 mmol, 4.0 equiv.) in anhydrous DMF (130 mL) at room temperature, HATU (5.64 g, 14.8 mmol, 1.0 equiv.) was added by portions over 10 min under nitrogen. After allowing it to stir for 20 min, a (R)-β-methylphenethylamine (2.11 mL, 14.83 mmol, 1.0 equiv.) was added dropwisely and the reaction mixture was allowed to stir for 20 min. DIPEA (5.68 mL, 32.6 mmol, 2.2 equiv.) was then added to the reaction mixture over 5 min and the resulting solution was stirred at room temperature for 18 h. The reaction mixture was then diluted with H_2O (200 mL), and extracted with DCM (5 × 50 mL), dried with MgSO₄, filtered, and concentrated in vacuo. The resulting residue was purified with flash column chromatography (DCM/EtOH 12:1, v/v) to give a white solid. (R)-6-(2phenylpropylcarbamoyl)picolinic acid: (1.93 g, 6.8 mmol, 46% yield) ¹H NMR (400 MHz, CD₃OD, 299 K, δ): 1.35 (d, J = 8.0 Hz, 3H), 3.17 (qin, J = 8 Hz, 1H), 3.61 (d, J = 8 Hz, 2H), 7.18–7.22 (m, 1H), 7.28–7.32 (m, 4H), 8.15 (t, J = 8 Hz, 1H), 8.30 (d, J = 8 Hz, 2H), 9.42 (m, 1H). ¹³C NMR (100.6 MHz, CD₃OD, 300 K, δ): 20.46, 41.80, 48.58, 127.34, 128.37, 129.05, 129.15, 130.36, 141.30, 146.59, 148.81, 151.99, 166.50, 168.28. HRMS

(ESI) calcd. for C₁₆H₁₆N₂O₃Na [M+Na]⁺: 307.1053, found 307.1053. The enantiomeric purity was determined with HPLC with AS-H column (Hexane/*i*-propanol: 80/20; flow rate: 0.25 ml/min) and compared with a racemic mixture according to the elution orders with retention times, $t_s = 46.33$ min and $t_R = 49.46$ min) to be 88% ee. (S)-6-(2phenylpropyl carbamoyl)picolinic acid was isolated, following the procedure for (R)-6-(2-phenylpropylcarbamoyl)picolinic with (S)-βacid the use of methylphenethylamine instead, in 43% yield (1.81 g, 6.36 mmol): ¹H NMR (400 MHz, CD₃OD, 299 K, δ): 1.36 (d, J = 8.0 Hz, 3H), 3.17 (qin, J = 8 Hz, 1H), 3.62 (d, J = 8 Hz, 2H), 7.19–7.22 (m, 1H), 7.28–7.31 (m, 4H), 8.15 (t, J = 8 Hz, 1H), 8.31 (d, J = 4 Hz, 2H). ¹³C NMR (100.6 MHz, CD₃OD, 300 K, δ): 20.46, 41.81, 48.60, 127.34, 128.38, 129.06, 129.16, 130.37, 141.32, 146.60, 148.83, 152.00, 166.51, 168.29. HRMS (ESI) calcd. for C₁₆H₁₆N₂O₃Na [M+Na]⁺: 307.1053, found 307.1052. The enantiomeric purity was determined to be 96% ee.

N,N'-(biphenyl-4,4'-diyl)bis[6-(R)-(2-phenylpropylcarbamoyl)-pyridine-2,6dicarboxamide] (L2^{RR}) and N,N'-(biphenyl-4,4'-diyl)bis[6-(S)-(2-phenylpropyl carbamoyl)-pyridine-2,6-dicarboxamide] (L2^{SS})



To a stirred solution of (*R*)-6-(2-phenylpropylcarbamoyl)picolinic acid (1.00 g, 3.52 mmol, 2.2 equiv.) in anhydrous DMF (16 mL) at room temperature, HATU (2.93 g, 7.69 mmol, 4.8 equiv.) was added under nitrogen. After allowing it to stir for 20 min, a benzidine (0.30 g, 1.61 mmol, 1.0 equiv.) was added and the reaction mixture was allowed to stir for 20 min in dark. DIPEA (3.51 mL, 20.16 mmol, 12.5 equiv.) was then

added to the reaction mixture and the resulting solution was stirred at room temperature for 14 h. The reaction mixture was then diluted with H₂O (20 mL) and extracted with DCM (5 \times 30 mL). After removing all of the organic volatile under reduced pressure, the residue was diluted with ethyl acetate (20 mL) and then washed with H_2O (5 × 30 mL) to remove the remained DMF. The organic layer was separated and concentrated directly under reduced pressure. The residue was then diluted with MeCN (20 mL), and fine powder was progressively precipitated out. Then the solid was collected by filtration and the desired compound was isolated. (L2^{RR}): (0.42 g, 0.588 mmol, 73% yield), ¹H NMR (400 MHz, CDCl₃, 299K, δ): 1.42 (d, *J* = 8 Hz, 6H), 3.11–3.17 (m, 2H), 3.48–3.54 (m, 2H), 4.00–4.06 (m, 2H), 7.26–7.31 (m, 2H), 7.34–7.42 (m, 8H), 7.60–7.69 (m, 10H), 8.09 (t, J = 8 Hz, 2H), 8.41 (d, J = 8 Hz, 2H), 8.44 (d, J = 8 Hz, 2H), 9.20 (s, 2H). ¹³C NMR (100.6 MHz, CDCl₃, 300 K, δ): 19.31, 39.97, 45.96, 120.70, 125.21, 125.45, 127.14, 127.31, 127.35, 128.95, 136.29, 136.89, 139.33, 143.91, 148.63, 148.88, 161.10, 163.09. HRMS (ESI) calcd. for C₄₄H₄₀N₆O₄Na [M+Na]⁺: 739.3003, found 739.2994. (L2^{ss}) was isolated, following the procedure for (L2^{RR}) with the use of (S)-6-(2-phenylpropyl carbamoyl)picolinic acid instead, in 69% yield (0.40 g, 0.555 mmol): ¹H NMR (400 MHz, CDCl₃, 298K, δ): 1.42 (d, *J* = 8 Hz, 6H), 3.13–3.15 (m, 2H), 3.48–3.53 (m, 2H), 4.01–4.06 (m, 2H), 7.27–7.31 (m, 2H), 7.35–7.42 (m, 8H), 7.58–7.60 (m, 2H), 7.65–7.70 (m, 8H), 8.10 (t, J = 8 Hz, 2H), 8.41 (d, J = 8 Hz, 2H), 8.45 (d, J = 8 Hz, 2H), 9.19 (s, 2H). ¹³C NMR (100.6 MHz, CDCl₃, 298K, δ): 19.32, 39.99, 45.96, 120.71, 125.24, 125.48, 127.17, 127.35, 127.37, 128.97, 136.30, 136.94, 139.36, 143.92, 148.63, 148.88, 161.11, 163.02. HRMS (ESI) calcd. for C₄₄H₄₀N₆O₄Na [M+Na]⁺: 739.3003, found 739.2996.

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{*N*,*N*[']-(biphenyl-4,4'-diyl)bis[6-(*R*)-(1-phenylethyl-carbamoyl)-pyridine-2dicarboxamide]}·2Eu·6(CF₃SO₃) [Eu₂(L1^{RR})₃] and 3{*N*,*N*[']-(biphenyl-4,4'- diyl)bis[6-(*S*)-(1-phenylethylcarbamoyl)-pyridine-2-dicarboxamide]}·2Eu·6(CF₃SO₃) [Eu₂(L1^{SS})₃]



To a white suspension of L1^{RR} (0.103 g, 0.150 mmol, 1.5 equiv.) in a mixture of 13 mL of DCM/MeOH (12:1, v/v), a solution of Eu(CF₃SO₃)₃ (0.060 g, 0.100 mmol, 1 equiv.) in 5 mL of MeCN was added. The suspension was changed to yellow turbidity immediately. The reaction mixture was then refluxed for 16 h and the solid progressively dissolved to give a resulting homogeneous yellow solution. The solvent was removed under reduced pressure. The solid was then washed with THF to give the desired product. [Eu₂(L1^{RR})₃]: (0.140 g, 0.043 mmol, 85% yield) ¹H NMR (400 MHz, CD₃CN, 298 K, δ): 1.54 (s, br., 3 × 6H, CH₃), 4.95 (s, br., 3 × 2H, N-H), 5.27 (s, br., 3 × 2H, (CH₃)CH), 6.36 (s, br., 3 × 4H), 6.71 (s, br., 3 × 4H, phenyl-H), 6.77 (s, br., 3 × 4H, phenyl-H), 6.83 (s, br., 3 × 6H), 6.95 (s, br., 3 × 2H, N-H), 7.09 (s, br. 3 × 2H), 7.87 (s, br. 3 × 4H, phenyl-H). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, δ): 22.45 (CH₃), 52.30 (CH), 93.11, 93.62, 123.32, 126.44, 128.14, 128.35, 129.44, 136.95, 139.06, 143.68, 144.82, 145.70, 156.35, 161.81 (CO), 165.23 (CO). HRMS (ESI) calcd. for C130H108N18Eu2O24S4F12 [M-20Tf]²⁺: 1481.2431 (¹⁵¹Eu based), found 1481.2432. Calculated for C₁₃₂H₁₀₈N₁₈O₃₀Eu₂F₁₈S₆·2H₂O: C, 48.03; H, 3.42; N, 7.64%; Found: C, 47.24; H, 3.37; N, 7.47 %; [Eu₂(L1^{SS})₃] was synthesized, following the procedure for [Eu₂(L1^{RR})₃] with the

use of L1^{SS} instead, in 92% yield (0.150 g, 0.046 mmol): ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 1.53 (s, br., 3 × 6H, CH₃), 4.92 (s, br., 3 × 2H, N-H), 5.25 (s, br., 3 × 2H, (CH₃)CH), 6.36 (s, br., 3 × 4H), 6.70 (s, br., 3 × 4H, phenyl-H), 6.74 (s, br., 3 × 4H, phenyl-H), 6.82 (s, br., 3 × 6H), 6.92 (s, br., 3 × 2H, N-H), 7.07 (s, br. 3 × 2H), 7.85 (s, br. 3 × 4H, phenyl-H). ¹³C NMR (100.6 MHz, CD₃CN, 300 K, δ): 22.46 (CH₃), 52.37 (CH), 93.34, 93.80, 123.35, 126.52, 128.19, 128.44, 129.51, 136.93, 139.12, 143.63, 144.90, 145.77, 156.35, 161.83 (CO), 165.23 (CO). HRMS (ESI) calcd. for C130H108N18Eu2O24S4F12 [M-20Tf]²⁺: (¹⁵¹Eu 1481.2431 based), found 1481.2433. Calculated for C₁₃₂H₁₀₈N₁₈O₃₀Eu₂F₁₈S₆·2H₂O: C, 48.03; H, 3.42; N, 7.64%; Found: C, 47.49; H, 3.42; N, 7.54 %.

 $3{N,N'-(biphenyl-4,4'-diyl)bis[6-(R)-(1-phenylethyl-carbamoyl)-pyridine-2$ $dicarboxamide]}·2La·6(CF₃SO₃) [La₂(L1^{RR})₃] and <math>3{N,N'-(biphenyl-4,4'-diyl)bis[6-(S)-(1-phenylethylcarbamoyl)-pyridine-2-dicarboxamide]}·2La·6(CF₃SO₃) [La₂(L1^{SS})₃]$



To a white suspension of $L1^{RR}$ (0.050 g, 0.073 mmol, 1.5 equiv.) in a mixture of 6 mL of DCM/MeOH (12:1, v/v), a solution of La(CF₃SO₃)₃ (0.028 g, 0.048 mmol, 1 equiv.) in 3 mL of MeCN was added. The suspension was changed to yellow turbidity immediately. The reaction mixture was then refluxed for 16 h and the solid progressively dissolved to give a resulting homogeneous yellow solution. The solvent was removed under

reduced pressure. The solid was then washed with THF to give the desired product. [La₂(L1^{RR})₃]: (0.071 g, 0.022 mmol, 93% yield) ¹H NMR (400 MHz, CD₃CN, 298 K, δ): 1.75 (d, J = 8 Hz, 3 × 6H, CH₃), 5.08–5.16 (m, 3 × 2H, (CH₃)CH), 6.64 (d, J = 8 Hz, 3 × 4H, phenyl-H), 7.11–7.17 (m, $3 \times 4H$, phenyl-H), 7.18–7.23 (overlapping of two type of peaks, m, 3 × 4H, phenyl-H and m, 3 × 2H), 7.71 (d, J = 8 Hz, 3 × 4H, phenyl-H), 8.50-8.55 (m, 3 × 4H), 8.56–8.60 (m, 3 × 2H), 9.09 (d, J = 8 Hz, 3 × 2H, N-H), 10.32 (s, 3 × 2H, N-*H*). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, δ): 22.15 (*C*H₃), 53.71 (*C*(CH₃)H), 122.86 (*C*H), 127.38 (CH), 127.93 (CH), 128.63 (CH), 129.08 (CH), 130.05 (CH), 136.86, 139.19, 143.17, 144.54 (CH), 150.14, 150.79, 168.12 (CO), 168.95 (CO). HRMS (ESI) calcd. for $C_{129}H_{108}N_{18}La_2O_{21}S_3F_9$ [M-3OTf]³⁺: 929.8355, found 929.8354. Calculated for C₁₃₂H₁₀₈N₁₈O₃₀La₂F₁₈S₆·5H₂O·2CH₃OH: C, 47.44; H, 3.74; N, 7.43%; Found: C, 46.14; H, 3.72; N, 7.32 %; [La₂(L1^{SS})₃] was synthesized, following the procedure for [La₂(L1^{RR})₃] with the use of L1^{ss} instead: (0.070 g, 0.022 mmol, 90% yield) ¹H NMR (400 MHz, CD₃CN, 298 K, δ): 1.75 (d, *J* = 8 Hz, 3 × 6H, CH₃), 5.09–5.16 (m, 3 × 2H, (CH₃)CH), 6.64 (d, J = 8 Hz, 3 × 4H, phenyl-H), 7.12–7.16 (m, 3 × 4H, phenyl-H), 7.18–7.23 (m, 3 × 4H, phenyl-*H*; m, 3 × 2H), 7.71 (d, J = 8 Hz, 3 × 4H, phenyl-*H*), 8.51 (t, J = 8 Hz, 3 × 2H), 8.56 (d, J = 8 Hz, 3 × 2H), 8.60 (d, J = 8 Hz, 3 × 2H), 9.09 (d, J = 8 Hz, 3 × 2H, N-H), 10.32 (s, 3 × 2H, N-*H*). ¹³C NMR (100.6 MHz, CD₃CN, 299 K, δ): 22.16 (*C*H₃), 53.73 (*C*(CH₃)H), 122.88 (CH), 127.41 (CH), 127.96 (CH), 128.65 (CH), 129.11 (CH), 130.08 (CH), 136.87, 139.22, 143.18, 144.57 (CH), 150.16, 150.81, 168.15 (CO), 168.98 (CO). HRMS (ESI) calcd. for C₁₂₉H₁₀₈N₁₈La₂O₂₁S₃F₉ [M-3OTf]³⁺: 929.8355, found 929.8352. Calculated for C₁₃₂H₁₀₈N₁₈O₃₀La₂F₁₈S₆·4H₂O: C, 47.89; H, 3.53; N, 7.62%; Found: C, 47.45; H, 3.46; N, 7.53 %.

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$3{N,N'-(biphenyl-4,4'-diyl)bis[6-(R)-(2-phenylpropylcarbamoyl)-pyridine-2,6$ $dicarboxamide]}·2Eu·6(CF₃SO₃) [Eu₂(L2^{RR})₃] and 3{ N,N'-(biphenyl-4,4'-diyl)bis[6- (S) (2-phenylpropylcarbamoyl)-pyridine-2,6-dicarboxamide]}·2Eu·6(CF₃SO₃) [Eu₂(L2^{SS})₃]$



To a white suspension of L2^{RR} (0.107 g, 0.150 mmol, 1.5 equiv.) in a mixture of 13 mL of DCM/MeOH (12:1, v/v), a solution of Eu(CF₃SO₃)₃ (0.060 g, 0.100 mmol, 1 equiv.) in 5 mL of MeCN was added. The solution was changed to yellow turbidity immediately. The solution was then refluxed and the solid progressively dissolved to give a resulting homogeneous yellow solution. After for 16 h, the solvent was removed under reduced pressure. The solid was then washed with DCM to give the desired product. [Eu₂(L2^{RR})₃]: (0.154 g, 0.046 mmol, 91% yield), ¹H NMR (400 MHz, CD₃CN, 298 K, some of the peaks are shown two sets of peaks, **A** and **B**, respectively in ~1.1:1 ratio, δ): 1.02 (s, br., 3 × 6H, CH₃, A), 1.33 (s, br., 3 × 6H, CH₃, B), 2.56 (s, br., 3 × 2H, (CH₃)CH, A), 2.96 (s, br., 3 × 2H, (CH₃)CH, B), 3.77 (overlapping of three type of peaks, br. 3 × 2H, CHH, A, 3 × 2H, CHH, A and 3 × 2H, CHH, B), 4.01 (s, br. 3 × 2H, CHH, B), 4.82 (s, br. 3 × 2H, NH, A), 4.96 (s, br. 3 × 2H, NH, B), 6.36 (s, br. 3 × 4H, A), 6.64 (m, br. 3 × 4H, B), 7.06-7.31 (m, br. 3 × 18H, A and 3 × 18H B), 8.36 (s, br. 3 × 4H, A), 8.44 (s, br. 3 × 4H, B). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, some of the peaks are shown two sets of peaks, δ): 19.35 (CH₃), 19.63 (CH₃), 41.00 (CH), 41.29 (CH), 47.94 (CH₂), 47.96 (CH₂), 93.07, 93.09, 93.59, 93.89, 123.63, 127.71, 127.90, 128.00, 128.41, 128.48, 129.62, 129.70, 137.26, 137.30, 139.33, 139.38, 144.36, 144.41, 144.55, 145.42, 145.88, 156.35, 161.94 (CO), 162.38 (CO), 165.13 (CO), 165.26 (CO). HRMS (ESI) calcd. for C₁₃₆H₁₂₀N₁₈Eu₂O₂₄S₄F₁₂ [M-2OTf]²⁺: 1523.2900 (¹⁵¹Eu based), found 1523.2905. Calculated for C₁₃₈H₁₂₀N₁₈O₃₀Eu₂F₁₈S₆·2H₂O: C, 48.97; H, 3.69; N, 7.45%; Found: C, 48.47; H, 3.63; N, 7.36 %; [Eu₂(L2^{SS})₃] was synthesized, following the procedure for [Eu₂(L2^{RR})₃] with the use of L2^{SS} instead, in 92% yield (0.154 g, 0.046 mmol): ¹H NMR (400 MHz, CD₃CN, 297 K, some of the signals are shown in two sets of peaks, **A** and **B**, respectively in $\sim 1.1:1$ ratio, δ): 0.90 (s, br., 3 × 6H, CH₃, A), 1.32 (s, br., 3 × 6H, CH₃, B), 2.54 (s, br., 3 × 2H, **CH, A**), 2.96 (s, br., 3 × 2H, **CH, B**), 3.74 (s, br. 3 × 2H, **CHH**, **A**), 3.78 (s, br., 3 × 2H, **CHH**, A and 3 × 2H, CHH, B), 4.02 (s, br. 3 × 2H, CHH, B), 4.73 (s, br. 3 × 2H, NH, A), 4.87 (s, br. 3 × 2H, **NH**, **B**), 6.33 (s, br. 3 × 4H, **A**), 6.64 (m, br. 3 × 4H, **B**), 7.07–7.31 (m, br. 3 × 18H, **A** and m, br. 3 × 18H **B**), 8.37 (s, br. 3 × 4H, **A**), 8.45 (s, br. 3 × 4H, **B**). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, some of the peaks are shown into two sets of peaks, δ): 19.35 (CH₃), 19.63 (CH₃), 41.00 (CH), 41.29 (CH), 47.93 (CH₂), 47.98 (CH₂), 93.13, 93.18, 93.71, 93.91, 123.63, 127.88, 127.91, 128.00, 128.41, 128.49, 129.62, 129.70, 137.23, 137.27, 139.34, 139.39, 144.36, 144.40, 144.60, 145.43, 145.90, 156.32, 161.91 (CO), 162.36 (**CO**), 165.13 (**CO**), 165.26 (**CO**). HRMS (ESI) calcd. for C₁₃₆H₁₂₀N₁₈Eu₂O₂₄S₄F₁₂ [M-2OTf]²⁺: 1523.2900 (¹⁵¹Eu based), found 1523.2889. Calculated for C₁₃₈H₁₂₀N₁₈O₃₀Eu₂F₁₈S₆·2H₂O: C, 48.97; H, 3.69; N, 7.45%; Found: C, 48.24; H, 3.60; N, 7.31 %.

3{*N*,*N*[']-(biphenyl-4,4'-diyl)bis[6-(*R*)-(2-phenylpropylcarbamoyl)-pyridine-2,6dicarboxamide]}·2La·6(CF₃SO₃) [La₂(L2^{RR})₃] and 3{*N*,*N*[']-(biphenyl-4,4'-diyl)bis[6-(*S*)-(2-phenylpropylcarbamoyl)-pyridine-2,6-dicarboxamide]}·2La·6(CF₃SO₃) [La₂(L2^{SS})₃]



To a white suspension of L2^{RR} (0.040 g, 0.056 mmol, 1.5 equiv.) in a mixture of 6 mL of DCM/MeOH (8:1, v/v), a solution of La(CF₃SO₃)₃ (0.022 g, 0.037 mmol, 1 equiv.) in 8 mL of MeCN was added. The suspension was changed to yellow turbidity immediately. The reaction mixture was then refluxed for 16 h and the solid progressively dissolved to give a resulting homogeneous yellow solution. The solvent was removed under reduced pressure. The solid was then washed with DCM to give the desired product. [La₂(L2^{RR})₃]: (0.055 g, 0.016 mmol, 89% yield) ¹H NMR (400 MHz, CD₃CN, 298 K, some of the signals are shown in two sets of peaks, **A** and **B**, respectively in ~1:1.1 ratio, δ): 1.02 (s, br., 3 × 6H, A), 1.18 (s, br., 3 × 6H, B), 2.77 (s, br., 3 × 2H, B), 2.93 (s, br., 3 × 2H, A), 3.40 (s, br., 3 × 6H), 3.57 (s, br., 3 × 2H), 6.72 (s, br. 3 × 8H, A), 7.05 (s, br., 3 × 8H, B), 7.18–7.30 (m, 3 × 12H), 7.82 (s, br. 3 × 8H), 8.40 (s, br., 3 × 4H, B), 8.59 (s, br., 3 × 4H, A), 8.75 (s, br., 3 × 4H, B), 8.87 (s, br., 3 × 4H, A), 10.47 (s, br., 3 × 4H). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, some of the peaks cannot be shown clearly due to limited solubility, δ): 19.85, 19.90, 40.29, 40.51, 49.19, 49.31, 122.97, 123.00, 127.29, 127.35, 128.26, 128.33, 128.46, 128.78, 130.11, 137.01, 139.29, 144.82, 144.89, 150.43, 151.16, 151.30. HRMS (ESI) calcd. for C₁₃₅H₁₂₀N₁₈La₂O₂₁S₃F₉ [M-3OTf]³⁺: 957.8668, found 957.8670. Calculated for C138H120N18O30La2F18S6·4H2O: C, 48.83; H, 3.80; N, 7.43%; Found: C, 48.02; H, 3.73; N, 7.30 %; [La₂(L2^{ss})₃] was synthesized, following the procedure for [La2(L2RR)3] with the use of L2SS instead, in 85% yield (0.052 g, 0.016

mmol): ¹H NMR (400 MHz, CD₃CN, 298 K, some of the signals are shown in two sets of peaks, **A** and **B**, respectively in ~1:1.1 ratio, δ): 1.02 (s, br., $3 \times 6H$, A), 1.16 (s, br., $3 \times 6H$, B), 2.77 (s, br., $3 \times 2H$, B), 2.93 (s, br., $3 \times 2H$, A), 3.39 (s, br., $3 \times 6H$), 3.57 (s, br., $3 \times 2H$), 6.72 (s, br. $3 \times 8H$, A), 7.04 (s, br., $3 \times 8H$, B), 7.17–7.39 (m, $3 \times 12H$), 7.82 (s, br. $3 \times 8H$), 8.40 (s, br., $3 \times 4H$, B), 8.58 (s, br., $3 \times 4H$, A), 8.75 (s, br., $3 \times 4H$, B), 8.89 (s, br., $3 \times 4H$, A), 10.47 (s, br., $3 \times 4H$). ¹³C NMR (100.6 MHz, CD₃CN, 298 K, some of the peaks cannot be shown clearly due to limited solubility, δ): 19.87, 40.24, 40.49, 49.14, 49.37, 122.91, 123.05, 127.32, 128.28, 128.49, 128.76, 130.14, 134.20, 137.02, 139.36, 144.86, 150.44, 151.33, 168.23, 168.41. HRMS (ESI) calcd. for C₁₃₅H₁₂₀N₁₈La₂O₂₁S₃F₉ [M-3OTf]³⁺: 957.8668, found 957.8662. Calculated for C₁₃₈H₁₂₀N₁₈O₃₀La₂F₁₈S₆·4H₂O: C, 48.83; H, 3.80; N, 7.43%; Found: C, 48.00; H, 3.78; N, 7.31 %.

3{*N*,*N*[']-(biphenyl-4,4'-diyl)bis[6dicarboxamide]}·2Gd·6(CF₃SO₃) [Gd₂(L2^{SS})₃]



To a white suspension of $L2^{ss}$ (0.050 g, 0.056 mmol, 1.5 equiv.) in a mixture of 6 mL of DCM/MeOH (12:1, v/v), a solution of Gd(CF₃SO₃)₃ (0.028 g, 0.047 mmol, 1 equiv.) in 8 mL of MeCN was added. The suspension was changed to yellow turbidity immediately. The reaction mixture was then refluxed for 16 h and the solid progressively dissolved to give a resulting homogeneous yellow solution. The solvent was removed under reduced pressure. The solid was then washed with DCM to give the desired product.

 $[Gd_2(L2^{ss})_3]$: (0.071 g, 0.021 mmol, 90% yield) HRMS (ESI) calcd. for C₁₃₅H₁₂₀N₁₈Gd₂O₂₁S₃F₉ [M-3OTf]³⁺: 970.5464, found 970.5457.

2a.4.2 X-ray crystallography



Chart S2a-1. Selected atomic numbering scheme of $[Eu_2(L1^{SS})_3](CF_3SO_3)_6$ in strand 1(top), 2 (middle), and 3 (bottom) for X-ray crystallography. The corresponding hydrogen atoms, H(number)A, with the same number of the attached carbons are not shown.

			Distances(Å)	
	Distance(Å)	Strand 1	Strand 2	Strand 3
Eu(1)–N		N(2), 2.566(5)	N(8), 2.537(5)	N(14), 2.595(5)
Eu(1)–O		O(2), 2.438(5)	O(5), 2.404(5)	O(9), 2.380(5)
Eu(1)–O		O(1), 2.438(5)	O(6), 2.407(5)	O(10), 2.424(4)
Eu(2)–N		N(5), 2.557(6)	N(11), 2.532(5)	N(17), 2.553(5)
Eu(2)–O		O(3), 2.435(4)	O(7), 2.434(4)	O(11), 2.441(5)
Eu(2)–O		O(4), 2.427(5)	O(8), 2.409(5)	O(12), 2.429(5)
Eu(1)-Eu(2)	15.055(1)			
		Bite angles		
	angles(°)			angles(°)
O(1)-Eu(1)-N(2)	62.96(17)	O(4)-	-Eu(2)–N(5)	63.87(17)
O(5)–Eu(1)–N(8)	63.87(18)	O(8)–	Eu(2)–N(11)	62.82(16)
O(9)-Eu(1)-N(14)	63.70(16)	O(12)-	-Eu(2)–N(17)	62.82(18)

Table S2a-1. Selected structural parameters for [Eu₂(L1^{SS})₃](CF₃SO₃)₆

N(2)-Eu(1)-O(2) 62	.92(16)	N(5)–Eu(2)–C)(3)	67.	17(18)
N(8)-Eu(1)-O(6) 62	77(17)	N(11)-Eu(2)-0	D(7)	63.	47(16)
N(14)-Eu(1)-O(1	0) 62	81(16)	N(17)-Eu(2)-O(11)		64.	06(17)
O(1)-Eu(1)-O(5) 77	77.70(16)		O(4)–Eu(2)–O(8)		17(17)
O(5)–Eu(1)–O(9) 75	75.94(16)		O(8)–Eu(2)–O(12)		51(16)
O(9)-Eu(1)-O(1) 75	.64(16)	O(12)-Eu(2)-O(4)		74.	20(16)
N(2)–Eu(1)–N(8)) 119	9.46(17)	N(5)–Eu(2)–N	(11)	122	.53(18)
N(8)–Eu(1)–N(14) 12:	1.88(17)	N(11)-Eu(2)-N(17)		117	.94(17)
N(14)-Eu(1)-N(2	2) 11	7.61(15)	N(17)–Eu(2)–N(5)		119	.00(18)
O(2)–Eu(1)–O(6) 76	5.01(16)	O(3)–Eu(2)–C)(7)	76.	01(14)
O(6)–Eu(1)–O(10)) 77	.39(15)	O(7)–Eu(2)–O	(11)	73.	34(15)
O(10)–Eu(1)–O(2	?) 76	5.41(16)	O(11)–Eu(2)–(D(3)	73.	63(15)
		Torsiona	l angles			
Strand 1	angles(°)	Strand 2	angles(°)	Stra	ind 3	angles(°)
N(2)–C(6)–C(7)– O(12)	- 22.97(103)	N(8)–C(32)– C(33)–O(6)	- 19.52(107)	N(14)- C(59) [.]	-C(58)– –O(10)	- 18.38(100)
C(6)–C(7)–N(3)– C(8)	- 175.63(70)	C(32)–C(33)– N(9)–C(34)	- 171.78(72)	C(58)- N(15)	-C(59)— —C(60)	- 164.52(72)
C(7)–N(3)–C(8)– C(9)	- 13.31(135)	C(33)–N(9)– C(34)–C(35)	160.48(77)	C(59)- C(60)	-N(15) C(61)	166.37(76)
C(11)–C(13)– C(14)–C(15)	150.24(75)	C(37)–C(39)– C(40)–C(41)	- 44.15(102)	C(63)- C(66)	-C(65)— —C(67)	- 35.42(103)
C(17)–C(19)– N(4)–C(20)	-5.56(126)	C(43)–C(45)– N(10)–C(46)	7.90(115)	C(69)- N(16)	-C(71)– –C(72)	-8.10(104)
C(19)–N(4)– C(20)–C(21)	- 167.72(71)	C(45)–N(10)– C(46)–C(47)	- 176.32(69)	C(71)- C(72)	-N(16)– –C(73)	- 167.21(59)
O(3)–C(20)– C(21)–N(5)	-8.27(99)	O(7)–C(46)– C(47)–N(11)	- 21.38(103)	O(11)- C(73)·	-C(72)- -N(17)	-17.69(92)
C–H…C	Dista	nce(Å) of (H…C)	Distance(Å)	of (C…C)	Angles(°) of (C–H–C)
C38–H38A…C1	0	2.987(8)	3.800(2	11)	14	6.81(51)
C64–H64A…C3	5	2.874(8)	3.72	8(11)	15	3.19(48)
C11–H11A…C6	1	2.908(8)	3.702(2	12)	14	4.23(56)
C67–H67A…C1	8	2.897(5)	3.78	8(9)	16	0.92(48)
C15–H15A…C4	4	3.154(6)	3.997(2	10)	15	1.76(44)
C41–H41A…C7	0	2.989(9)	3.851(2	12)	15	4.92(50)

Table	S2a-2.	Crystal	data	and	structure	refinement	of	[Eu ₂ (L1 ^{SS}) ₃](CF ₃ SO ₃) ₆ ,
[Eu ₂ (L2	2 ^{RR})₃](CF	₃ SO₃) 6, a	nd L2 ^{RI}	R				

	[Eu2(L1 ^{SS})3](CF3SO3)6,	[Eu2(L2 ^{RR})3](CF3SO3)6	L2 ^{RR}
Empirical formula	C ₁₂₆ H ₁₀₈ Eu ₂ N ₁₈ O ₁₂	C ₁₃₂ H ₁₂₀ Eu ₂ N ₁₈ O ₁₂	C44 H40N6O4
Formula weight	2370.22	2454.38	716.82
Temperature	296(2) K	296(2) K	296(2) K
Wavelength	0.71073 Å	0.71073 Å	0.71073 Å
Crystal system, space group	Orthorhombic, P 21 21 2	Triclinic, P-1	Monoclinic, P 2 ₁
Unit cell dimensions	$\begin{array}{l} a = 26.0606(10), \qquad b = \\ 28.4219(9), \qquad c = \\ 23.3299(9), \qquad \alpha = \beta = \gamma \\ = 90 \end{array}$	$\begin{array}{ll} a = 16.6790(5), & b = \\ 20.3099(6), & c = \\ 24.4218(8), & \alpha = \\ 87.025(2), & \beta = \\ 70.961(2), & \gamma = \\ 89.269(2) \end{array}$	$\begin{array}{ll} a = 10.3285(6), & b = \\ 17.8133(12), & c = \\ 10.5822(7), & \alpha = 90, \\ \beta = 100.506(4) & \alpha = \\ 90 \end{array}$
Volume	17280.3(11) Å ³	7809.7(4) Å ³	1914.3(2) Å ³
Z, Calculated density	4, 0.911 mg/m ³	2, 1.044 mg/m ³	2, 1.244 mg/m ³
Absorption coefficient	0.765 mm ^{−1}	0.849 mm ^{−1}	0.081 mm ⁻¹
F(000)	4848	2520	756
Crystal size	0.20 x 0.46 x 0.48 mm	0.04 x 0.16 x 0.24 mm	0.04 x 0.36 x 0.56 mm
θ range for data collection	2.60 to 25.35 deg	2.58 to 26.37 deg	2.78 to 24.71 deg
Limiting indices	-31 ≤ <i>h</i> ≤ 24, -34 ≤ <i>k</i> ≤ 34, -28 ≤ <i>l</i> ≤ 28	$-20 \le h \le 20, -25 \le k$ $\le 25, -30 \le l \le 30$	$-12 \le h \le 12, -20 \le k$ $\le 20, -12 \le l \le 12$
Reflections collected / unique	144075/31600	196288/31901	23536 / 6477
Absorption correction	Semi-empirical from equivalents	Semi-empirical from equivalents	Semi-empirical from equivalents
Refinement method	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²
Data/restraints/ parameters	31600 / 0 / 1261	31901 / 6 / 1161	6477 / 2 / 487
Goodness-of-fit on F ²	1.010	0.952	1.021

Final R indices [I>2σ(I)]	$R_1 = 0.0607, \qquad wR_2 = 0.0986$	$R_1 = 0.0743, \qquad wR_2 = 0.1956$	$R_1 = 0.0583, \qquad wR_2 = 0.1407$
R indices (all data)	$R_1 = 0.1062, \qquad wR_2 = 0.1073$	$R_1 = 0.1465, \qquad wR_2 = 0.2181$	$R_1 = 0.1163, \qquad wR_2 = 0.1712$
Flack parameter	0.018(8)	N/A	0(2)
Extinction coefficient	N/A	N/A	0.0054(14)
Largest diff. peak and hole	0.749 and -0.355 Å ⁻³	1.034 and -0.622 Å ⁻³	0.253 and -0.193 Å ⁻³

Note: All data were collected at ambient temperature. [Eu₂(L1^{ss})₃](CF₃SO₃)₆, and [Eu₂(L2^{RR})₃](CF₃SO₃)₆ were refined using PLATON/SQUEEZE, as such parameters such as empirical formula, formula weight, calculated density, etc. only reflect the refined structure as is.

Crystal data for Tb₂(L1^{ss})₃: *formula*, $M_r = 2372.04$, *crystal system*, space group P 2₁ 2₁ 2, Z = 4, a = 26.075(3) Å, b = 28.363(4)Å, c = 23.396(3)Å, $a = 90^{\circ}$, $\theta = 90^{\circ}$, $\gamma = 90^{\circ}$, V = 17304(4) Å³, μ (Mo_{Ka}) = 0.857 mm⁻¹, $\rho_{calc} = 0.911$ mgm⁻³, T = 296(2) K. The crystallographic data for the structural analyses have been deposited with the Cambridge Crystallographic Data Centre, CCDC No. 1009616, and the data can be obtained free of charge via <u>www.ccdc.cam.ac.uk/data_request/cif</u>.

Information	Identifier from cif file
Empirical formula	$C_{126}H_{96}N_{18}O_{12}Tb_2$
Formula weight	2372.04
Temperature/K	296(2)
Crystal system	orthorhombic
Space group	P21212
a/Å	26.075(3)
b/Å	28.363(4)
c/Å	23.396(3)
α/°	90
β/°	90

γ/°	90
Volume/Å ³	17304(4)
Z	4
ρ _{calc} g/cm ³	0.911
µ/mm ⁻¹	0.857
F(000)	4816.0
Crystal size/mm ³	0.480 × 0.320 × 0.200
Radiation	ΜοΚα (λ = 0.71073)
20 range for data collection/°	5.202 to 50.7
Index ranges	-29 ≤ h ≤ 31, -34 ≤ k ≤ 29, -28 ≤ l ≤ 28
Reflections collected	210003
Independent reflections	31661 [R _{int} = 0.0893, R _{sigma} = 0.0808]
Data/restraints/parameters	31661/17/1363
Goodness-of-fit on F ²	0.941
Final R indexes [I>=2 σ (I)]	R ₁ = 0.0524, wR ₂ = 0.1159
Final R indexes [all data]	R ₁ = 0.0774, wR ₂ = 0.1240
Largest diff. peak/hole / e Å ⁻³	0.59/-0.33
Flack parameter	0.002(5)

Th. (1 255).	Shape measure (°)				
TD2(LZ ~)3	Tricapped trigonal prism (D _{3h})	Capped square antiprism (C _{4v})			
Tb1	7.04	10.89			
Tb2	6.77	10.43			

Table S2a-3. Results of the Shape Analysis for $Tb_2(L1^{ss})_3$ helix.

Shape analysis was performed by the software developed by Raymond's group.^{24, 25}. The coordination geometry of the $Tb_2(L1^{ss})_3$: helix can be best described as tricapped trigonal prism.

2a.4.3 Photophysical measurement

Table S2a-1. A summary of selected photophysical properties, UV-Vis absorption and luminescence data, of $[Eu_2(L)_3](CF_3SO_3)_6$ in acetonitrile solution^a

λ_{abs} ε λ_{em} ι		$\lambda_{ m abs}{}^{ m max}$	${m arepsilon}^{\sf max}$	$\lambda_{em}{}^{max}$	τ
--	--	-------------------------------	---------------------------	------------------------	---

	(nm)	(L·mol ^{−1} ·cm [−]	(nm)	(ms)
		¹)		
[Eu ₂ (L1 ^{RR}) ₃](CF ₃ SO ₃) ₆	340	102300	616	0.22
[Eu ₂ (L1 ^{SS}) ₃](CF ₃ SO ₃) ₆	340	107000	616	0.22
[Eu ₂ (L2 ^{RR}) ₃](CF ₃ SO ₃) ₆	340	99000	616	0.22
[Eu ₂ (L2 ^{SS}) ₃](CF ₃ SO ₃) ₆	340	98000	616	0.22
[La ₂ (L1 ^{SS}) ₃](CF ₃ SO ₃) ₆	338	103000	449 ^b	0.014 ^b

^aUsing a 1mm cuvette and filter 455 nm. ^bMeasurement performed at 77 K in 1:4 of MeOH/EtOH and using a 10 mm cuvette.

2a.5 References

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Chapter 2b: Helicate-to-tetrahedron transformation of chiral lanthanide supramolecular complexes

2b.1 Introduction

2b.1.1 Induced structural change of supramolecules

Self-assembly supramolecular complexes have been proven to be an excellent candidate for various application such as catalysts^{1,2} and luminescent sensors.^{3,4} The applications of these self-assembled complex are all structural dependent and indepth investigation of ligand design principle was performed.⁵ Beyond the ligand design principles, understanding the factors that affect the structural formation of self-assembled complexes is important for engineering functional supramolecular complexes. In terms of lanthanide systems, recent research has been demonstrated that the formation of supramolecular structures is governed by five factors including concentration,^{6,7} anion and solvent,^{8,9} ionic radii of cation,^{10,11} stoichiometric ^{ratio12-14} and UV.¹⁵ This chapter will focus on concentration- and cationic radii dependent supramolecular formation.

2b.1.2 Concentration-dependent supramolecular formation

Generally, lanthanide ions coordinate with organic ligands by weak noncovalent interaction. Upon self-assembly of lanthanide ions with ligands, different supramolecular products can co-exist in solution state if there is no clear thermodynamic preference. Due to the labile feature of lanthanides, the supramolecular products can be changed from one to another depending on the external environment. In the case of concentration effect, increasing either complex or ligand concentration can promote the formation of higher order supramolecular products due to the aggregation of building blocks.¹⁶ An example is shown in figure 2b-1. Ln₂L₃ helicate was formed at low ligand concentration (6mM). When ligand concentration was increased to 48 mM, higher ordered Ln₄L₆ tetrahedron was formed.¹⁷



Figure 2b-1. Concentration supramolecular formation reported by Sun's and co-workers.¹⁷

2b.1.3 Cationic radii-dependent supramolecular formation

Unlike preorganized ligand such as cyclen, the coordinating sites of dipodal ligands and tripodal ligands are not well defined.¹⁸⁻²⁰ Slight change in reaction condition may lead to a different supramolecular product. As mentioned in chapter 1,

lanthanide ions are considered as having similar ionic radii and thus should exhibited similar reactivity profile. However, recent research has demonstrated that the cationic radii of lanthanide ions would affect the self-assembly process. Differences in ionic radii may affect the binding between ligands and metal ions, leading to different supramolecular products. One of the examples was shown in figure 2b-2. Coordination of ligand with light lanthanide led to the formation of cubane core, while the use of heavy lanthanide will lead to the formation of dimer core.²¹



Figure 2b-2. Cationic radii-dependent supramolecular formation reported by our group.²¹

<u>2b.1.4 Scope of study</u>

Recent research has reported that the offset distance of coordination units leads to an influence on the supramolecular formation. In this chapter, two pairs of new *C*₂-symmetric bis-tridentate ligands **L3** and **L4** were designed and synthesized. The offset distance of the coordination units in **L1**, **L3** and **L4** remained unchanged, and the effect of linker length was investigated. By varying the length of the linker, supramolecular transformation was observed for **L3**. Effect of concentration and cationic radii towards supramolecular formation will be also examined.



Figure 2b-3. Supramolecular formation of L1, L2 and L3.

2b.2 Result and discussion

2b.2.1 Design and synthesis of L3 and L4

In the previous chapter, **L1** was found to form enantiomeric pure lanthanide bimetallic triple helicate induced by point chirality effect. Based on this result, the length of the central bridging linker was varied and the coordinating sites was kept to the same. The structures of ligands were shown in figure 2b-4.



Figure 2b-4. Ligand structures of L1, L3 and L4.

Similar to the design of L1, pyridine-2,6-dicarboxamide (pcam) moieties were employed as coordinating units in L3 and L4, which tends to form stable ninecoordinated lanthanide complexes.²² The shorter ligand L3 was prepared by connecting two chiral metal chelating pcam moieties with a monophenyl linker. For L4, a longer triphenyl linker was employed. Compare the ligand design of L1 with L3 and L4, the offset distance of two chelating moieties did not change and we expected that both L3 and L4 could also form stable Ln₂L₃ (L = L3 and L4) helicates. Following the previous procedures, two pairs of ligands L3^{R/S} and L4^{R/S} were prepared in two steps using generic HATU peptide coupling reactions and were fully characterized.

2b.2.2 Conformation of metal-to-ligand ratio

The metal-to-ligand ratio is essential for complexation and it can be determined by ¹H NMR and UV-CD titration. Poor ligand solubility was the major challenge when performing titration as it is only soluble in a mixture of chloroform and methanol. Addition methanol may cause dissocation of lanthanide complex. Therefore, ¹H NMR and UV-CD titration were done by using a mixture of CHCl₃/MeOH/MeCN in a ratio of 50:5:45 and 12:1:12 respectively to minimize the effect brought by methanol.

¹H NMR and UV-CD titration were performed by adding metal into a solution of ligand and then observed the changes. For ¹H NMR titration (figure 2b-5), the ¹H NMR signal at 1.7 ppm, which corresponds to the CH₃ group of the ligand, decreased gradually upon addition of Eu(OTf)₃ to a solvent mixture containing **L3** and a new siganl at 2.1 ppm in the aliphatic region increased progressively. When 0.67 eq. of Eu(OTf)₃ was added, all the ligand peaks disappeared when confirmed the metal-to-ligand ratio was 2 to 3.



Figure 2b-5. ¹H NMR titration of L1^{ss} (1.09 x 10⁻³M in 50:5:45, v/v/v of CDCl₃/CD₃OD/CD₃CN) with Eu(OTf)₃ (0.108M in CD₃OD) at 298 K.

Similar result was also observed for UV-Vis titration. UV-Vis titration for **L3** was performed in a solvent mixture of CHCl₃/MeOH/MeCN (12:1:12, v/v/v). Upon addition of Eu(OTf)₃ to the ligand solution **L3**, a progressive decrease of 281, 315 and 353 nm was observed which corresponded to the ligand absorption (figure 2b-6). As evolution of complex absorption at 320 nm was observed simultaneously. A plot of changes in molar absorptivity at four wavelengths reveal an end point of 0.67.



Figure 2b-6. UV-Vis titration of **L3** with $Eu(OTf)_{3.}(a)$ Variation in UV-Vis absorption spectra of titrating **L3**^{SS} (2.79 x 10⁻⁴M, in 12:1:12, v/v/v, of CHCl₃/MeOH/CH₃CN) with $Eu(OTf)_{3}$ (0.036M in MeOH) at 298K (Eu:L3^{SS} = 0.0–2.0). (b) Variation of absorbance at four different wavelengths upon titrating L3^{SS} with

Eu(OTf)₃.

Given that **L4** exhibit similar ligand design and coordinating site, a hypothesis of metal-to-ligand ratio 2 to 3 was also investigated by ¹H NMR and UV-CD titration. For ¹H NMR titration, the signal at 5.3 ppm corresponding to the chiral proton of the ligand was monitored (figure 2b-7). Upon addition of Eu(OTf)₃ to a solution of **L4**, the proton signal at 5.3 ppm decreased gradually and a A new chiral proton signal at 5.5 ppm increased progressively. When 0.67 eq. of Eu(OTf)₃ was added, all the ligand signal disappeared which confirmed the metal-to-ligand ratio is 2 to 3. Similar result was also observed for UV-Vis titration and an end point of 0.67 was observed (figure 2b-8).



Figure 2b-7. ¹HNMR titration of L4^{ss} (1.09 x 10^{-3} M in 50:5:45, v/v/v of CDCl₃/CD₃OD/CD₃CN) with

Eu(OTf)₃ (0.108M in CD₃OD) at 298 K.



Figure 2b-8. UV-Vis titration of L4 with $Eu(OTf)_{3.}(a)$ Variation in UV-Vis absorption spectra of titrating L4^{SS} (1.96 x 10⁻⁴M, in 12:1:12, v/v/v, of CHCl₃/MeOH/CH₃CN) with $Eu(OTf)_{3}$ (0.014M in MeOH) at 298K (Eu:L4^{SS} = 0.0–2.0). (b) Variation of absorbance at four different wavelengths upon titrating L4^{SS} with $Eu(OTf)_{3.}$

2b.2.3 Synthesis and characterization of [Eu₂L3₃] helicates

The Eu metal complex was synthesized by treating 8 x 10^{-3} M ligand with 0.67 equivalents of Eu(OTf)₃ in a mixed solution of CHCl₃/MeOH/CH₃CN (12/1/12, v/v/v).By comparing the ¹H NMR of free ligand, most signals arising from the europium complexes are shifted due to the formation of paramagnetic europium complex. Interestingly, two sets of signals in a ratio of ~96:4 was observed and the signal of minor species disappeared after dissolution in CD₃CN for three days (figure 2b-9).



Figure 2b-9. (a) ¹H NMR of crude Eu complex mixture containing both of major species and minor species in a ratio of 96:4. (b) ¹HNMR showing pure major species by dissolving crude mixture after 3 days.

Several investigations were done to promote the formation of either minor species or major species including solvent effect, counterion effect and concentration effect. For solvent effect, the role of methanol in complexation was investigated. Since methanol is well known to compete with ligands for lanthanide ions during complexation, the self-assembly process may be affected. Therefore, complexation was done without the use of methanol. ¹H NMR result showed that the addition of methanol was not necessary as the it did not affect the result of complexation (figure 2b-10). Lanthanide complexes was found to be stable in 4% methanol without dissociation. To keep consistent to the complexation condition of ¹H NMR titration, methanol was added in further preparation of different lanthanide complexes.



Figure 2b-10. ¹*HNMR of crude Eu complex mixture prepared using CD₃Cl and CD₃CN as solvent.*

Triflate ion was replaced by chloride and perchlorate to investigate the counterions effect. Similar result was also observed when counterion was changed to chloride and perchlorate and Eu₂L3₃ was formed as major species. However, both chloride and perchlorate ion were not suitable for further experiment due to poor stability of resulting complexes. For perchlorate ion, the water content of the stock Europium perchlorate solution is too high (50%) and high-water content interfered the complexation. Free ligand signals were observed in ¹H NMR spectrum (figure 2b-11). For chloride ion, the complex was only soluble in methanol and dissociation of complex was found in ESI-HRMS (figure 2b-12).



Figure 2b-11. ¹*HNMR of* [*Eu*₂*L3*₃](*ClO*₄)₆.



Figure 2b-12. (a) ¹HNMR and (b) ESI-HRMS of [Eu₂L**3**₃]Cl_{6.}

Ligand concentration was found to have a great influence towards the supramolecular formation of L3. In a lower ligand concentration (1 x 10^{-3} M), only single species was observed in ¹H NMR (figure 2b-13). When the ligand concentration increased, another minor species was observed as evidenced by another ¹H NMR titration, in which the minor species can be observed when 2.15 x 10^{-3} M of ligand was titrated with Eu(OTf)₃ (figure 2b-14).



Figure 2b-13. ¹HNMR of (a) crude Eu complexes prepared by 0.008M L3 and (b) 0.0001M L3 in

CDCl₃/MeOD/CD₃CN (12/1/12, v/v/v).



Figure 2b-14. ¹HNMR titration of **L3** (2.15 x 10⁻³M in 70:5:25, v/v/v of CDCl₃/CD₃OD/CD₃CN) with Eu(OTf)3 (0.09M in CD₃OD) at 298 K.

Due to the poor ligand solubility, attempt to prepare pure minor species was not successful. The high ligand concentration was then mimicked by dissolving Eu complexes and observed the ¹H NMR. Generally, increasing the Eu complex concentration will lead to an increase in formation of minor species. The maximum ratio of major species to minor species was determined to be 2.9:1 when ligand concentration is 88.9 mM. Further increase in ligand concentration will not affect the ratio significantly (figure 2b-15).



Figure 2b-15. ¹HNMR spectra of [Eu major species]/[Eu minor species] at different ligand concentration in CD₃CN at 298 K.

The metal-to-ligand ratio of major species was also confirmed by ESI-HRMS. From the ESI-HRMS spectrum, only single set of independent signal series was observed corresponding to the dinuclear species [Eu₂L3₃]. Three different clusters of signals can be observed which correspond to three different charged [Eu₂L3₃] species (+2, +3 and +4). Each signal can be assigned to the charged species with the loss of either triflate ion or proton. For example, signal with peaks with *m/z* equal to 1369.1962, 1294.2137 and 1219.2385 can be assigned to the species of [{[Eu₂L1^{SS}₃](OTf)_m}-H_n]²⁺ with m = 4, n = 0; m = 3, n = 1; m = 2, n = 2. The assignments for each signal were also further confirmed by comparing the corresponding experimental isotopic pattern to the stimulated isotopic pattern.

2b.2.4 Synthesis and characterization of [Sm₂L3₃], [Gd₂L3₃] and [Tb₂L3₃]

Sm was used to replace Eu for preparing $[Sm_2L3_3]$ complexes. By using the same protocol, similar result was observed. The ¹H NMR showed the existence of a major species and a minor species in a ratio of 94:6 and the minor species was found to undergo transformation to the major species after 3 days (figure 2b-16). The ESI-HMRS of the Sm major species showed only single set of independent peak series, corresponding to the $[Sm_2L3_3]$ complex with different charges. The preparation of $[Gd_2L3_3]$ and $[Tb_2L3_3]$ complexes were done by treating 1 x 10⁻³ M ligand with the correspond Gd(OTf)₃ and Tb(OTf)₃. Characterization by NMR is not possible due to the strong paramagnetic nature. The ESI-HRMS revealed single species corresponding to $[Gd_2L1_3]$ and $[Tb_2L1_3]$ with different clusters corresponding to the progressive loss of triflate anion and proton.



Figure 2b-16. (a) ¹H NMR of crude Sm complex mixture containing both of major species and minor species in a ratio of 94:6. (b) ¹HNMR showing pure major species by dissolving crude mixture after 3

days.

2b.2.5 Synthesis and characterization of [La₂L3₃] and [Lu₂L3₃]

The corresponding La and Lu complexes were synthesized by treating 2 eq. of lanthanide ions to 3 eq. of L3. For La complexes, the crude NMR showed two different sets of signals corresponding to a major and minor species. However, unlike the minor species of Eu complex, which transformed to major species slowly upon dilution (figure 2b-17), the La minor species exhibited a fast transformation upon dilution (figure 2b-18).



Figure 2b-17. ¹HNMR of (a) crude Eu complexes prepared by 0.006M L3 in CD₃CN and (b) subjected to 5-fold dilution and performed ¹HNMR instantaneously.



Figure 2b-18. ¹HNMR of (a) crude La complexes prepared by 0.006M L3 in CD₃CN and (b) subjected to 5-fold dilution and performed ¹HNMR instantaneously.

For Lu complexes, insignificant amount of minor species was observed when 0.008M L3 was treated with Lu(OTf)₃. Similar with the result of Eu and Sm, pure major species can be obtained by decreasing the ligand concentration to 0.001M (figure 2b-19). The metal-to-ligand ratio of both La and Lu complexes were even evidenced by ESI-HRMS in which only $[Ln_2L3_3]$ were shown in the spectrum and no other supramolecular species can be observed.



Figure 2b-19. ¹HNMR of (a) crude Lu complexes prepared by 0.008M L3, (B) crude Lu complexes prepared by 0.001M L3.

<u>2b.2.5 Synthesis and characterization of [Ln₂L4₃]</u>

Following the previous synthetic procedures of $[Ln_2L1_3]$, the corresponding $[Eu_2L4_3]$ was prepared by addition of two equivalents $Eu(OTf)_3$ into the solution containing three equivalent of L4. Due to the paramagnetic europium ions, most NMR signal of the $[Eu_2L4_3]$ was shifted which indicated the successful coordination of europium ion to the ligand. The ¹H NMR displayed only single set of signals and the ESI-HRMS revealed one set of independent peak series, corresponding to the dinuclear species $[Eu_2L4_3]$. In the ESI-HRMS spectrum, $[Eu_2L4_3]$ exhibits three different clusters of peaks which correspond to three different species with +3 and +4 charged states. For example, peaks with m/z equal to 1015.2181, 965.2259, 915.2409 and 865.2620 can be assigned to the species of $[[Eu_2L4^{SS}_3](OTf)_m]-H_n]^{3+}$ with m = 3, n = 0; m = 2, n = 1; m = 1, n = 2; m = 0, n = 3. Other lanthanide ions were used to replace Eu for preparation of a series of $[Ln_2L4_3]$ including La, Sm, Gd, Tb and Lu. All the $[Ln_2L4_3]$ were successfully synthesized and well characterized by NMR and ESI-HRMS.

2b.2.6 Helicate-to-tetrahedron transformation

Single X-ray diffraction is a powerful tool to determine the exact structure of a supramolecular complex. Several attempts were done to prepare crystal of [Eu₂L3^{SS}₃] including evaporation and solvent diffusion. A single crystal of Eu complex was obtained by slow diffusion of diisopropyl ether into an acetonitrile solution of the complex. Similar result was observed when ether was used including methyl t- butyl ether, diethyl ether, diisopropyl ether. No crystal can be formed when ether was replaced by ethyl acetate, dichloromethane and acetone.

Single X-ray diffraction showed a $[Eu_4L3^{SS}_6]$ tetrahedral structure which confirmed a helicate-to-tetrahedron transformation through crystallization (figure 2b-20). In the crystal structure of $[Eu_4L3^{SS}_6]$ tetrahedron, each europium metal centre can be best described as tricapped trigonal prism. Four metal ions adopted the same Δ configurations, which shows the successful transfer of chiral information from the chiral moiety to the metal centre. In the tetrahedron structure, four europium ions occupied the vertices, and six ligands act as the edges. Eu-N distances range from 2.52(2) to 2.59(2) Å and Eu-O distances range from 2.38(2) to 2.44(2) Å.



Figure 2b-20. X ray crystal structure of [Eu₄L3^{ss}₆] showing a tetrahedron structure.

Further characterization of $[Eu_4(L3^{SS})_6]$ was done by washing the crystal with diisopropyl ether and then dried under vacuum. Compared with $[Eu_2(L3)_3]$ helicate, $[Eu_4(L3)_6]$ showed a very different ¹H NMR signal, which suggested that only pure $[Eu_4(L3)_6]$ in the solution (figure 2b-21). The integration of $[Eu_4(L3)_6]$ integrations are equivalent to the ligand in C_2 -symmetric nature which revealed the similar lanthanide geometry with $[Eu_2(L3)_3]$.



Figure 2b-21. ¹H NMR of (a) pure [Eu₂L1₃] and (b) pure [Eu₄L1₆] that was done simultaneously after dissolution of [Eu₄L1₆].

Comparing the ¹H NMR of $[Eu_2(L3)_3]$ with $[Eu_4(L3)_6]$, the peaks of $[Eu_4(L3)_6]$ are relatively broad (figure 2b-22) The broadening of peaks can be attributed to the pseudocontact shifts of paramagnetic europium ion. Since the pseudocontact shifts is governed by the distance between the paramagnetic metal ion and proton, it can be deduced that the proton in $[Eu_4(L3)_6]$ experienced a stronger paramagnetic shift. The chemical formulas of $[Eu_4(L3)_6]$ was further characterized by ESI-HRMS. In the ESI-HRMS spectrum, only single set of independent peaks was observed. Each cluster of peaks can be assigned to the tetranuclear species with the progressive loss of anions and protons. For example, peaks with m/z equal to 1065.3668, 1035.3778, 1005.3876 and 975.3884 can be assigned to the species of $[{[Eu_4L3^{SS}_6](OTf)_m}-H_n]^{5+}$ with m =7, n = 0; m = 6, n = 1; m = 5, n = 2; m = 4, n = 3. Upon dilution of $[Eu_4(L3^{SS})_6]$ in MeCN, the tetrahedral $[Eu_4(L3^{SS})_6]$ is unstable and will slowly rearranged back to $[Eu_2(L3^{SS})_3]$ in 11 days (figure 2b-23).



Figure 2b-22. Paramagnetic shift of [Eu₂(L3)₃] and [Eu₄(L3)₆] in CD₃CN.



Figure 2b-23. Tetrahedron-to-helicate transformation from $[Eu_4L3_6]$ to $[Eu_2L3_3]$ (4.53 x 10⁻⁴M in CD₃CN).

With this interesting helicate-to-tetrahedron transformation via ether crystallization, the effect of lanthanide ionic radii towards this transformation was further investigated. By replacing Eu to Sm for coordination, helicate-to-tetrahedron transformation was also observed. Single crystal of [Sm₄L3₆] was obtained upon slow diffusion of diisopropyl ether into a solution containing pure [Sm₂L3₃]. The crystal was then dried and characterized by NMR and ESI-HRMS. In the ¹H NMR spectrum, the [Sm₄L3₆] showed a different signal compared with that of [Sm₂L1₃] (figure 2b-24). Similar with [Eu₄L3₆], [Sm₄L3₆] is unstable upon dissolution in MeCN and will slowly transform back to [Sm₂L3₃] in 14 days.



Figure 2b-24. Chemical shift of $[Sm_2(L3)_3]$ and $[Sm_4(L3)_6]$ in CD₃CN.

By replacing Eu with Gd and Tb, similar helicate-to-tetrahedron was also observed as evidenced by ESI-HRMS. However, characterization by NMR is not possible due to the strong paramagnetic nature of Gd and Tb. Tetrahedron-to-helicate transformation process was monitored by dissolving the tetrahedron in MeCN and
wait for 14 days before performing ESI-HRMS. Result showed that that $[Gd_4L3_6]$ and $[Tb_4L3_6]$ are unstable upon dissolution in MeCN and will rearrange back to $[Gd_2L3_3]$ and $[Tb_2L3_3]$ respectively (figure 2b-25).



Figure 2b-25. ESI-HRMS of tetrahedron-to-helicate transformation after dissolution in CH₃CN for 14 days (a) [Gd₄L**3**₆] and (b) [Tb₄L**3**₆].

When Eu was replaced by Lu, [Lu₄L**3**₆] can be also prepared by helicate-totetrahedron transformation through ether crystallization. The chemical formula and structure were well characterized by NMR and ESI-HRMS. Similar with [Eu₄L**3**₆] and [Sm₄L**3**₆], the ¹H NMR of [Lu₄L**3**₆] also showed a very different signal compared with that of $[Lu_2L3_3]$ (figure 2b-26). Interestingly, different from that of $[Eu_4L3_6]$ and $[Sm_4L3_6]$, $[Lu_4L3_6]$ was found to be relatively stable and only insignificant amount of $[Lu_2L3_3]$ was found upon dissolution in MeCN after 10 days (figure 2b-27).



Figure 2b-26. Chemical shift of $[Lu_2(L3)_3]$ and $[Lu_4(L3)_6]$ in CD₃CN.



Figure 2b-27. ¹*H* NMR of $[Lu_4(L3)_6]$ upon dissolution in CD₃CN at different timespan.

When the metal source is changed from Eu to La, no crystal can be obtained by slow diffusion of ether into a solution containing [La₂L**3**₃]. The powder that obtained from diffusion of ether was then characterized by NMR. The ¹HNMR of the powder revealed that the La complex remained in the most stable form [La₂L**3**₃] helicate form and no other supramolecular complexes can be observed (figure 2b-28).



Figure 2b-28. ¹HNMR of (a) [La₂(**L3**)₃] in CD₃CN and (b) the powder left after diffusing ether into a solution containing [La₂(**L3**)₃].

2b.2.7 Analysis of the effect of ligand design and ionic radii

Result showed that the ligand design has a great influence towards the supramolecular formation as well as the helicate-to-tetrahedron transformation. In this study, both **L3** and **L4** were expected to form stable Ln_2L_3 helicate as the two metal ions shared the same C_3 axis upon coordination. According to the symmetry design principle that proposed by Raymond and co-workers, the use of longer and flexible central bridging linker should be avoided since it may increase the chance of forming

unwanted clusters.²³ In the previous chapter, diphenyl unit was used as central bridging linker in **L1** and **L2** and both can form stable Ln₂L₃ helicate. In this study, monophenyl unit was used as central bridging linker in **L3** and it is expected form Ln₂L₃ helicate since shorter linker may promote the formation of helicate without other unwanted supramolecular clusters. Interestingly, helicate-to-tetrahedron transformation was observed for **L3** and pure tetrahedron can be isolated through either crystallization. Apart from the offset distance of two chelating groups, this study showed that the distance of two metal ions also plays an important role in supramolecular formation.

The stability of the supramolecular complex also varies across the lanthanide series (figure 2b-29). Generally, lanthanide ions are considered as having similar physical and chemical properties due to their similar ionic radii. In this study, the stability and the supramolecular formation were governed by the ionic radii which is consistent with the work that reported by Hamacek and co-workers.²⁴ When the largest La ion was used, formation of stable $[La_2(L3)_3]$ helicate was highly preferred and labile $[La_4(L3)_6]$ tetrahedron can be only observed under high ligand concentration. Stable helicate and tetrahedron can be formed when the smallest Lu ion was used. The lanthanide ions that lies between La and Lu (Sm, Eu, Gd and Tb) were found to form stable helicate at low concentration and stable tetrahedron at high ligand concentration.



Figure 2b-29. Summary of helicate-to-tetrahedron transformation governed by different lanthanide ions.

Kinetic studies were done to estimate the activation parameter of tetrahedron-to-helicate transformation process. Arrhenius plot was used since the tetrahedron-to-helicate transformation is not an equilibrium process. The transformation process was monitored by NMR and two experimental error were observed. Firstly, tetrahedron did not undergo transformation to helicate even heating at 338K. Moreover, the rate of conversion at 338K is slower than that of 328K in some of the trials. 8 trials were performed for Eu complex and 5 trials were performed for Sm complex. After omitting the outliner, the kinetic studies showed that both [Eu₄L₃₆] and [Sm₄L₃₆] exhibited similar activation energy which was 97.1 (5) and 85.2 (4) kJmol⁻¹ respectively due to the similar ionic radii of Eu and Sm. The positive value of ΔH_{298} also showed that the tetrahedron-to-helicate transformation is endothermic.

This study also revealed that the Ln_2L3_3 helicate is an essential intermediate for the formation of Ln_4L_6 tetrahedron. It was proposed that the ligand and lanthanide ion will first self-assembled to form the entropically favored helicate. Then the helicate will rearrange to form the Ln_4L3_6 tetrahedron under high ligand concentration. Crystallization process was proven to be an alternative way to mimic high ligand concentration environment. This hypothesis was supported by attempting to prepare Lu tetrahedron in a one pot self-assembly reaction. Despite varying the reaction condition such as temperature and solvent, direction formation of Lu tetrahedron was unsuccessful. All the tetrahedron seems to form by crystallization of a solution of helicate only.

2b.2.8 Photophysical studies

Photophysical measurement was done for the complex of **L3** and **L4**. To avoid tetrahedron-to-helicate transformation, all the [Ln₄**L3**₆] tetrahedron were performed within 1 hr after dissolution of sample. For UV-Vis absorption measurement, the λ_{max} of [Eu₂**L3**₃] was blueshifted for 19 nm compared with that of [Eu₄**L3**₆] (figure 2b-30).



Figure 2b-30. Normalized UV-Vis spectrum of $[Eu_2(L3^{SS})_3]$ and $[Eu_4(L3^{SS})_6]$ in CH₃CN.

The chiroptic properties of Eu complexes were examined by solution CD and CPL measurement. Similar with that of UV-Vis absorption, $[Ln_2L3_3]$ and $[Ln_4L3_6]$ (Ln = La, Eu and Sm) also showed slight differences in the CD absorption. For $[Ln_2L3_3]$, strong cotton effects were observed at 212, 240, 259, 273, 312 and 381 nm while for $[Ln_4L3_6]$,

strong cotton effects were observed at 212, 240, 259, 273, 312 and 362 nm. Mirror image of CD spectrum were observed for *R*- and *S*-isomers. For $[Ln_2L3_3]$ (Ln = La and Eu), strong Cotton effects were observed at 209, 240, 263, 290, 334 and 381 nm.

For CPL measurement, both $[Eu_2L3^{ss}_3]$ and $[Eu_4L3^{ss}_6]$ exhibited fairly strong CPL signal for the for the ${}^5D_0 -> {}^7F_1$ transition, with $g_{lum} = +0.10$ for both complexes. Weaker g_{lum} value of -0.02 was observed for the the ${}^5D_0 -> {}^7F_2$ transition. Mirror CPL spectrum was also observed for these Eu complexes (figure 2b-31a). The CD and CPL result were consistent with the diastereoselective and diastereoselective breaking supramolecular formation behaviour that reported in Chapter 2a.

The luminescent properties of $[Eu_2L3_3]$ and $[Eu_4L3_6]$ were examined. Upon excitation of $[Eu_2L3_3]$ at 311 nm, characteristics narrow Eu red emission lines at 595, 616, 688 and 697 nm were observed which indicated the successful sensitization of $Eu^{III 5}D_0$ excited state via antenna effect (figure 2b-31b). Similar result was observed upon excitation of $[Eu_4L3_6]$ at 330 nm. The relative quantum yield of $[Eu_2L3^{RR}_3]$ was determined to be 4.41% which is higher than that of $[Eu_4L3^{RR}_6]$ (0.83%).



Figure 2b-31. (a) Normalized CPL spectra of [Eu4L1^{RR/SS}₆] in MeCN. (b) Normalized excitation and

emission spectra of [Eu₄L1^{RR/SS}₆] in MeCN.

2b.3 Conclusion

In conclusion, this work demonstrated that the lanthanide self-assembly supramolecular formation is highly sensitive to the lanthanide ionic radii as well as the length of central bridging linker. The formation of higher ordered tetrahedron can be favored by varying the distance of two metal chelating groups. Result showed that the formation of helicates is favored when larger lanthanide ion was used, while smaller lanthanide ions tend to form tetrahedron. This work is important as it demonstrated two important parameters for designing supramolecular lanthanide edifices.

2b.4 Experimental section

2b.4.1 Synthesis

 $N^{2},N^{2'}-(1,4-phenylene)bis(N^{6}-((S)-1-phenylethyl)pyridine-2,6-dicarboxamide)$ (L3^{SS}) and $N^{2},N^{2'}-(1,4-phenylene)bis(N^{6}-((R)-1-phenylethyl)pyridine-2,6-dicarboxamide)$ (L3^{RR})



To a stirred solution of **(R)-6-((1-phenylethyl)carbamoyl)picolinic acid** (0.25 g, 0.92 mmol, 2.2 equiv.) in anhydrous DMF (4 mL) at room temperature, HATU (0.765 g, 2.01 mmol, 4.8 equiv.) was added under nitrogen. After allowing it to stir for 20 min, a 4,4'- diamino-p-terphenyl (0.045 g, 0.417 mmol, 1.0 equiv.) was added and the reaction mixture was allowed to stir for 20 min in dark. DIPEA (0.89 mL, 5.1 mmol, 12.5 equiv.) was then added to the reaction mixture and the resulting solution was stirred at room

temperature for 14 h. The reaction mixture was then diluted with H_2O (10 mL) and extracted with DCM (5×3 mL). After removing the organic volatile under reduced pressure, the residue was then washed with CH₃CN (10 mL), and fine powder was progressively precipitated out. Then the solid was collected by centrifugation and the desired compound was isolated. (L3^{RR}): (0.20 g, 0.33 mmol, 70% yield), ¹H NMR (400 MHz, $(CD_3)_2$ SO, 299 K, δ): 1.67 (d, J = 8 Hz, 6H), 5.35–5.45 (m, 2H), 7.24 (t, J = 8 Hz, 2H), 7.34 (t, J = 8 Hz, 4H), 7.45 (d, J = 8 Hz, 4H), 7.70 (s, 4H), 8.07 (t, J = 8 Hz, 2H), 8.35 (d, J = 8 Hz, 2H), 8.39 (d, J = 8 Hz, 2H), 9.27 (d, J = 8 Hz, 2H). ¹³C NMR (100 MHz, (CD₃)₂SO, 300 K, δ): 22.17(CH₃), 48.81(CH), 122.36, 125.41, 126.52, 127.28, 128.80, 134.71, 140.20, 144.52, 149.14, 149.42, 162.28, 163.39. HRMS (ESI) calcd. for C₇₂H₆₄N₁₂O₈Na [2L3^{RR}+Na]⁺: 1247.4862, found 1247.4850. (L3^{SS}) was synthesized, following the procedure for (L3^{RR}) with the use of (S)-6-((1-phenylethyl)carbamoyl)picolinic acid instead, in 60% yield (0.17 g, 0.28 mmol): ¹H NMR (400 MHz, (CD₃)₂SO, 299 K, δ): 1.67 (d, J = 8 Hz, 6H), 5.35–5.45 (m, 2H), 7.24 (t, J = 8 Hz, 2H), 7.34 (t, J = 8 Hz, 4H), 7.45 (d, J = 8 Hz, 4H), 7.70 (s, 4H), 8.07 (t, J = 8 Hz, 2H), 8.35 (d, J = 8 Hz, 2H), 8.39 (d, J = 8 Hz, 2H), 9.27 (d, J = 8 Hz, 2H). ¹³C NMR (100 MHz, (CD₃)₂SO, 300 K, δ): 22.15, 48.78, 122.35, 125.39, 125.58, 126.51, 127.26, 128.78, 134.71, 140.19, 144.51, 149.13, 149.41, 162.28, 163.20. HRMS (ESI) calcd. for C₇₂H₆₄N₁₂O₈Na [2L3^{SS}+Na]⁺: 1247.4862, found 1247.4850.

 N^{2} , $N^{2'}$ -([1,1':4',1''-terphenyl]-4,4''-diyl)bis(N^{6} -((R)-1-phenylethyl)pyridine-2,6-dicarboxamide) (L4^{RR}) and N^{2} , $N^{2'}$ -([1,1':4',1''-terphenyl]-4,4''-diyl)bis(N^{6} -((R)-1-phenylethyl)pyridine-2,6-dicarboxamide) (L4^{RR})



To a stirred solution of (R)-6-((1-phenylethyl)carbamoyl)picolinic acid (0.5 g, 1.85 mmol, 2.2 equiv.) in anhydrous DMF (8 mL) at room temperature, HATU (1.65 g, 4.06 mmol, 4.8 equiv.) was added under nitrogen. After allowing it to stir for 20 min, a 4,4'diamino-p-terphenyl (1.10 g, 4.21 mmol, 1.0 equiv.) was added and the reaction mixture was allowed to stir for 20 min in dark. DIPEA (1.78 mL, 10.2 mmol, 12.5 equiv.) was then added to the reaction mixture and the resulting solution was stirred at room temperature for 14 h. The reaction mixture was then diluted with H_2O (20 mL) and extracted with DCM (5 × 6 mL). After removing all of the organic volatile under reduced pressure, the residue was then washed with CH₃CN three times (20 mL x 3), and fine powder was progressively precipitated out. Then the solid was collected by filtration and the desired compound was isolated. (L4^{RR}): (0.62 g, 0.81 mmol, 95.28% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 299 K, δ): 1.69 (d, J = 6.96 Hz, 6H), 5.35–5.45 (m, 2H), 7.29 (t, J = 2.65 Hz, 2H), 7.39 (t, J = 8 Hz, 4H), 7.46 (d, J = 8 Hz, 4H), 7.66 (d, J = 8 Hz, 8H), 7.78 (d, J = 8 Hz, 4H), 8.09 (t, J = 8 Hz, 2H), 8.38 (d, J = 8 Hz, 2H), 8.44 (d, J = 8 Hz, 2H), 8.54 (d, J = 8 Hz, 2H), 10.01 (s, 2H). ¹³C NMR (101 MHz, 300 K, δ): ¹³C NMR (101 MHz, (CD₃)₂SO, 300K, δ): 163.11, 162.22, 149.59, 149.14, 144.60, 140.23, 138.74, 137.85, 135.95, 128.83, 127.29, 126.62, 125.66, 125.40, 122.16, 48.68, 22.28. HRMS (ESI) calcd. for C₄₈H₄₀N₆O₄Na [L4^{RR}+Na]⁺: 787.3003, found 787.3038. (L4^{SS}) was synthesized, following procedure for (L4^{RR}) with the the use of (S)-6-((1phenylethyl)carbamoyl)picolinic acid instead, in 80% yield (0.4 g, 0.52 mmol): ¹H NMR (400 MHz, (CD₃)₂SO, 299 K, δ): 1.72 (d, J = 8 Hz, 6H), 5.40–5.48 (m, 2H), 7.30 (t, J = 8

Hz, 2H), 7.41 (t, *J* = 8 Hz, 4H), 7.49 (d, *J* = 8 Hz, 4H), 7.70 (d, *J* = 8 Hz, 8H), 7.83 (d, *J* = 8 Hz, 4H), 8.13 (t, 2H), 8.40 (d, *J* = 8 Hz, 2H), 8.47 (d, *J* = 8 Hz, 2H), 8.77 (d, *J* = 8 Hz, 2H). ¹³C NMR (100 MHz, (CD₃)₂SO, 300 K, δ): 163.11, 162.22, 149.61, 149.15, 144.60, 140.23, 138.75, 137.86, 135.95, 128.83, 127.29, 126.62, 125.65, 125.40, 122.15, 48.67, 22.28. HRMS (ESI) calcd. for C₄₈H₄₀N₆O₄Na [**L4**^{RR}+Na]⁺: 787.3003, found 787.3038.

General synthetic procedures of [Ln₂L3₃]



To a white suspension of **L3** (10 mg, 0.016 mmol, 1.5 equiv.) in a mixture of 8.49 ml of DCM/MeOH (12:1, v/v), a solution of $Ln(OTf)_3$ (0.011 mmol, 1 equiv.) (Ln = La, Sm, Eu, Gd, Tb and Lu) in 7.83 ml of CH₃CN was added. The solution was changed to homogeneous colorless solution immediately. The solution was then reacted for 16 hrs at room temperature and pressure. After 16 hrs, the solvent was removed under reduced pressure to give desired product.

[La₂L3^{ss}₃]: (12.0 mg, 3.99 x 10⁻³ mmol, 73.3% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): δ 10.32 (s, 3 x 2H), 9.00 (d, J = 6.9 Hz, 3 x 2H), 8.56 – 8.35 (m, 3 x 6H), 7.24 – 7.01 (m, 3 x 14H), 5.11 (p, J = 7.0 Hz, 3 x 2H), 1.71 (d, J = 7.1 Hz, 3 x 6H). ¹³C NMR (100 MHz, CD₃CN, 300 K, δ): 167.57, 166.99, 148.75, 148.51, 143.14, 141.70, 133.52, 128.51, 127.66, 126.60, 126.31, 125.93, 123.56, 52.25, 20.61. HRMS (ESI) calcd. for C₁₁₂H₉₆F₁₂La₂N₁₈O₂₄S₄ [La₂(L3^{ss})₃ + 4OTf⁻]²⁺: 1355.6841, found 1355.6792. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈La₂N₁₈O₃₀S₆·8H₂O: C 43.41, H 3.58, N 7.99, found: C 43.35, H 3.60, N 7.92.

[La₂L**3**^{RR}₃]: (13.5 mg, 4.48 x 10⁻³ mmol, 82.4% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.34 (s, 3 x 2H), 9.01 (d, J = 7.0 Hz, 3 x 2H), 8.61 – 8.39 (m, 3 x 6H), 7.25 – 6.97 (m, 3 x 14H), 5.11 (p, J = 7.0 Hz, 3 x 2H), 1.71 (d, J = 7.1 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300 K, δ): 167.52, 166.92, 148.69, 148.45, 143.05, 141.64, 133.46, 128.44, 127.59, 126.53, 126.25, 125.86, 123.48, 52.21, 20.55. HRMS (ESI) calcd. for C₁₁₁H₉₅F₉La₂N₁₈O₂₁S₃ [La₂(L**3**^{RR})₃ + 3OTf⁻ - H⁺]²⁺: 1280.7042, found 1280.7103. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈La₂N₁₈O₃₀S₆·19H₂O: C 40.80, H 4.03, N 7.52, found: C 40.90, H 3.57, N 7.30.

[Sm₂L3^{SS}₃]: (14.5 mg, 4.78 x 10⁻³ mmol, 87.9% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.40 (s, 3 x 2H), 9.11 (d, J = 6.8 Hz, 3 x 2H), 8.59 – 8.31 (m, 3 x 6H), 7.18 (dd, J = 5.2, 1.9 Hz, 3 x 6H), 7.07 (dd, J = 6.8, 2.8 Hz, 3 x 4H), 6.98 (s, 3 x 4H), 5.08 – 4.92 (m, 3 x 2H), 1.66 (d, J = 7.0 Hz, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 169.49, 168.71, 149.49, 149.22, 143.59, 141.77, 133.34, 128.61, 127.66, 125.87, 125.66, 125.32, 123.32, 52.29, 20.63. HRMS (ESI) calcd. for C₁₁₁H₉₆F₉N₁₈O₂₁S₃Sm₂ [Sm₂(L3^{SS})₃ + 3OTf⁻]³⁺: 861.8131, found 861.8192. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈Sm₂N₁₈O₃₀S₆·18H₂O: C 40.78, H 3.96, N 7.51, found: C 40.80, H 3.53, N 7.42.

[Sm₂L**3**^{RR}₃]: (12.1 mg, 3.99 x 10⁻³ mmol, 73.3% yield), ¹H NMR (495 MHz, CD₃CN, 299 K, δ): 10.15 (s, 3 x 2H), 8.89 (s, 3 x 2H), 8.45 – 8.32 (m, 3 x 6H), 7.13 (s, 3 x 6H), 7.03 (s, 3 x 4H), 6.92 (s, 3 x 4H), 5.01 – 4.95 (m, 3 x 2H), 1.72 – 1.50 (m, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 169.48, 168.71, 149.49, 149.23, 143.59, 141.76, 133.34, 128.61, 127.67, 125.87, 125.66, 125.31, 123.32, 52.28, 20.63. HRMS (ESI) calcd. for $C_{111}H_{96}F_9N_{18}O_{21}S_3Sm_2$ [Sm₂(L3^{RR})₃ + 3OTf⁻]³⁺: 861.8131, found 861.8160. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Sm_2N_{18}O_{30}S_6\cdot 18H_2O$: C 40.78, H 3.96, N 7.51, found: C 40.80, H 3.36, N 7.35.

[Eu₂L3^{SS}₃]: (14.1 mg, 4.64 x 10⁻³ mmol, 85.4% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 8.12 (s, 3 x 4H), 7.40 (s, 3 x 2H), 7.20 (t, *J* = 7.9 Hz, 3 x 2H), 7.05 (d, *J* = 3.9 Hz, 3 x 10H), 6.45 (d, *J* = 8.0 Hz, 3 x 2H), 6.36 (d, *J* = 7.9 Hz, 3 x 2H), 6.14 (s, 3 x 2H), 5.04 (s, 3 x 2H), 2.04 (d, *J* = 6.0 Hz, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 160.24, 155.37, 142.84, 142.47, 141.94, 134.83, 128.50, 127.50, 125.65, 125.29, 91.92, 91.80, 51.70, 21.73. HRMS (ESI) calcd. for C₁₁₂H₉₆Eu₂F₁₂N₁₈O₂₄S₄ [Eu₂(L3^{SS})₃ + 4OTf⁻]²⁺: 1369.1985, found 1369.1962. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈Eu₂N₁₈O₃₀S₆·18H₂O: C 40.74, H 3.96, N 7.50, found: C 40.83, H 3.56, N 7.44.

[Eu₂L**3**^{RR}₃]: (13.6 mg, 4.48 x 10⁻³ mmol, 82.3% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 8.07 (s, 3 x 4H), 7.13 (d, J = 15.9 Hz, 3 x 4H), 6.99 (s, 3 x 10H), 6.37 (s, 3 x 2H), 6.28 (d, J = 8.2 Hz, 3 x 2H), 6.09 (s, 3 x 2H), 4.80 (s, 3 x 2H), 1.99 (s, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 163.43, 160.28, 155.37, 142.83, 142.46, 142.02, 134.81, 128.50, 127.50, 125.66, 125.29, 91.94, 91.80, 51.70, 21.73. HRMS (ESI) calcd. for C₁₁₂H₉₆Eu₂F₁₂N₁₈O₂₄S₄ [Eu₂(L**3**^{RR})₃ + 4OTf⁻]²⁺: 1369.1985, found 1369.1949. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈Eu₂N₁₈O₃₀S₆·4H₂O: C 44.05, H 3.37, N 8.11, found: C 44.10, H 3.38, N 7.99.

 $[Gd_2L3^{SS}_3]$: (12.5 mg, 4.10 x 10⁻³ mmol, 75.4% yield), HRMS (ESI) calcd. for $C_{111}H_{96}F_9N_{18}O_{21}S_3Gd_2$ $[Gd_2(L3^{SS})_3 + 3OTf^-]^{3+}$: 866.4836, found 861.4828. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Gd_2N_{18}O_{30}S_6\cdot21H_2O$: C 39.98, H 4.06, N 7.36, found: C 40.01, H 3.54, N 7.30.

 $[Gd_2L3^{RR}_3]$: (14.3 mg, 4.69 x 10⁻³ mmol, 86.2% yield), HRMS (ESI) calcd. for $C_{111}H_{96}F_9N_{18}O_{21}S_3Gd_2$ $[Gd_2(L3^{RR})_3 + 3OTf^{-}]^{3+}$: 866.4836, found 861.4865. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Gd_2N_{18}O_{30}S_6\cdot 22H_2O$: C 39.77, H 4.10, N 7.32, found: C 39.88, H 3.30, N 7.20.

 $[Tb_2L3^{SS}_3]$: (14.7 g, 4.82 x 10⁻³ mmol, 88.6% yield), HRMS (ESI) calcd. for $C_{109}H_{94}F_3N_{18}O_{15}STb_2 [Tb_2(L3^{SS})_3 + OTf^- - 2H^+]^{3+}$: 767.5113, found 767.5154. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Tb_2N_{18}O_{30}S_6 \cdot 16H_2O$: C 41.01, H 3.86, N 7.55, found: C 41.07, H 3.49, N 7.48.

 $[Tb_2L3^{RR}_3]$: (13.6 g, 4.46 x 10⁻³ mmol, 81.9% yield), HRMS (ESI) calcd. for $C_{112}H_{96}Eu_2F_{12}N_{18}O_{24}S_4$ $[Tb_2(L3^{RR})_3 + 4OTf^-]^{2+}$: 1300.7232, found 1300.7169. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Tb_2N_{18}O_{30}S_6 \cdot 15H_2O$: C 41.24, H 3.82, N 7.59, found: C 41.36, H 3.33, N 7.48.

[Lu₂L3^{SS}₃]: (14.2 g, 4.61 x 10⁻³ mmol, 84.7% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.42 (s, 3 x 2H), 9.02 (d, J = 6.8 Hz, 3 x 2H), 8.56 – 8.33 (m, 3 x 6H), 7.17 (ddd, J = 8.9, 5.5, 3.3 Hz, 3 x 6H), 7.02 (d, J = 5.0 Hz, 3 x 8H), 4.97 (p, J = 6.9 Hz, 3 x 2H), 1.75 – 1.58 (m, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 166.82, 166.38, 147.04, 146.66, 143.08, 141.74, 133.26, 128.66, 127.61, 126.31, 125.79, 125.69, 123.07, 52.29, 20.77. HRMS (ESI) calcd. for C₁₁₁H₉₆F₉Lu₂N₁₈O₂₁S₃ [Lu₂(L3^{SS})₃ + 3OTf⁻]³⁺: 878.1615, found 878.1616. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈Lu₂N₁₈O₃₀S₆·7H₂O: C 42.68, H 3.46, N 7.86, found: C 42.66, H 3.39, N 7.77.

[Lu₂L3^{RR}₃]: (14.4 g, 4.67 x 10⁻³ mmol, 85.9% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.31 (s, 3 x 2H), 8.93 (s, 3 x 2H), 8.38 (s, 3 x 6H), 7.13 (s, 3 x 6H), 6.97 (s, 3 x 8H),

4.92 (d, J = 7.5 Hz, $3 \times 2H$), 1.61 (d, J = 7.1 Hz, $3 \times 6H$). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 166.41, 165.98, 146.63, 146.33, 142.69, 141.32, 128.26, 127.21, 125.87, 125.43, 125.29, 122.64, 51.86, 20.36. HRMS (ESI) calcd. for C₁₁₂H₉₆F₁₂Lu₂N₁₈O₂₄S₄ [Lu₂(L3^{RR})₃ + 4OTf⁻]²⁺: 1391.7185, found 1391.7209. Elemental analysis calcd. for C₁₁₄H₉₆F₁₈Lu₂N₁₈O₃₀S₆·7H₂O: C 42.68, H 3.46, N 7.86, found: C 42.74, H 3.54, N 7.84.

General synthetic procedures of [Ln₄L3₆]



Ln₂L3₃ (0.66 mmol) were dissolved in CH₃CN (0.6 ml) and ether was allowed to slowly diffuse into the solution. The solution was decanted, and the crystal was washed with ether and dried under vacuum to obtain the homometallic tetrahedron crystal.

[Eu₄L3^{ss}₆]: (1.80 mg, 2.96 x 10⁻⁴ mmol, 90.0% yield), ¹H NMR (495 MHz, CD₃CN, 299 K, δ): 9.04 (s, 6 x 4H), 7.81 (s, 6 x 2H), 7.31 (s, 6 x 2H), 6.79 (s, 6 x 4H), 6.40 (d, J = 31.8 Hz, 6 x 8H), 5.88 (s, 6 x 2H), 5.47 (s, 6 x 2H), 4.09 (s, 6 x 2H), 2.34 (s, 6 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 161.73, 156.50, 153.58, 142.91, 136.72, 135.57, 133.80, 127.79, 126.76, 125.52, 124.96, 121.93, 119.36, 92.00, 91.32, 52.29, 22.27. HRMS (ESI) calcd. for C₂₂₅H₁₉₂Eu₄F₂₇N₃₆O₅₁S₉ [Eu₄(L3^{ss})₆ + 9 OTf⁻]³⁺: 1875.2490, found 1875.2516. Elemental analysis calcd. for C₂₂₈H₁₉₂Eu₄F₃₆N₃₆O₆₀S₁₂·11H₂O·C₆H₁₄O: C 44.10, H 3.61, N 7.91, found: C 44.12, H 3.68, N 7.83.

[Eu₄L3^{RR}₆]: (1.68 mg, 2.77 x 10⁻⁴ mmol, 83.8% yield), ¹H NMR (495 MHz, CD₃CN, 299 K, δ) 9.03 (s, 6 x 4H), 7.80 (s, 6 x 2H), 7.33 (s, 6 x 2H), 6.79 (s, 6 x 4H), 6.40 (d, J = 30.9 Hz, 6 x 8H), 5.89 (s, 6 x 2H), 5.47 (s, 6 x 2H), 4.11 (s, 6 x 2H), 2.33 (s, 6 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 161.81, 156.51, 153.57, 142.91, 136.77, 135.56, 133.85, 128.50, 127.79, 126.76, 125.52, 124.96, 121.96, 119.39, 92.10, 91.38, 52.29, 22.26. HRMS (ESI) calcd. for C₂₂₅H₁₉₂Eu₄F₂₇N₃₆O₅₁S₉ [Eu₄(L3^{RR})₆ + 9 OTf⁻]³⁺: 1875.2490, found 1875.2516. Elemental analysis calcd. for C₂₂₈H₁₉₂Eu₄F₃₆N₃₆O₆₀S₁₂·14H₂O·2(C₆H₁₄O): C 44.15, H 3.83, N 7.72, found: C 44.13, H 3.78, N 7.72.

[Sm₄L3^{SS}₆]: (1.74 mg, 2.87 x 10⁻⁴ mmol, 86.9% yield), ¹H NMR (495 MHz, CD₃CN, 299 K, δ) 9.91 (s, 6 x 2H), 9.20 (s, 6 x 2H), 8.76 (d, J = 9.6 Hz, 6 x 2H), 8.63 (s, 6 x 2H), 8.54 (d, J = 8.6 Hz, 6 x 2H), 7.19 – 6.98 (m, 6 x 10H), 6.14 (s, 6 x 4H), 4.52 (d, J = 8.7 Hz, 6 x 2H), 1.43 (s, 6 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 170.25, 167.32, 150.56, 148.82, 143.61, 142.40, 132.68, 128.54, 127.64, 126.67, 125.68, 125.39, 122.20, 121.60, 119.64, 52.47, 20.73. HRMS (ESI) calcd. for C₂₂₂H₁₉₁F₁₈N₃₆O₄₂S₆Sm₄ [Sm₄(L3^{SS})₆ + 6 OTf⁻ H⁺]⁵⁺: 1034.1745, found Elemental 1034.1748. analysis calcd. for C₂₂₈H₁₉₂Sm₄F₃₆N₃₆O₆₀S₁₂·15H₂O·2(C₆H₁₄O): C 44.07, H 3.85, N 7.71, found: C 43.91, H 3.51, N 7.76.

[Sm₄L3^{RR}₆]: (1.89 g, 3.12 x 10⁻⁴ mmol, 94.4% yield), ¹H NMR (495 MHz, CD₃CN, 299 K, δ) ¹H NMR (495 MHz, Acetonitrile- d_3) δ 9.93 (s, 6 x 2H), 9.22 (s, 6 x 2H), 8.76 (d, J = 8.9 Hz, 6 x 2H), 8.74 – 8.59 (m, 6 x 2H), 8.54 (d, J = 9.3 Hz, 6 x 2H), 7.07 (dd, J = 29.5, 9.9 Hz, 6 x 10H), 6.13 (s, 6 x 4H), 4.52 (s, 6 x 2H), 1.44 (s, 6 x 6H). ¹³C NMR (125 MHz, CD₃CN,

300 K, δ): 170.26, 167.34, 150.55, 148.83, 143.61, 142.42, 132.68, 128.54, 127.64, 126.67, 125.86, 125.67, 125.41, 122.21, 121.62, 119.64, 52.47, 20.73. HRMS (ESI) calcd. for C₂₂₅H₁₉₂Sm₄F₂₇N₃₆O₅₁S₉ [Sm₄(**L3^{RR}**)₆ + 9 OTf⁻]³⁺: 1873.2460, found 1873.2405. Elemental analysis calcd. for C₂₂₈H₁₉₂Sm₄F₃₆N₃₆O₆₀S₁₂·12H₂O·C₆H₁₄O: C 44.02, H 3.63, N 7.90, found: C 44.03, H 3.72, N 7.92.

 $[Gd_4L3^{SS}_6]$: (1.45 mg, 2.38 x 10⁻⁴ mmol, 72.1% yield), HRMS (ESI) calcd for $C_{224}H_{192}F_{24}Gd_4N_{36}O_{48}S_8$ $[Gd_4(L3^{SS})_6 + 8 \text{ OTf}^-]^{4+}$ is 1374.4520, found 1374.4512. Elemental analysis calcd. for $C_{228}H_{192}Gd_4F_{36}N_{36}O_{60}S_{12}\cdot12H_2O$: C 43.40, H 3.45, N 7.99, found: C 43.31, H 3.38, N 7.78.

[Gd4L3^{RR}₆]: (1.23 mg, 2.02 x 10^{-4} mmol, 61.2% yield), HRMS (ESI) calcd for C₂₂₅H₁₉₂F₂₇Gd₄N₃₆O₅₁S₉ [Gd₄(L1^{RR})₆ + 9 OTf⁻]⁴⁺ is 1882.2535 (100%), found 1882.2499. Elemental analysis calcd. for C₂₂₈H₁₉₂Gd₄F₃₆N₃₆O₆₀S₁₂·13H₂O: C 43.28, H 3.47, N 7.97, found: C 43.22, H 3.58, N 7.85.

[Tb₄L3^{ss}₆]: (1.37 mg, 2.25 x 10^{-4} mmol, 68.1% yield), HRMS (ESI) calcd for C₂₂₂H₁₉₁F₁₈N₃₆O₄₂S₆Tb₄ [Tb₄(L3^{ss})₆ + 6 OTf⁻ - H⁺]⁵⁺ is 1040.9802, found 1040.9800. Elemental analysis calcd. for C₂₂₈H₁₉₂Tb₄F₃₆N₃₆O₆₀S₁₂·12H₂O·C₆H₁₄O: C 43.79, H 3.61, N 7.86, found: C 43.74, H 3.68, N 7.81.

[Tb₄L**3**^{RR}₆]: (1.66 mg, 2.72 x 10^{-4} mmol, 82.5% yield), HRMS (ESI) calcd for C₂₂₂H₁₉₁F₁₈N₃₆O₄₂S₆Tb₄ [Tb₄(L**3**^{SS})₆ + 6 OTf⁻ - H⁺]⁵⁺ is 1040.9802, found 1040.9839. Elemental analysis calcd. for C₂₂₈H₁₉₂Tb₄F₃₆N₃₆O₆₀S₁₂·17H₂O·C₆H₁₄O: C 43.18, H 3.72, N 7.75, found: C 43.13, H 3.46, N 7.67.

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[Lu₄L3^{ss}₆]: (1.89 mg, 3.07 x 10⁻⁴ mmol, 92.9% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ) 10.20 (s, 6 x 2H), 9.07 (s, 6 x 2H), 8.46 (dd, J = 10.6, 5.7 Hz, 6 x 6H), 7.05 (dt, J = 21.9, 6.3 Hz, 6 x 10H), 6.71 (d, J = 4.9 Hz, 6 x 4H), 5.02 (q, J = 6.5 Hz, 6 x 2H), 1.65 (t, J = 5.9Hz, 6 x 6H). HRMS (ESI) calcd. for C₂₂₅H₁₉₂Lu₄F₂₇N₃₆O₅₁S₉ [Lu₄(L3^{SS})₆ + 9 OTf⁻]³⁺: 1905.6090, found 1905.6060.

[Lu₄L**3**^{RR}₆]: (1.34 mg, 2.17 x 10⁻⁴ mmol, 65.9% yield), ¹H NMR (405 MHz, CD₃CN, 299 K, δ) 10.22 (s, 6 x 2H), 9.07 (s, 6 x 2H), 8.47 (d, J = 5.5 Hz, 6 x 6H), 7.21 – 6.91 (m, 6 x 10H), 6.72 (s, 6 x 4H), 5.02 (p, J = 7.0 Hz, 6 x 2H), 1.66 (d, J = 7.1 Hz, 6 x 6H). HRMS (ESI) calcd. for C₂₂₅H₁₉₂Lu₄F₂₇N₃₆O₅₁S₉ [Lu₄(L**3**^{RR})₆ + 9 OTf⁻]³⁺: 1905.6090, found 1905.6080.

General synthetic procedures of [Ln₂L4₃]



To a white suspension of L4 (30 mg, 0.039 mmol, 1.5 equiv.) in a mixture of 2.55 ml of DCM/MeOH (12:1, v/v), a solution of $Ln(OTf)_3$ (0.026 mmol, 1 equiv.) (Ln = La, Sm, Eu, Gd, Tb and Lu) in 2.35 ml of CH₃CN was added. The solution was changed to yellow immediately. The solution was then reacted for 16 hrs at room temperature and pressure. After 16 hrs, the solvent was removed under reduced pressure to give desired product.

[La₂L4^{ss}₃]: (39.4 mg, 0.114 mmol, 86.9% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.21 (s, 3 x 2H), 9.07 (d, *J* = 7.1 Hz, 3 x 2H), 8.67 – 8.40 (m, 3 x 6H), 7.88 (d, *J* = 8.3 Hz, 3 x 4H), 7.31 – 7.20 (m, 3 x 6H), 7.14 (s, 3 x 4H), 7.02 (d, J = 8.3 Hz, 3 x 4H), 6.95 (s, 3 x 4H), 5.14 – 5.05 (m, 3 x 2H), 1.76 (d, J = 7.1 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300 K, δ): 168.78, 168.17, 150.80, 150.04, 144.52, 142.98, 139.58, 130.07, 129.10, 128.65, 128.40, 127.82, 127.37, 123.02, 119.33, 118.78, 55.51, 22.11. HRMS (ESI) calcd. for C₁₄₇H₁₂₀F₉La₂N₁₈O₂₁S₃ [La₂(L4^{SS})₃ + 3 OTf⁻]³⁺: 1006.2012, found 1006.2037. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆La₂·7H₂O: C 50.14, H 3.76, N 7.02, found: C 50.24, H 4.21, N 6.71.

[La₂L4^{RR}₃]: (37.9 mg, 0.109 mmol, 83.7% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.21 (s, 3 x 2H), 9.07 (d, *J* = 6.6 Hz, 3 x 2H), 8.54 (d, *J* = 21.5 Hz, 3 x 6H), 7.88 (d, *J* = 8.3 Hz, 3 x 4H), 7.25 (s, 3 x 6H), 7.13 (s, 3 x 4H), 7.02 (d, *J* = 8.2 Hz, 3 x 4H), 6.96 (s, 3 x 4H), 5.08 (s, 3 x 2H), 1.76 (d, *J* = 7.1 Hz, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 167.40, 166.79, 149.42, 148.66, 143.13, 141.62, 138.50, 138.19, 135.54, 128.70, 127.72, 127.27, 127.02, 126.39, 125.98, 121.64, 52.14, 22.24. HRMS (ESI) calcd. for C₁₄₇H₁₂₀F₉La₂N₁₈O₂₁S₃ [La₂(L4^{RR})₃ + 3 OTf⁻]³⁺: 1006.2012, found 1006.2037. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆La₂·6H₂O: C 50.40, H 3.72, N 7.05, found: C 50.42, H 3.95, N 6.90.

[Sm₂L4^{SS}₃]: (38.1 mg, 0.0109 mmol, 83.5% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.15 (s, 3 x 2H), 9.03 (d, J = 7.0 Hz, 3 x 2H), 8.44 (dq, J = 7.3, 4.3, 3.5 Hz, 3 x 6H), 7.89 (d, J = 8.3 Hz, 3 x 4H), 7.35 – 7.16 (m, 3 x 6H), 7.10 (dd, J = 7.0, 2.5 Hz, 3 x 4H), 6.98 (d, J = 8.2 Hz, 3 x 4H), 6.91 (s, 3 x 4H), 5.19 (q, J = 7.0 Hz, 3 x 2H), 1.77 (d, J = 6.9 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300K, δ): 169.41, 168.43, 149.63, 148.99, 143.29, 141.71, 138.44, 137.96, 135.57, 128.69, 127.62, 127.20, 126.80, 125.81, 125.20, 121.36, 52.31, 20.90. HRMS (ESI) calcd. for C₁₄₇H₁₂₀F₉Sm₂N₁₈O₂₁S₃ [Sm₂(L4^{SS})₃ + 3 OTf⁻]³⁺: 1013.8759, found 1013.8749. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆Sm₂·8H₂O: C 49.58, H 3.77, N 6.94, found: C 49.59, H 4.07, N 6.81.

[Sm₂L4^{RR}₃]: (36.5 mg, 0.0104 mmol, 79.9% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.11 (s, 3 x 2H), 9.00 (d, J = 7.0 Hz, 3 x 2H), 8.43 (dq, J = 7.3, 4.3, 3.5 Hz, 3 x 6H), 7.89 (d, J = 8.3 Hz, 3 x 4H), 7.42 – 7.17 (m, 3 x 6H), 7.10 (dd, J = 7.0, 2.5 Hz, 3 x 4H), 6.98 (d, J = 8.2 Hz, 3 x 4H), 6.91 (s, 3 x 4H), 5.19 (q, J = 7.0 Hz, 3 x 2H), 1.78 (d, J = 6.9 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300K, δ): 169.42, 168.46, 149.74, 149.03, 143.40, 141.67, 138.49, 138.02, 135.53, 128.68, 127.62, 127.22, 126.87, 125.80, 125.25, 121.32, 52.25, 20.81. HRMS (ESI) calcd. for $C_{144}H_{118}N_{18}O_{12}Sm_2 [Sm_2(L4^{RR})_3 - 3H^+]^{3+}$: 863.9162, found 863.9186. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆Sm₂·17H₂O: C 47.46, H 4.09, N 6.64, found: C 47.34, H 3.96, N 6.48. [Eu₂L4^{SS}₃]: (36.4 mg, 0.0104 mmol, 79.8 % yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 8.21 – 7.96 (m, 3 x 4H), 7.42 (s, 3 x 2H), 7.24 (s, 3 x 4H), 7.15 (d, J = 5.8 Hz, 3 x 6H), 7.08 (s, 3 x 4H), 6.96 (s, 3 x 4H), 6.76 (s, 3 x 2H), 6.58 (s, 3 x 4H), 5.36 (s, 3 x 2H), 4.99 (s, 3 x 2H), 1.76 (s, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 155.25, 142.22, 138.62, 137.71, 135.36, 128.25, 127.19, 126.81, 126.69, 125.21, 122.15, 92.13, 91.52, 50.88, 21.13. HRMS (ESI) calcd. for C₁₄₇H₁₂₀Eu₂F₉N₁₈O₂₁S₃ [Eu₂(L4^{SS})₃ + 3OTf⁻]³⁺: 1015.2111, found 1015.2097. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆Eu₂·4H₂O: C 50.54,

H 3.62, N 7.07, found: C 50.55, H 3.90, N 6.50.

[Eu₂L4^{RR}₃]: (37.6 mg, 0.0108 mmol, 82.3% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 8.02 (d, *J* = 49.6 Hz, 3 x 4H), 7.47 (d, *J* = 38.6 Hz, 3 x 2H), 7.23 (s, 3 x 4H), 7.16 (d, *J* = 5.2 Hz, 3 x 6H), 7.07 (s, 3 x 4H), 6.97 (s, 3 x 4H), 6.79 (s, 3 x 2H), 6.59 (s, 3 x 4H), 5.36 (s, 3 x 2H), 5.03 (s, 3 x 2H), 1.73 (d, *J* = 22.2 Hz, 3 x 6H). ¹³C NMR (125 MHz, CD₃CN, 300 K, δ): 155.62, 142.66, 138.99, 138.10, 135.80, 128.64, 127.58, 127.20, 127.07, 125.61, 122.56, 92.56, 91.97, 51.29, 21.55. HRMS (ESI) calcd. for $C_{147}H_{120}Eu_2F_9N_{18}O_{21}S_3$ $[Eu_2(L4^{RR})_3 + 3OTf^-]^{3+}$: 1015.2111, found 1015.2097. Elemental analysis calcd. for $C_{150}H_{120}F_{18}N_{18}O_{30}S_6Eu_2 \cdot 5H_2O$: C 50.28, H 3.66, N 7.04, found: C 50.26, H 4.03, N 6.99. $[Gd_2L4^{SS}_3]$: (39.5 mg, 0.0113 mmol, 86.2% yield), HRMS (ESI) calcd. for $C_{144}H_{117}Gd_2N_{18}O_{12}[Gd_2(L4^{SS})_3 - 3H^+]^{3+}$: 868.5869, found 868.5845. Elemental analysis

calcd. for $C_{150}H_{120}F_{18}N_{18}O_{30}S_6Gd_2 \cdot 18H_2O$: C 47.07, H 4.11, N 6.59, found: C 46.97, H 3.72, N 6.40.

 $[Gd_2L4^{RR}_3]$: (37.6 mg, 0.0107 mmol, 82.0% yield), HRMS (ESI) calcd. for $C_{147}H_{120}F_9Gd_2N_{18}O_{21}S_3$ $[Gd_2(L4^{RR})_3 + 3OTf^-]^{3+}$: 1018.5452, found 1018.5465. Elemental analysis calcd. for $C_{150}H_{120}F_{18}N_{18}O_{30}S_6Gd_2\cdot 23H_2O$: C 45.99, H 4.27, N 6.44, found: C 45.88, H 3.76, N 6.21.

 $[Tb_2L4^{ss}_3]$: (37.2 mg, 0.0106 mmol, 81.1% yield), HRMS (ESI) calcd. for $C_{147}H_{120}F_9Tb_2N_{18}O_{21}S_3 [Tb_2(L4^{ss})_3 + 3OTf^-]^{3+}$: 1019.5472, found 1019.5485. Elemental analysis calcd. for $C_{150}H_{120}F_{18}N_{18}O_{30}S_6Tb_2 \cdot 16H_2O$: C 47.47, H 4.04, N 6.64, found: C 47.58, H 3.87, N 6.56.

 $[Tb_2L4^{RR}_3]$: (38.6 g, 0.0110 mmol, 84.1% yield), HRMS (ESI) calcd. for $C_{147}H_{120}F_9Tb_2N_{18}O_{21}S_3 [Tb_2(L4^{RR})_3 + 3OTf^-]^{3+}$: 1019.5472, found 1019.5485. Elemental analysis calcd. for $C_{150}H_{120}F_{18}N_{18}O_{30}S_6Tb_2 \cdot 18H_2O$: C 47.03, H 4.10, N 6.58, found: C 47.02, H 4.06, N 6.19.

[Lu₂L4^{ss}₃]: (38.6 mg, 0.0109 mmol, 83.4% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.13 (s, 3 x 2H), 8.99 (d, *J* = 7.0 Hz, 3 x 2H), 8.54 (dd, *J* = 6.0, 2.8 Hz, 3 x 2H), 8.46 (d, *J* = 6.0 Hz, 3 x 4H), 7.83 – 7.66 (m, 3 x 4H), 7.27 (dt, J = 5.6, 2.8 Hz, 3 x 6H), 7.12 – 7.00 (m, 3 x 4H), 6.89 (d, J = 8.5 Hz, 3 x 4H), 6.83 (s, 3 x 4H), 4.95 (t, J = 7.0 Hz, 3 x 2H), 1.69 (d, J = 7.0 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300K, δ): 166.79, 166.00, 147.36, 146.91, 142.84, 141.66, 138.40, 137.88, 135.37, 128.72, 127.58, 127.16, 126.75, 126.06, 125.62, 120.98, 52.16, 20.90. HRMS (ESI) calcd. for C₁₄₅H₁₁₈F₃Lu₂N₁₈O₁₅S [Lu₂(L4^{SS})₃ + OTf⁻ - 2H⁺]³⁺: 930.2509, found 930.2584. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆Lu₂·3H₂O: C 50.14, H 3.53, N 7.02, found: C 50.10, H 3.74, N 6.90.

[Lu₂L4^{RR}₃]: (39.3 mg, 0.0111 mmol, 85.0% yield), ¹H NMR (400 MHz, CD₃CN, 299 K, δ): 10.18 (s, 3 x 2H), 9.03 (d, *J* = 7.1 Hz, 3 x 2H), 8.54 (d, *J* = 6.2 Hz, 3 x 2H), 8.52 – 8.38 (m, 3 x 4H), 7.76 (d, *J* = 8.4 Hz, 2H), 7.36 – 7.21 (m, 3 x 4H), 7.11 – 6.99 (m, 3 x 4H), 6.89 (d, *J* = 8.5 Hz, 3 x 4H), 6.83 (s, 3 x 4H), 4.94 (t, *J* = 7.0 Hz, 3 x 2H), 1.69 (d, *J* = 7.0 Hz, 3 x 6H). ¹³C NMR (101 MHz, CD₃CN, 300 K, δ): 166.79, 165.98, 147.36, 146.92, 142.82, 141.67, 138.38, 137.83, 135.37, 128.72, 127.58, 127.14, 126.74, 126.07, 125.63, 120.99, 52.18, 20.92. HRMS (ESI) calcd. for C₁₄₇H₁₂₀F₉Lu₂N₁₈O₂₁S₃ [Lu₂(L4^{SS})₃ + 3OTf⁻]³⁺: 1030.2241, found 1030.2292. Elemental analysis calcd. for C₁₅₀H₁₂₀F₁₈N₁₈O₃₀S₆Lu₂·3H₂O: C 50.14, H 3.53, N 7.02, found: C 50.09, H 3.95, N 7.00.

2b.4.2 Kinetic studies

The activation energy of tetrahedron-to-helicate transformation was determined by Arrhenius Equation that shown below,

$$lnk = lnA - \frac{E_a}{RT}$$

where k is the rate constant, A is the frequency factor, E_a is the activation energy (Jmol⁻¹), R is the gas constant (8.314 JK⁻¹mol⁻¹), T is the temperature (K).

Enthalpy ΔH was determined by the following equation

$$E_a = \Delta H + RT$$

where E_a is the activation energy (Jmol⁻¹), R is the gas constant (8.314 JK⁻¹mol⁻¹), T is the temperature (K).

Activation energy was determined by dissolving Eu or Sm tetrahedron in CD₃CN. The stock solution was transferred to five NMR tubes after performing the first NMR data point. The temperature (308K, 318K, 328K, 338K) of NMR tubes were maintained by water bath. The concentration of the mixture of tetrahedron and helicate was calculated by the following equations depending on the resolution of NMR spectrum:

2 $[Ln_2L3_3] / 4[Ln_4L3_6]$ = integration of linker proton in helicate / integration of linker proton in tetrahedron OR 6 $[Ln_2L3_3] / 4[Ln_4L3_6]$ = integration of [NH+phenylproton] in helicate / integration of linker proton in tetrahedron WITH 3 $[Ln_2L3_3] + 6$ $[Ln_4L3_6] = [L3].$

	E _a (kJmol⁻ ¹)	А	∆H ₂₉₈ (kJmol ⁻¹)
Eu_A	90.1	2.93 x 10 ¹³	+87.6
Eu_B	98.8	2.63 x 10 ¹³	+96.3
Eu_C	102.3	1.16 x 10 ¹⁵	+99.8
Average	97.1 (5)	4.05 x 10 ¹⁴	+94.6 (5)

Table S2b-4. Kinetic data of Eu tetrahedron-to-helicate transformation.

	E _a (kJmol⁻ ¹)	А	ΔH ₂₉₈ (kJmol ⁻¹)
Sm_A	78.8	2.25 x 10 ¹¹	+76.3
Sm_B	89.5	2.06 x 10 ¹³	+87.0
Sm_C	84.5	4.51 x 10 ¹²	+82.0
Sm_D	88.1	2.17 x 10 ¹³	+85.6
Average	85.2 (4)	1.18 x 10 ¹³	+82.7 (4)

Table S2b-5. Kinetic data of Sm tetrahedron-to-helicate transformation.

<u>2b.4.3 Crystal structure of Eu₄(L3^{SS})₆:</u>

Information	Identifier from cif file
Empirical formula	C ₂₂₅ H ₁₉₂ Eu ₄ F ₂₇ N ₃₆ O ₅₁ S ₉
Formula weight	5625.51
Temperature/K	220.0
Crystal system	trigonal
Space group	R3
a/Å	29.9510(4)
b/Å	29.9510(4)
c/Å	71.8428(16)
α/°	90
β/°	90
γ/°	120
Volume/Å ³	55813.1(19)
Z	6
$\rho_{calc}g/cm^3$	1.004
µ/mm⁻¹	5.808
F(000)	17046.0
Crystal size/mm ³	0.24 × 0.17 × 0.14
Radiation	CuKα (λ = 1.54178)
20 range for data collection/°	5.902 to 100.82

Index ranges	-28 ≤ h ≤ 27, -27 ≤ k ≤ 29, -69 ≤ l ≤ 71
Reflections collected	41125
Independent reflections	20620 [R _{int} = 0.0639, R _{sigma} = 0.0648]
Data/restraints/parameters	20620/1946/2017
Goodness-of-fit on F ²	1.055
Final R indexes [I>=2σ (I)]	R ₁ = 0.0553, wR ₂ = 0.1497
Final R indexes [all data]	R ₁ = 0.0685, wR ₂ = 0.1594
Largest diff. peak/hole / e Å ⁻³	0.66/-0.18
Flack parameter	0.030(4)

	Shape measure (°)			
Eu ₄ (L3^{ss}) ₆	Tricapped trigonal prism	Capped square antiprism		
	(D _{3h})	(C _{4v})		
Eu1	6.45	11.13		
Eu2	5.82	10.33		
Eu3	5.94	10.23		
Eu4	6.11	10.93		

Supplementary Table 7. Results of the Shape Analysis for Eu₄(L3^{SS})₆ tetrahedron.

Shape analysis was performed by the software developed by Raymond's group.²⁵⁻²⁶ The coordination geometry of the $Eu_4(L3^{SS})_6$ tetrahedron can be best described as tricapped trigonal prism.

2b.4.4 Photophysical data

	λ_{abs}^{max}	${m arepsilon}^{\sf max}$
	(nm)	(L·mol ⁻¹ ·cm ⁻¹)
[La ₂ (L4^{SS}) ₃](CF ₃ SO ₃) ₆	345	125117.0
$[La_2(\mathbf{L4^{RR}})_3](CF_3SO_3)_6$	345	128419.6
[Sm ₂ (L4^{ss}) ₃](CF ₃ SO ₃) ₆	345	104030.4
[Sm ₂ (L4^{RR}) ₃](CF ₃ SO ₃) ₆	345	104038.3
[Eu ₂ (L4^{ss}) ₃](CF ₃ SO ₃) ₆	345	124128.9
[Eu ₂ (L4^{RR}) ₃](CF ₃ SO ₃) ₆	345	125331.4
[Gd ₂ (L4^{ss}) ₃](CF ₃ SO ₃) ₆	345	108324.9
$[Gd_2(\mathbf{L4^{RR}})_3](CF_3SO_3)_6$	345	115018.7
[Tb ₂ (L4^{ss}) ₃](CF ₃ SO ₃) ₆	345	114213.3
[Tb ₂ (L4 ^{RR}) ₃](CF ₃ SO ₃) ₆	345	119793.5
[Lu ₂ (L4^{ss}) ₃](CF ₃ SO ₃) ₆	345	117521.9
[Lu ₂ (L4^{RR}) ₃](CF ₃ SO ₃) ₆	345	115357.7

Table S2b-1. A summary of selected photophysical properties, UV-Vis absorption and luminescence data of $[Ln_2L4_3]$ in acetonitrile solution using a 10 mm cuvette and filter 380 nm.

	$\lambda_{abs}{}^{max}$	$\boldsymbol{\varepsilon}^{max}$	$\lambda_{ ext{em}}{}^{ ext{max}}$	ф _х ^ь (%)	τ
	(nm)	(L·mol ⁻¹ ·cm ⁻¹)	(nm)		(ms)
[La ₂ (L3 ^{ss}) ₃](CF ₃ SO ₃) ₆	311	59052.4	/	/	/
[La ₂ (L3^{RR}) ₃](CF ₃ SO ₃) ₆	311	58787.3	/	/	/
[Sm ₂ (L3^{ss}) ₃](CF ₃ SO ₃) ₆	311	60056.4	/	/	/
[Sm ₂ (L3^{RR}) ₃](CF ₃ SO ₃) ₆	311	62036.6	/	/	/
[Eu ₂ (L3^{ss}) ₃](CF ₃ SO ₃) ₆	311	62925.5	616	4.07 (0.015)	1.42
[Eu ₂ (L3^{RR}) ₃](CF ₃ SO ₃) ₆	311	61958.6	616	4.41 (0.005)	1.41
$[Gd_2(L3^{SS})_3](CF_3SO_3)_6$	311	58396.8	462	/	0.0073
$[Gd_2(\mathbf{L3^{RR}})_3](CF_3SO_3)_6$	311	59227.0	/	/	/
[Tb ₂ (L3^{SS}) ₃](CF ₃ SO ₃) ₆	311	61469.6	/	/	/
[Tb ₂ (L3^{RR}) ₃](CF ₃ SO ₃) ₆	311	60113.3	/	/	/
[Lu ₂ (L3^{SS}) ₃](CF ₃ SO ₃) ₆	311	60668.1	/	/	/
[Lu ₂ (L3^{RR}) ₃](CF ₃ SO ₃) ₆	311	59773.5	/	/	/
[Sm ₄ (L3^{ss}) ₆](CF ₃ SO ₃) ₁₂	330	130019.8	/	/	/
[Sm ₄ (L3^{RR}) ₆](CF ₃ SO ₃) ₁₂	330	139354.6	/	/	/
[Eu ₄ (L3^{SS}) ₆](CF ₃ SO ₃) ₁₂	330	123947.0	616	0.81 (0.035)	1.28
[Eu4(L3^{RR}) 6](CF3SO3)12	330	121335.7	616	0.83 (0.035)	1.27
[Gd ₄ (L3^{SS}) ₆](CF ₃ SO ₃) ₁₂	330	123657.1	/	/	/
$[Gd_4(L3^{RR})_6](CF_3SO_3)_{12}$	330	121829.6	466	/	0.0087 ^c
[Tb ₄ (L3^{ss}) ₆](CF ₃ SO ₃) ₁₂	330	131075.9	/	/	/
$[Tb_4(\mathbf{L3^{RR}})_6](CF_3SO_3)_{12}$	330	127713.5	/	/	/
[Lu ₄ (L3^{SS}) ₆](CF ₃ SO ₃) ₁₂	330	110557.9	/	/	/
[Lu4(L3^{RR}) 6](CF3SO3)12	330	112691.3	/	/	/

Table S2b-2. A summary of selected photophysical properties, UV-Vis absorption and luminescence data of $[Ln_2L3_3]$ and $[Ln_4L3_6]$ complexes in acetonitrile solution^a. ^ausing a 10 mm cuvette and filter 380 nm. ^bThe relative quantum yields were referenced with quinine sulfate in 0.1 M sulfuric acid (ϕ = 0.577, λ_{ex} = 350nm) with 10 mm cuvette. ^cMeasurement performed at 77K in 1:4 MeOH/EtOH.

	glum				
	⁵ D ₀ -> ⁷ F ₁	⁵ D ₀ -> ⁷ F ₂	⁵ D ₀ -> ⁷ F ₄	⁵ D ₀ -> ⁷ F ₄	
	ΔJ = 1	ΔJ = 2	ΔJ = 4	∆J = 4	
Complex	(592.5 nm)	(615 nm)	(695.5 nm)	(704 nm)	
[Eu ₂ (L3^{ss}) ₃]	+0.10	-0.02	_	_	
[Eu ₂ (L3^{RR}) ₃]	- 0.10	+0.02	_	_	
[Eu ₄ (L3^{SS}) ₆]	+0.10	-0.02	-0.05	+0.23	
[Eu ₄ (L3^{RR}) ₆]	- 0.08	+0.02	+0.04	-0.23	

*All g_{lum} values have associated nominal assumed instrumental uncertainty of ± 0.01 .

Table S2b-3. g_{lum} values for $[Eu_2(L3^{SS/RR})_3]$ and $[Eu_4(L3^{SS/RR})_6]$ calculated from total intensity and CPL spectra.

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Chapter 3: Formation of chiroptical lanthanide heterometallic Ln₄L₆ tetrahedral cage

3.1 Introduction

3.1.1 Introduction to lanthanide heterometallic systems

Transition metal based supramolecular complexes have been proven to be an excellent candidate for various applications including catalyst¹⁻⁵ and luminescent probes.⁶⁻⁸ In order to fine tune the functional role of supramolecular materials, numerous hybrid metal supramolecular systems have been reported. For instance, some supramolecular catalysts involved the use of two different metal ions.⁹⁻¹¹ One can act as structural nodes to provide a well-defined structure for matching substrate, while another one act as a catalytic centre to interact with target substrates.

For lanthanide supramolecular complexes, numerous d-f hybrid complexes topologies¹²⁻¹⁴ as well as heterometallic lanthanide based supramolecular complexes have been prepared.¹⁵⁻²⁰ It was found that the combination of different lanthanide ions can lead to an enhancement in photophysical properties including improvement in up-conversion efficiency²¹ as well as better optimization in the optical properties.²² The enhancement in photophysical properties can help to develop potential dual imaging agent that cover both visible and near-infrared region.²³

3.1.2 Preparation of lanthanide heterometallic complexes

As mentioned in the previous chapters, preparation of lanthanide supramolecular complexes is difficult compared with transition metal due to their labile coordination number and poor stereochemical preferences. Preparation of lanthanide heterometallic systems is more challenging due to the similar ionic radii and chemical properties of lanthanide ions.²⁴ During the one-pot mixed lanthanide self-assembly process, lanthanide ions will bind to the ligand randomly and complexes with different lanthanide content will result.

Generally, there are two strategies for preparing lanthanide heterometallic complexes. The first strategy is to synthesize an organic ligand that consists of two different chelating groups.²⁵⁻²⁷ These two coordination sites are well designed so that they can bind to the lanthanide ions selectively. However, selective coordination with different lanthanide ions is difficult using the same ligand. Undesired supramolecular products with different fractions of lanthanide content can be only partially removed from the reaction mixture and thus the complexes can be only studied as a part of a statistical mixture.²⁶ Rare successful examples of pure heterometallic complexes were demonstrated by Aromí's group.²⁶ They have synthesized a series of asymmetric ligands with excellent selectivity towards lanthanide ions by employing β-diketone and dipicolinate-like as the coordination sites. Figure 3-1 showed the ligand examples.



Figure 3-1. Ligand examples reported by Aromí's group.²⁶

Lanthanide heterometallic complexes can be also prepared by stepwise reaction. Different from one-pot self-assembly process, stepwise reaction approach required the preparation of lanthanide complexes followed by the attachment of two lanthanide complexes.^{28, 29} Stepwise reaction approach is a better approach since the lanthanide contents of the resulting complexes can be controlled easily. However, this strategy strongly depends on the use of ligand having strong chelating power such as macrocyclic ligands.³⁰⁻³² Otherwise, the complexes are not stable enough to survive the next steps in the reaction. Figure 3-2 showed an example of lanthanide heterometallic complexes that prepared by stepwise reaction approach.³²



Figure 3-2. Lanthanide heterometallic complexes prepared by stepwise reaction approach.³²

3.1.3 Scope of study



Figure 3-3. Self-assembly of a mixed heterometallic tetrahedron from metal helicates by one-step process by ether diffusion crystallization. The heterometallic tetrahedron consists of Eu and Gd in a ratio of 1:1.

Inspired by the potential application of heterometallic lanthanide systems, we investigate the possibility of developing heterometallic lanthanide tetrahedron via crystallization. In this chapter, a series of mixed lanthanide heterometallic tetrahedra were developed with the use of *C*₂-symmetical bis(tridentate) ligand. Crystallization of two lanthanide helicates by slow diffusion of diisopropyl ether leads to a formation of mixed heterometallic tetrahedron. Detailed ESI-HRMS deconvolution was done to analyze the percentage amount of tetrahedron. The effect of ionic radii towards the formation of heterometallic tetrahedron was also investigated. By varying the ratio of two different lanthanide helicates, the percentage amount of tetrahedra varies. Photophysical studies of the tetrahedra were further investigated.

3.2 Result and discussion

3.2.1 Preparation of $[Eu_nGd_{4-n}L3_6](n = 0-4)$

A C_2 -symmetrical bis(tridentate) ligand L3 was reported in Chapter 2b and the corresponding lanthanide helicate was found to have an helicate-to-tetrahedron transformation. Such helicate-to-tetrahedron transformation depends on the size of lanthanide cations and the linker length and pure tetrahedron can be only isolated via crystallization. With this interesting findings, further investigation of the helicate-to-tetrahedron transformation was done by crystallization of different mixture of [Ln₂L3₃] helicate. It was hypothesized that the helicate will either undergo a self-sorting behavior to form pure homometallic tetrahedron or forming a mixture of tetrahedron with different metal content due to the similar ionic radii of lanthanide ions.

Interestingly, upon crystallization of a mixture of $[Eu_2(L3)_3]$ and $[Gd_2(L3)_3]$ helicates in a ratio of 1 to 1, a single crystal of mixed heterometallic $[Eu_nGd_{4-n}L3_6]$ (n = 0-4) was obtained. X-ray crystal structure of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4) revealed a tetrahedral topology constructed by four lanthanide ions and six ligands (figure 3-4).


Figure 3-4. X-ray crystal structure of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4). The Eu and Gd metal centers were refined as 50:50 by X-ray crystallography however depicted as a single element in the illustration.

Precise differentiation by X-ray crystallography between Eu and Gd metal centre is no possible due to their similar electron density. Thus, each Eu and Gd metal centre is refined as 50:50. In the crystal structure, all the lanthanide centres are arranged in the same Δ-configurations and each metal centre can be best described as as tricapped trigonal prism. Average Ln – N (Ln = Eu or Gd) distance is 2.553 Å, comparable to an average Eu – N (2.561 Å) and Gd – N (2.581 Å) distance of pyridinedicarboxylic chelating unit. Average Ln – O distance was found to be 2.406 Å, which is also comparable to the average Gd – O and distance Eu – O (2.426 Å and 2.423 Å) that reported in literature.³³ Characterization by NMR is not possible due to the strong paramagnetic nature of Gd. Therefore, the chemical formulas were examined by ESI-HRMS. In the ESI-HRMS spectrum, peaks series were observed corresponding to the heterometallic [Eu_nGd_{4-n}L3₆] (n = 0-4) species with the progressive loss of anions and protons (figure 3-5).



Figure 3-5. ESI-HRMS spectra of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4) with insets showing the observed and isotropic pattern of the peaks corresponding to $[Eu_2Gd_2L3_6 + 70T_f^{2}]^{5+}$.

3.2.3 Effect of difference in ionic radii

Gd was replaced by other lanthanide ions to investigate the effect of ionic radii. When Gd was replaced by Sm, which exhibited less paramagnetic nature and a slightly small ionic radii difference with Eu (Δr_{Eu-Gd} 1.3 pm vs Δr_{Eu-Sm} 1.2 pm),³⁴ similar mixed heterometallic tetrahedron was formed upon crystallization of a mixture of [Eu₂(L3)₃] and [Sm₂(L3)₃] in a ratio of 1 to 1. The chemical formulas were also confirmed by ESI-HRMS, in which single set of peaks corresponding to [Eu_nSm_{4-n}L3₆] (n = 0-4) species were observed (figure 3-6). Similar with the result in chapter 2b, ¹H NMR revealed that [Eu₂Sm₂L3₆] was not stable upon dissolution and it slowly transformed back to a mixture of [Eu₂(L3)₃], [EuSm(L3)₃] and [Sm₂(L3)₃] after 13 days (figure 3-7).



Figure 3-6. ESI-HRMS spectra of $[Eu_nSm_{4-n}L3_6]$ (n = 0-4) with insets showing the observed and isotropic



pattern of the peaks corresponding to $[Eu_2Sm_2L3_6 + 7OTf]^{5+}$.

Figure 3-7. Variation of ¹H NMR of $[Eu_nSm_{4-n}L3_6]$ (n = 0-4) upon dissolution in CD₃CN.

Different result was obtained when La, Tb and Lu were used to replace Gd. In the case of Tb, five different types of tetrahedra were observed in the ESI-HRMS upon crystallization of a mixture of [Eu₂L3₃] and [Tb₂L3₃] in a ratio of 1 to 1. These five tetrahedra can be assigned as [Eu₄L3₆], [Eu₃TbL3₆] [Eu₂Tb₂L3₆], [EuTb₃L3₆] and [Tb₄L3₆] (figure 3-8). Similar result was also observed when Gd was replaced by Lu. The ESI-HRMS also showed the existence of a mixture of [Eu₄L3₆], [Eu₃LuL3₆] [Eu₂Lu₂L3₆], [EuLu₃L3₆] and [Lu₄L3₆] (figure 3-9).



Figure 3-8. Expanded region of the ESI-HRMS spectrum of a mixture of [Eu₄L**3**₆], [Eu₃TbL**3**₆], [Eu₂Tb₂L**3**₆], [EuTb₃L**3**₆] and [Tb₄L**3**₆].



Figure 3-9. Expanded region of the ESI-HRMS spectrum of a mixture of [Eu₄L3₆], [Eu₃LuL3₆],

[Eu₂Lu₂L**3**₆], [EuLu₃L**3**₆] and [Lu₄L**3**₆]. 133 When Gd was replaced by La, ESI-HRMS result revealed that the major species were $[La_2L3_3]$, $[EuLaL3_3]$ and $[Eu_2L3_3]$, and minor species were found to be $[Eu_2La_2L3_6]$ and $[Eu_3LaL3_6]$ (figure 3-10). As discussed in chapter 2b, La highly prefers to form helicates upon coordinating with lanthanide. Thus, it is expected that La also biases the helicate-to-tetrahedron transformation. $[La_2L3_3]$ was also observed in the crystallized product.



Figure 3-10. Expanded region of the ESI-HRMS spectrum of a mixture of [Eu2L33], [EuLaL33], [La2L33], [Eu2La2L36] and [Eu3LaL36].

Mass deconvolution using OrginPro was performed and figure 3-11 summarizes the effect of ionic radii towards the helicate-to-tetrahedron transformation via crystallization. Generally, the formation of lanthanide heterometallic tetrahedron is highly sensitive to the difference of ionic radii between the lanthanide ions used in crystallization. In the case of Eu and Gd, the ionic radii differ from 1.3 pm, and the largest percentage amount of complex was found to be $[Eu_2Gd_2L3_6]$ heterometallic tetrahedron. Similar result can be observed when the difference of ionic radii slightly decreased to 2.5 pm, which is the case of Eu and Tb.

Interestingly, the intensity of $[Ln_2Ln'_2L3_6]$ heterometallic tetrahedron also decreased if the difference of ionic radii increased. As evidenced in the combination of Eu and Lu, the major species was found to be $[Eu_4L3_6]$ and the heterometallic $[Eu_2Lu_2L3_6]$ became minor species when the difference of ionic radii became 8.8 pm.

The maximum difference of ionic radii that this system can tolerate to form heterometallic tetrahedron, i.e., the cutoff ionic radii, was further investigated by utilizing different pairs of lanthanides. Due to the similar mass difference, selection of lanthanide pairs was challenging. Based on the MS deconvolution result, the cutoff ionic radii was found to be 8.8 pm in the combination of Eu and Lu.



Figure 3-11. Quantitative analysis of the lanthanide heterometallic tetrahedra. The ESI-MS data was calculated based on the intensity of $[complex + 9 OTf]^{3+}$

3.2.3 Variation of metal content of the heterometallic tetrahedra

The preferential formation of heterometallic tetrahedra via crystallization is important for preparing of lanthanide heterometallic material. Further investigation has been done to study the formation process. However, the entire process of helicate-to-tetrahedron transformation via crystallization cannot be monitored by NMR and ESI-HRMS. Therefore, the solution state of the mixed complexes first studied before crystallization. Interestingly, formation of intermediate was observed in ¹HNMR. Upon mixing [Eu₂L**3**₃] and [Sm₂L**3**₃] in MeCN, ¹HNMR revealed a formation of new species correspond to the heterometallic helicate [EuSmL**3**₃] (figure 3-12). The signal of [EuSmL**3**₃] increased gradually. After 5 hrs, the system reached to an equilibrium and the ratio of [Eu₂L**3**₃]: [Sm₂L**3**₃]: [EuSmL**3**₃] was found to be 1:1:1.



Figure 3-12. Variation of ¹H NMR when [Eu₂L**3**₃] was mixed with [Sm₂L**3**₃] after (A) 0 hr and (B) 5 hrs in CD₃CN.

The ionic radii also governed the transformation process from homometallicto heterometallic helicate. Upon mixing [Eu₂L3₃] and [Lu₂L3₃] in MeCN, similar homometallic- to heterometallic helicate transformation was observed. However, the rate was slower compared with that of the case in Eu and Sm. No new NMR signal can be observed after mixing $[Eu_2L3_3]$ and $[Lu_2L3_3]$ for 5 hrs and insignificant amount of $[EuLuL3_3]$ was formed after 7 days (figure 3-13).



Figure 3-13. Variation of ¹H NMR when [Eu₂L**3**₃] was mixed with [Lu₂L**3**₃] after (A) 5 hrs and (B) 7 days in CD₃CN.

3.2.4 Proposed mechanism of helicate-to-tetrahedron transformation process

During the crystallization process, no precipitate was observed. As ligand was not soluble in MeCN, it was hypothesized that the helicate did not undergo complete dissociation during helicate-to-tetrahedron transformation. Based on the above homometallic helicate-to-heterometallic helicate transformation study, it was further hypothesized the rate of association and dissociation of lanthanide ions was governed by the ionic radii. The statistical and experimental result were shown in figure 3-14.

Statistical	result (1:1)	1.1	experimental result (1:1)					
Species	Percentage (%)		Species	Percentage (%)	Species	Percentage (%)	Species	Percentage (%
Eu ₄ L3 ₆	11.1		Eu ₄ L3 ₆	2.85	Eu ₄ L3 ₆	2.74	Eu ₄ L3 ₆	59.07
Eu ₃ SmL3 ₆	22.2		Eu ₃ GdL3 ₆	19.68	Eu ₃ TbL3 ₆	17.70	Eu ₃ LuL3 ₆	25.88
Eu ₂ Sm ₂ L3 ₆	33.3		Eu2Gd2L36	45.77	Eu ₂ Tb ₂ L3 ₆	45.33	Eu ₂ Lu ₂ L3 ₆	8.55
EuSm ₃ L3 ₆	22.2		EuGd ₃ L3 ₆	26.64	EuTb ₃ L3 ₆	29.84	EuLu ₃ L3 ₆	2.55
Sm ₄ L3 ₆	11.1		Gd ₄ L3 ₆	5.07	Tb ₄ L3 ₆	4.39	Lu ₄ L3 ₆	3.96

Figure 3-14. Comparison of statistical and experimental result of MS deconvolution of mixed lanthanide heterometallic tetrahedra.

It was hypothesized that lanthanide ions having similar ionic radii will have similar rate of association and dissociation, which lead to the formation of heterometallic tetrahedron. Therefore, the crystallization process is governed by both ionic radii as well as the ratio of lanthanide ions that present in the helicate solution. It was then suggested that the metal ratio of the resulting tetrahedron can be simply altered by varying the ratio of the two lanthanide helicates in the mixed solution before crystallization. Upon crystallization of a mixture containing [Eu₂(L3)₃] and [Gd₂(L3)₃] helicates in a ratio of 3:1, the MS deconvolution result showed the formation of highest amount of heterometallic [Eu₃GdL3₆] (figure 3-15). The metal content of the tetrahedron was further varied by changing the ratio of the helicate and similar result was observed in the MS deconvolution spectra (figure 3-16).



Figure 3-15. MS deconvolution result of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4) prepared by crystallization of a mixture

of [Eu₂**L3**₃] and [Gd₂**L3**₃] in a ratio of 1:3.



Figure 3-15. MS deconvolution result of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4) prepared by crystallization of a mixture

of $[Eu_2L3_3]$ and $[Gd_2L3_3]$ in a ratio of 3:1.

3.2.4 Photophysical properties of heterometallic tetrahedra

The photophysical properties of emissive heterometallic tetrahedra [Eu_nGd_{4-n}L3₆] (n = 0-4) prepared by different ratio of helicate were examined. Upon excitation at 330 nm, successful sensitization of Eu^{3+ 5}D₀ excited state through antenna effect was observed. The luminescence spectrum revealed four narrow Eu emission lines at 595, 616, 688 and 697 nm, which corresponds to the transition ⁵D₀ to ⁷F_J (*J* = 1, 2 and 4) transitions (figure 3-16a). Quantum yield measurement was also done and [Eu₄L3^{RR}₆] exhibited the highest quantum yield of 0.83%. Due to the decrease in luminescent Eu³⁺ content of the tetrahedron, the quantum yield decreased gradually from 0.70% [Eu_nGd_{4-n}L3^{RR}₆] (prepared by ratio Eu/Gd: 3/1) to 0.50% [Eu_nGd_{4-n}L3^{RR}₆] (prepared by ratio Eu/Gd: 1/3).

Solution circular dichroism and circularly polarized luminescence were also done to investigate the chiroptical properties of lanthanide heterometallic tetrahedra [Eu_nGd_{4-n}L3₆] (n = 0-4). Generally, five tetrahedra showed similar CD spectrum. Mirror image and strong cotton effects were observed for *R*- and *S*-isomers at 212, 240, 259, 273, 312 and 362 nm. Four Eu³⁺-containing heterometallic tetrahedra also exhibit CPL. Based on the luminescence result, tetrahedron with the highest content of emissive Eu³⁺ ions were expected to have the strongest CPL intensity. Indeed, [Eu₄L3₆] was found to exhibit the highest CPL intensity. The normalized CPL intensity decreased gradually from [Eu_nGd_{4-n}L3^{RR}₆] (prepared by ratio Eu/Gd: 3/1) to 0.50% [Eu_nGd_{4-n}L3^{RR}₆] (prepared by ratio Eu/Gd: 1/1) to 0.34% [Eu_nGd_{4-n}L3^{RR}₆] (prepared by ratio Eu/Gd: 1/3) as expected. Four Eu³⁺-containing tetrahedra were found to have the same *g*_{lum} value (587 nm) = 0.15 and mirror image of CPL spectra confirmed the successful stereoselective control via point chirality (figure 3-16b).



Figure 3-16. Photophysical properties of Eu³⁺-containing tetrahedra. (a) Normalized emission spectrum and (b) Normalized CPL spectrum of [Eu_nGd_{4-n}L**3**₆].

3.3 Conclusion

To conclude, with the use of C_2 -symmetrical bis(tridentate) ligand L3, a mixture of heterometallic tetrahedra [Eu_nGd_{4-n}L3₆] (n = 0-4) can be prepared via crystallization. The chemical formulae of the tetrahedra were confirmed by ESI-HRMS and X-ray crystallography. The formation of heterometallic tetrahedra was highly sensitive to the ionic radii of lanthanide ions. By varying the ratio of helicate solution before crystallization, the metal content of the tetrahedron can be fine-tuned. The Eu³⁺⁻ containing tetrahedra were luminescent and exhibited CPL signal with g_{lum} (587 nm) value = 0.15.

3.4 Experimental

3.4.1 Synthesis

General synthetic procedures of [Ln₂L3₃]



To a white suspension of L3 (10 mg, 0.016 mmol, 1.5 equiv.) in a mixture of 8.49 ml of DCM/MeOH (12:1, v/v), a solution of $Ln(OTf)_3$ (0.011 mmol, 1 equiv.) (Ln = Dy and Ho) in 7.83 ml of MeCN was added. The solution was changed to homogeneous colorless solution immediately. The solution was then reacted for 16 hrs at room temperature and pressure. After 16 hrs, the solvent was removed under reduced pressure to give desired product.

 $[Dy_2L3^{SS}_3]$: (14.2 mg, 4.64 x 10⁻³ mmol, 85.4% yield), HRMS (ESI) calcd. for $C_{112}H_{96}Dy_2F_{12}N_{18}O_{24}S_4$ $[Dy_2(L3^{SS})_3 + 4OTf^{-}]^{2+}$: 1379.7060, found 1379.7075. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Dy_2N_{18}O_{30}S_6\cdot11H_2O$: C 42.06, H 3.65, N 7.74, found: C 42.10, H 3.14, N 7.78.

 $[Ho_2L3^{SS}_3]$: (13.9 mg, 4.54 x 10⁻³ mmol, 83.4% yield), HRMS (ESI) calcd. for $C_{112}H_{96}Eu_2F_{12}N_{18}O_{24}S_4$ $[Ho_2(L3^{SS})_3 + 4OTf^-]^{2+}$: 1381.7072, found 1381.7081. Elemental analysis calcd. for $C_{114}H_{96}F_{18}Ho_2N_{18}O_{30}S_6\cdot 20H_2O$: C 40.01, H 4.01, N 7.37, found: C 40.07, H 3.29, N 7.24.

General procedures for preparing homometallic tetrahedra



 Ln_2L3_3 (0.66 mmol) was dissolved in MeCN (0.6 ml) and ether was allowed to slowly diffuse into the solution containing Ln_2L_3 . The solution was decanted, and the crystal was washed with ether and dried under vacuum to obtain the homometallic tetrahedra crystal.

Dy4L3^{SS}₆: (1.66 mg, 2.71 x 10⁻⁴ mmol, 82.4% yield), HRMS (ESI): m/z calcd. for C₂₂₃H₁₉₂Dy4F₂₁N₃₆O₄₅S₇ [Dy4L3^{SS}₆ + 7 OTf⁻]⁵⁺: 1073.9743, found 1073.9764; Calcd for C₂₂₂H₁₉₁Dy4F₁₈N₃₆O₄₂S₆ [Dy4L3^{SS}₆ + 6 OTf⁻ H⁺]⁵⁺: 1043.7821, found 1043.7826; Calcd. for C₂₂₁H₁₉₀Dy4F₁₅N₃₆O₃₉S₅ [Dy4L3^{SS}₆ + 5 OTf⁻ 2H⁺]⁵⁺: 1013.7902, found 1013.7969; Calcd. for C₂₂₀H₁₈₉Dy4F₁₂N₃₆O₃₆S₄ [Dy4L3^{SS}₆ + 4 OTf⁻ 3H⁺]⁵⁺: 983.7983, found 983.7963; Calcd. for C₂₂₀H₁₉₀Dy4F₁₂N₃₆O₃₆S₄ [Dy4L3^{SS}₆ + 4 OTf⁻ 2H⁺]⁶⁺: 819.9998, found 820.0034. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆ Dy₃TbN₃₆O₆₀S₁₂·11H₂O·C₆H₁₄O: C 43.81, H 3.58, N 7.86, found: C 43.88, H 3.63, N 7.46.

General procedures for preparing heterometallic tetrahedra

 Ln_2L3_3 and Ln'_2L3_3 (depends on different mole ratio) were dissolved in MeCN (0.6 ml) and ether was allowed to slowly diffuse into the solution containing a mixture of Ln_2L3_3 and Ln'_2L3_3 . The solution was decanted, and the crystal was washed with ether and dried under vacuum to obtain the homometallic tetrahedra crystal.

 $[Eu_{n}Gd_{4-n}L3^{SS}_{6}] \text{ (prepared by ratio Eu/Gd: 3/1): (1.49 mg, 2.45 x 10⁻⁴ mmol, 74.3% yield),} HRMS (ESI):$ *m/z*calcd. for C₂₂₅H₁₉₂Eu₃F₂₇GdN₃₆O₅₁S₉ [Eu₃GdL3^{SS}₆ + 9 OTf⁻]³⁺: 1876.9166, found 1876.9222; Calcd. for C₂₂₃H₁₉₁Eu₃F₂₁GdN₃₆O₄₅S₇ [Eu₃GdL3^{SS}₆ + 7 OTf⁻ - H⁺]⁴⁺: 1332.9595, found 1335.9584; Calcd. for C₂₂₃H₁₉₂Eu₃F₂₁GdN₃₆O₄₅S₇ [Eu₃GdL3^{SS}₆ + 7 OTf⁻ - H⁺]⁴⁺: 0Tf⁻]⁵⁺: 1066.5690, found 1066.5699; Calcd. for C₂₂₂H₁₉₁Eu₃F₁₈GdN₃₆O₄₂S₆ [Eu₃GdL3^{SS}₆ + 7 OTf⁻ - H⁺]⁵⁺: 1036.3769, found 1036.3811; Calcd. for C₂₂₀H₁₉₀Eu₃F₁₂GdN₃₆O₃₆S₄ [Eu₃GdL3^{SS}₆ + 4 OTf⁻ - 2H⁺]⁶⁺: 813.8287, found 813.8317. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆Eu₃GdN₃₆O₆₀S₁₂·14H₂O: C 43.26, H 3.50, N 7.97, found: C 43.20, H 3.32, N 7.92.

 $[Eu_nGd_{4-n}L3^{RR}_6] \text{ (prepared by ratio Eu/Gd: 3/1): (1.57 mg, 2.58 x 10⁻⁴ mmol, 78.3\% yield), HRMS (ESI):$ *m/z* $calcd. for C₂₂₅H₁₉₂Eu₃F₂₇GdN₃₆O₅₁S₉ [Eu₃GdL3^{RR}_6 + 9 OTf⁻]³⁺: 1876.9166, found 1876.9133; Calcd. for C₂₂₃H₁₉₂Eu₃F₂₁GdN₃₆O₄₅S₇ [Eu₃GdL3^{RR}_6 + 7 OTf⁻]⁵⁺: 1066.5690, found 1066.5623; Calcd. for C₂₂₂H₁₉₁Eu₃F₁₈GdN₃₆O₄₂S₆ [Eu₃GdL3^{RR}_6 + 7 OTf⁻]⁵⁺: 1036.3769, found 1036.3728; Calcd. for C₂₂₁H₁₉₀Eu₃F₁₅GdN₃₆O₃₉S₅ [Eu₃GdL3^{RR}_6 + 5 OTf⁻ - 2H⁺]⁵⁺: 1006.3850, found 1006.3813; Calcd. for C₂₂₀H₁₉₀Eu₃F₁₂GdN₃₆O₃₆S₄ [Eu₃GdL3^{RR}_6 + 4 OTf⁻ - 2H⁺]⁶⁺: 813.8287, found 813.8316. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆Eu₃GdN₃₆O₆₀S₁₂·13H₂O: C 43.38, H 3.48, N 7.99, found: C 43.35, H 3.35, N 7.88.$

 $[Eu_nGd_{4-n}L3^{ss}_6] \text{ (prepared by ratio Eu/Gd: 1/1): (1.72 mg, 2.83 x 10^{-4} mmol, 85.7\% yield), } \\ HRMS (ESI): m/z \text{ calcd. for } C_{224}H_{192}Eu_2F_{24}Gd_2N_{36}O_{48}S_8 \text{ [Eu_2Gd_2L3^{SS}_6 + 8 OTf^-]^{4+}: } \\ 1371.7002, \text{ found } 1371.7015; \text{ Calcd. for } C_{223}H_{192}Eu_2F_{21}Gd_2N_{36}O_{45}S_7 \text{ [Eu_2Gd_2L3^{SS}_6 + 7 OTf^-]^{5+}: } \\ 1067.5697, \text{ found } 1067.5664; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ 1037.5777, \text{ found } 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}Gd_2N_{36}O_{42}S_6 \text{ [Eu_2Gd_2L3^{SS}_6 + 6 OTf^- - H^+]^{5+}: } \\ \\ 1037.5746; \text{ Calcd. for } C_{222}H_{191}Eu_2F_{18}O_{22}O_{$

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 $C_{222}H_{192}Eu_2F_{18}Gd_2N_{36}O_{42}S_6$ [Eu₂Gd₂L**3**^{SS}₆ + 6 OTf⁻]⁶⁺: 864.8160, found 864.8126; Calcd. for $C_{220}H_{190}Eu_2F_{12}Gd_2N_{36}O_{36}S_4$ [Eu₂Gd₂L**3**^{SS}₆ + 4 OTf⁻ - 2H⁺]⁶⁺: 814.8295, found 814.8297. Elemental analysis calcd. for $C_{228}H_{192}F_{36}Eu_2Gd_2N_{36}O_{60}S_{12}\cdot 14H_2O$: C 43.22, H 3.50, N 7.96, found: C 43.26, H 3.54, N 8.00.

 $[Eu_nGd_{4-n}L3^{RR}_6] \text{ (prepared by ratio Eu/Gd: 1/1): (1.70 mg, 2.79 x 10⁻⁴ mmol, 84.7% yield), HRMS (ESI):$ *m/z*calcd. for C₂₂₅H₁₉₂Eu₂F₂₇Gd₂N₃₆O₅₁S₉ [Eu₂Gd₂L3^{RR}₆ + 9 OTf⁻]³⁺: 1878.5844, found 1878.5778; Calcd. for C₂₂₃H₁₉₂Eu₂F₂₁Gd₂N₃₆O₄₅S₇ [Eu₂Gd₂L3^{RR}₆ + 7 OTf⁻]⁵⁺: 1067.5697, found 1067.5664; Calcd. for C₂₂₁H₁₉₀Eu₂F₁₅Gd₂N₃₆O₃₉S₅ [Eu₂Gd₂L3^{RR}₆ + 5 OTf⁻ - 2H⁺]⁵⁺: 1007.5858, found 1007.5864; Calcd. for C₂₂₂H₁₉₂Eu₂F₁₈Gd₂N₃₆O₄₂S₆ [Eu₂Gd₂L3^{RR}₆ + 6 OTf⁻]⁶⁺: 864.8160, found 864.8129; Calcd. for C₂₂₀H₁₉₀Eu₂F₁₂Gd₂N₃₆O₃₆S₄ [Eu₂Gd₂L3^{RR}₆ + 4 OTf⁻ - 2H⁺]⁶⁺: 814.8295, found 814.8281. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆Eu₂Gd₂N₃₆O₆₀S₁₂·13H₂O: C 43.35, H 3.48, N 7.98, found: C 43.36, H 3.39, N 7.92.

 $[Eu_{n}Gd_{4-n}L3^{SS}_{6}] \text{ (prepared by ratio Eu/Gd: 1/3): (1.66 mg, 2.73 x 10⁻⁴ mmol, 82.6% yield), } \\ HRMS (ESI):$ *m/z* $calcd. for C₂₂₅H₁₉₂EuF₂₇Gd_3N₃₆O₅₁S₉ [EuGd_3L3^{SS}₆ + 9 OTf⁻]³⁺: 1880.2521, found 1880.2518; Calcd. for C₂₂₄H₁₉₂EuF₂₄Gd_3N₃₆O₄₈S₈ [EuGd_3L3^{SS}₆ + 8 OTf⁻]⁴⁺: 1372.9510, found 1372.9503; Calcd. for C₂₂₃H₁₉₂EuF₂₁Gd_3N₃₆O₄₅S₇ [EuGd_3L3^{SS}₆ + 7 OTf⁻]⁵⁺: 1068.5703, found 1068.5665; Calcd. for C₂₂₂H₁₉₁EuF₁₈Gd_3N₃₆O₄₂S₆ [Eu₂Gd₂L3^{SS}₆ + 6 OTf⁻ - H⁺]⁵⁺: 1038.5784, found 1038.5743; Calcd. for C₂₂₀H₁₉₀EuF₁₂Gd_3N₃₆O₃₆S₄ [Eu₂Gd₂L3^{SS}₆ + 4 OTf⁻ - 2H⁺]⁶⁺: 815.6633, found 815.6631. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆EuGd_3N₃₆O₆₀S₁₂·15H₂O·C₆H₁₄O: C 43.50, H 3.68, N 7.80, found: C 43.43, H 3.68, N 7.82.$

 $[Eu_nGd_{4-n}L3^{RR}_6] \text{ (prepared by ratio Eu/Gd: 1/3): (1.75 mg, 2.87 x 10⁻⁴ mmol, 87.1% yield), HRMS (ESI):$ *m/z* $calcd. for C₂₂₅H₁₉₂EuF₂₇Gd₃N₃₆O₅₁S₉ [EuGd₃L3^{RR}_6 + 9 OTf⁻]³⁺: 1880.2521, found 1880.2430; Calcd. for C₂₂₄H₁₉₂EuF₂₄Gd₃N₃₆O₄₈S₈ [EuGd₃L3^{RR}_6 + 8 OTf⁻]⁴⁺: 1372.9510, found 1372.9489; Calcd. for C₂₂₁H₁₉₀EuF₁₅Gd₃N₃₆O₃₉S₅ [EuGd₃L3^{RR}_6 + 5 OTf⁻ - 2H⁺]⁵⁺: 1008.5864, 1008.5843; Calcd. for C₂₂₀H₁₉₀EuF₁₂Gd₃N₃₆O₃₆S₄ [EuGd₃L3^{RR}_6 + 4 OTf⁻ - 2H⁺]⁶⁺: 815.6633, found 815.6630; Calcd. for C₂₂₀H₁₉₂EuF₁₂Gd₃N₃₆O₃₆S₄ [EuGd₃L3^{RR}_6 + 4 OTf⁻ - 2H⁺]⁷⁺: 699.2839, found 699.2811. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆EuGd₃N₃₆O₆₀S₁₂·15H₂O·C₆H₁₄O: C 43.50, H 3.68, N 7.80, found: C 43.53, H 3.42, N 7.89.$

 $[Eu_nSm_{4-n}L3^{SS}_6] \text{ (prepared by ratio Eu/Sm: 1/1): (1.79 mg, 2.95 x 10^{-4} mmol, 89.4\% yield), HRMS (ESI):$ *m/z*calcd. for C₂₂₃H₁₉₂Eu₂F₂₁N₃₆O₄₅S₇Sm₂ [Eu₂Sm₂L3^{SS}₆ + 7 OTf⁻]⁵⁺: 1064.7673, found 1064.7710; Calcd. for C₂₂₂H₁₉₁Eu₂F₁₈N₃₆O₄₂S₆Sm₂ [Eu₂Sm₂L3^{SS}₆ + 6 OTf⁻ - H⁺]⁵⁺: 1034.7754, found 1034.7769; Calcd. for C₂₂₁H₁₉₀Eu₂F₁₅N₃₆O₃₉S₅Sm₂ [Eu₂Sm₂L3^{SS}₆ + 5 OTf⁻ - 2H⁺]⁵⁺: 1004.7834, found 1004.7823; Calcd. for C₂₂₂H₁₉₂Eu₂F₁₈N₃₆O₄₂S₆Sm₂ [Eu₂Sm₂L3^{SS}₆ + 6 OTf⁻]⁶⁺: 862.4807, found 862.4775; Calcd. for C₂₂₀H₁₉₁Eu₂F₁₂N₃₆O₃₆S₄Sm₂ [Eu₂Sm₂L3^{SS}₆ + 4 OTf⁻ - H⁺]⁷⁺: 696.5675, 696.5658. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆Eu₂Sm₂N₃₆O₆₀S₁₂·14H₂O·2(C₆H₁₄O): C 44.17, H 3.83, N 7.73, found: C 44.11, H 3.54, N 7.75.

 $[Eu_{n}Sm_{4-n}L3^{RR}_{6}] \text{ (prepared by ratio Eu/Sm: 1/1): (1.68 mg, 2.77 x 10⁻⁴ mmol, 83.9% yield), HRMS (ESI):$ *m/z*calcd. for C₂₂₅H₁₉₂Eu₂F₂₇N₃₆O₅₁S₉Sm₂ [Eu₂Sm₂L3^{RR}₆ + 9 OTf⁻]³⁺: 1873.9137, found 1873.9077; Calcd. for C₂₂₄H₁₉₂Eu₂F₂₄N₃₆O₄₈S₈Sm₂ [Eu₂Sm₂L3^{RR}₆ + 8 OTf⁻]⁴⁺: 1368.1972, found 1368.1980; Calcd. for C₂₂₃H₁₉₁Eu₂F₂₁N₃₆O₄₅S₇Sm₂ [Eu₂Sm₂L3^{RR}₆ + 7 OTf⁻ - H⁺]⁴⁺: 1330.7073, found 1330.7100; Calcd. for

 $\begin{aligned} C_{223}H_{192}Eu_2F_{21}N_{36}O_{45}S_7Sm_2 & [Eu_2Sm_2\textbf{L3}^{\textbf{RR}}_6 + 7 \text{ OTf}^{-}]^{5+}: 1064.7673, \text{ found } 1064.7668; \\ Calcd. \text{ for } C_{222}H_{191}Eu_2F_{18}N_{36}O_{42}S_6Sm_2 & [Eu_2Sm_2\textbf{L3}^{\textbf{RR}}_6 + 6 \text{ OTf}^{-} - H^+]^{5+}: 1034.7754, \text{ found } \\ 1034.7715; \text{ Elemental analysis calcd. for } C_{228}H_{192}F_{36}Eu_2Sm_2N_{36}O_{60}S_{12}\cdot 12H_2O\cdot 2(C_6H_{14}O): \\ C 44.42, H 3.79, N 7.77, \text{ found: } C 44.40, H 3.82, N 7.80. \end{aligned}$

Dy4L3^{SS}₆: (1.66 mg, 2.71 x 10⁻⁴ mmol, 82.4% yield), HRMS (ESI): m/z calcd. for C₂₂₃H₁₉₂Dy4F₂₁N₃₆O₄₅S₇ [Dy4L3^{SS}₆ + 7 OTf⁻]⁵⁺: 1073.9743, found 1073.9764; Calcd for C₂₂₂H₁₉₁Dy4F₁₈N₃₆O₄₂S₆ [Dy4L3^{SS}₆ + 6 OTf⁻ H⁺]⁵⁺: 1043.7821, found 1043.7826; Calcd. for C₂₂₁H₁₉₀Dy4F₁₅N₃₆O₃₉S₅ [Dy4L3^{SS}₆ + 5 OTf⁻ 2H⁺]⁵⁺: 1013.7902, found 1013.7969; Calcd. for C₂₂₀H₁₈₉Dy4F₁₂N₃₆O₃₆S₄ [Dy4L3^{SS}₆ + 4 OTf⁻ 3H⁺]⁵⁺: 983.7983, found 983.7963; Calcd. for C₂₂₀H₁₉₀Dy4F₁₂N₃₆O₃₆S₄ [Dy4L3^{SS}₆ + 4 OTf⁻ 2H⁺]⁶⁺: 819.9998, found 820.0034. Elemental analysis calcd. for C₂₂₈H₁₉₂F₃₆ Dy₃TbN₃₆O₆₀S₁₂·11H₂O·C₆H₁₄O: C 43.81, H 3.58, N 7.86, found: C 43.88, H 3.63, N 7.46. 3.4.2 ESI-HRMS deconvolution analysis

	Precentage (%)
[Eu ₄ L 3 ₆ + 9OTf ⁻] ³⁺	2.85
[Eu ₃ Gd L3 ₆ + 9OTf ⁻] ³⁺	19.68
[Eu ₂ Gd ₂ L 3 ₆ + 9OTf ⁻] ³⁺	45.77
[EuGd ₃ L 3 ₆ + 9OTf ⁻] ³⁺	26.64
[Gd₄ L3 ₆ + 9OTf ⁻] ³⁺	5.07

Table S3-1. Percentage of the selected ESI-HRMS peak for $[Eu_nGd_{4-n}L3_6]$ (n = 0-4), Eu/Gd: 1:1.

	Percentage (%)
[Eu4 L3 6 + 90Tf ⁻] ³⁺	2.74
[Eu ₃ Tb L3 ₆ + 9OTf ⁻] ³⁺	17.70
[Eu ₂ Tb ₂ L 3 ₆ + 9OTf ⁻] ³⁺	45.33
[EuTb ₃ L 3 ₆ + 9OTf ⁻] ³⁺	29.84
[Eu4 L3 6 + 9OTf ⁻] ³⁺	4.39

Table S3-2. Percenatge of the selected ESI-HRMS peak for $[Eu_nTb_{4-n}L3_6]$ (n = 0-4), Eu/Tb: 1:1.

	Percentage (%)
[Eu ₄ L3 ₆ + 9OTf ⁻] ³⁺	59.07
[Eu ₃ Lu L3 ₆ + 9OTf ⁻] ³⁺	25.88
[Eu ₂ Lu ₂ L 3 ₆ + 9OTf ⁻] ³⁺	8.55
[EuLu ₃ L3 ₆ + 9OTf ⁻] ³⁺	2.55
[Lu ₄ L3 ₆ + 9OTf ⁻] ³⁺	3.96

Table S3-3. Percenatge of the selected ESI-HRMS peak for $[Eu_nLu_{4-n}L3_6]$ (n = 0-4), Eu/Lu: 1:1.

	Percentage
[Eu4 L3 6 + 9OTf ⁻] ³⁺	25.88
[Eu ₃ Gd L3 ₆ + 9OTf ⁻] ³⁺	53.37
[Eu ₂ Gd ₂ L 3 ₆ + 9OTf ⁻] ³⁺	17.92
[EuGd ₃ L 3 ₆ + 9OTf ⁻] ³⁺	3.89
[Gd4 L3 6 + 9OTf ⁻] ³⁺	0

Table S3-4. Percentage of the selected ESI-HRMS peak for $[Eu_nGd_{4-n}L3_6]$ (n = 0-4), Eu/Gd: 3:1.

	Percentage
[Eu4 L3 6 + 9OTf ⁻] ³⁺	0
[Eu ₃ Gd L3 ₆ + 9OTf ⁻] ³⁺	2.66
[Eu ₂ Gd ₂ L3 ₆ + 9OTf ⁻] ³⁺	13.33
[EuGd ₃ L 3 ₆ + 9OTf ⁻] ³⁺	57.06
[Gd4 L3 6 + 9OTf ⁻] ³⁺	27.30

Table S3-5. Percentage of the selected ESI-HRMS peak for $[Eu_nGd_{4-n}L3_6]$ (n = 0-4), Eu/Gd: 1:3.

3.4.3 Photophysical measurement

		<u>g</u> lum							
		-		[EunGd4	[EunGd4			[EunGd4	[EunGd4
				₋n L3 6	-n L3 6 ^{кк}]	[EunGd4-	[EunGd₄-	-n L3 6	-n L3 6 ^{кк}]
Electro				(n = 0-	(n = 0-	ո Լ3 ցss] (n	n L3 6 ^{RR}] (n	(n = 0-	(n = 0-
nic	λ			4),	4),	= 0-4),	= 0-4),	4),	4),
transiti	(nm	[Eu4 L3^s	[Eu4 L3 ^R	Eu/Gd:	Eu/Gd:	Eu/Gd:	Eu/Gd:	Eu/Gd:	Eu/Gd:
on)	s ₆]	^R ₆]	3:1	3:1	1:1	1:1	1:3	1:3
$^{5}D_{0} \rightarrow$	587	0.15	-0.15	0.14	-0.15	0.15	-0.15	0.14	-0.15
′F1	601	0.06	-0.06	0.05	-0.06	0.06	-0.05	0.06	-0.06
$^{5}D_{0} \rightarrow$ $^{7}F_{2}$	613	-0.05	0.04	0.04	-0.05	-0.05	0.05	-0.04	0.04
$^{5}D_{0} \rightarrow$ $^{7}F_{3}$	652	0.12	-0.11	0.10	-0.10	0.06	-0.06	0.03	-0.04
50)	685	0.01	-0.01	0.01	-0.01	0.02	-0.01	0.02	-0.01
$^{5}D_{0} \rightarrow$ $^{7}F_{4}$	697	-0.04	0.04	-0.04	0.04	-0.04	0.04	-0.05	0.05
	704	0.12	-0.12	0.10	-0.11	0.11	-0.10	0.07	-0.07

Table S3-6. Summary of CPL results for the eight homometallic and heterometallictetrahedra.

	$\lambda_{abs}{}^{max}$	ϵ^{max}	$\lambda_{ ext{em}}{}^{ ext{max}}$	ф _х ^ь (%)	τ
	(nm)	(L·mol ^{−1} ·cm ^{−1})	(nm)		(ms)
[Eu ₄ (L3^{SS}) ₆](CF ₃ SO ₃) ₁₂	330	121335.7	616	0.81 (0.035)	1.28
[Eu4(L3^{RR}) 6](CF3SO3)12	330	123947.0	616	0.83 (0.035)	1.27
[EunGd _{4-n} L 3 6 ^{ss}] (n = 0-4), Eu/Gd: 3:1	330	122963.9	616	0.76 (0.03)	1.27
[EunGd _{4-n} L 3 6 ^{RR}] (n = 0-4), Eu/Gd: 3:1	330	124822.0	616	0.70 (0.025)	1.27
[EunGd _{4-n} L3 ₆ ss] (n = 0-4), Eu/Gd: 1:1	330	124869.7	616	0.49 (0.025)	1.28
[EunGd _{4-n} L 3 6 ^{RR}] (n = 0-4), Eu/Gd: 1:1	330	125035.4	616	0.50 (0.02)	1.27
[EunGd _{4-n} L3 ₆ ^{ss}] (n = 0-4), Eu/Gd: 1:3	330	123626.7	616	0.30 (0.02)	1.27
[EunGd _{4-n} L 3 6 ^{RR}] (n = 0-4), Eu/Gd: 1:3	330	131345.8	616	0.34 (0.03)	1.27
$[Gd_4(L3^{SS})_6](CF_3SO_3)_{12}$	330	121829.6	/	/	/
$[Gd_4(L3^{RR})_6](CF_3SO_3)_{12}$	330	123657.1	466	/	0.0087 ^c
[Eu _n Sm _{4-n} L3 ₆ ^{ss}] (n = 0-4), Eu/Sm: 1:1	330	130409.0	616	0.33 (0.045)	1.27
[EunSm _{4-n} L3 ₆ ^{RR}] (n = 0-4), Eu/Sm: 1:1	330	119229.7	616	0.34 (0.03)	1.28

Table S3-7. A summary of selected photophysical properties, UV-Vis absorption and luminescence data of lanthanide complexes in acetonitrile solution^a. ^ausing a 10 mm cuvette and filter 380 nm. ^bThe relative quantum yields were referenced with quinine sulfate in 0.1 M sulfuric acid (ϕ = 0.577, λ_{ex} = 350nm) with 10 mm cuvette. ^cMeasurement performed at 77K in 1:4 MeOH/EtOH.

3.4.4 Crystal structure of $[Eu_nGd_{4-n}L3_6]$ (n = 0-4)

Crystal

Data

for

 $C_{224.998875}H_{191.99904}Eu_{1.99999}F_{26.999865}Gd_{1.99999}N_{35.99982}O_{50.999745}S_{8.999955}$ (*M* =5636.07 g/mol): trigonal, space group R3 (no. 146), a = 30.0304(9) Å, c = 71.671(3) Å, V = 55975(4) Å³, Z = 6.00003, T = 220.0 K, μ (CuK α) = 5.684 mm⁻¹, Dcalc = 1.003 g/cm³, 70262 reflections measured (5.886° $\leq 2\Theta \leq 117.84$ °), 34927 unique ($R_{int} = 0.0543$, $R_{sigma} = 0.0620$) which were used in all calculations. The final R_1 was 0.0655 (I > 2 σ (I)) and wR₂ was 0.1960 (all data). The crystallographic data for the structural analyses have been deposited with the Cambridge Crystallographic Data Centre, CCDC No. 2092789, and the data can be obtained free of charge via www.ccdc.cam.ac.uk/data request/cif.

Information	Identifier from cif file
Empirical formula	C ₂₂₅ H ₁₉₂ Eu ₂ F ₂₇ Gd ₂ N ₃₆ O ₅₁ S ₉
Formula weight	5636.07
Temperature/K	220.0
Crystal system	trigonal
Space group	R3
a/Å	30.0304(9)
b/Å	30.0304(9)
c/Å	71.671(3)
α/°	90
β/°	90
γ/°	120
Volume/Å ³	55975(4)
Z	6.00003
ρ _{calc} g/cm ³	1.003
µ/mm⁻¹	5.684
F(000)	17058.0
Crystal size/mm ³	0.23 × 0.18 × 0.13
Radiation	CuKα (λ = 1.54178)

20 range for data collection/°	5.886 to 117.84
Index ranges	-33 ≤ h ≤ 33, -33 ≤ k ≤ 33, -79 ≤ l ≤ 79
Reflections collected	70262
Independent reflections	34927 [R _{int} = 0.0543, R _{sigma} = 0.0620]
Data/restraints/parameters	34927/2794/1969
Goodness-of-fit on F ²	0.983
Final R indexes [I>=2σ (I)]	R ₁ = 0.0655, wR ₂ = 0.1771
Final R indexes [all data]	R ₁ = 0.0805, wR ₂ = 0.1960
Largest diff. peak/hole / e Å ⁻³	0.80/-0.82
Flack parameter	0.011(3)

	Shape measure (°)					
	Tricapped trigonal prism	Capped square antiprism				
	(D _{3h})	(C _{4v})				
Eu1	5.70	10.30				
Eu2	5.70	10.30				
Eu3	5.78	10.79				
Eu4	5.70	10.30				

Table S3-8. Results of the Shape Analysis for Eu₂Gd₂(L3^{SS})₆

Shape analysis was performed using the software that developed by Raymond's group.³⁵⁻³⁶ The coordination geometry of the $Eu_2Gd_2(L3^{ss})_6$ can be best described as tricapped trigonal prism.

3.5 References

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Chapter 4: Remote stereo-control of chiral lanthanide supramolecular complexes

4.1 Introduction

4.1.1 Chiral domino effect induced by α , α -disubstituted amino acids

Helical peptides are inherently chiral even if they lack asymmetric centers in the peptide backbone.¹ However, isomers of the helical peptides are isoenergetic and thus they have an equal population in the achiral environment. Control chirality of peptides is essential for biological application. Therefore, scientists were interested in restricting the conformation of peptides through chiral domino effect.

'Domino effect' is a term that describes the transfer and amplification of chiral information from the chiral moiety to the rest of the achiral moiety. For covalent chiral domino effect on peptides, it strongly relies on the presence of chiral amino acid units to transfer the chiral information throughout the entire peptide.²⁻⁴



Figure 4-1. Structures of Aib and Ac₆c.

One of the solutions is the use of α , α -disubstituted amino acids.⁵ Replacement of the α -hydrogen with an alkyl substituent can increase the chemical stability and restrict the conformational freedom of the peptide. Common α , α -disubstituted amino acids include α -aminoisobutyric acid (Aib) and 1-aminocyclohexanecarboxylic acid (Ac₆c) (figure 4-1).⁶

The secondary structures of peptide containing Aib has been well investigated. Homopeptide consists of Aib is preferred to form 3_{10} helicates in both solution and crystal state. The enantiomeric ratio of right-handed helicates and left-handed helicates is 1 to $1.^{7.8}$ There is no critical chain length for the formation of 3_{10} helicates.⁹ The minimal number of amino acids for 3_{10} helicates might be 3. When the L- α -amino acid residue is incorporated with Aib units, right-handed helicates will be preferred. It is found that the peptides prefer to form 3_{10} helicates when it is short (<7 amino acid units) and containing high percentage of Aib (>50%),¹⁰ while α -helices will be formed when the peptide is long (>9 amino acid units) and contains little Aib residue (<50%).¹¹

4.1.2 Remote control of chirality in transition metal complexes

Control of the chirality of the self-assembly supramolecule is one of the major research areas. In most of the examples, chirality of the metal complexes can be successfully controlled by the chiral group in the ligand through covalent domino effect.¹² However, the position of the chiral group should be close enough to the chelating group so that the chiral information could be transmitted to the metal ion.

In 2011, Yashima's group reported an example of remote control of chirality of a metal complex.¹³ The complex was coordinated by a ligand bearing a helical oligopeptide. The general idea of the remote control of chirality was based on the covalent chiral domino effect. Although the chiral Val residue was located far from the metal center, chiral information can be still transferred through the strongly helicogenic achiral peptide. The mechanism involved the transfer of chiral information along the whole peptide, resulting in either *P*- (right-handed twisted) or *M*- (left-handed twisted) helix. The handedness can control the chirality of the metal center. It was found the most effective distance is 4 amino acid units (n=2), while no stereocontrol effect can be found when n =0 or 1.

Based on the result in 2011, Yashima's group further demonstrated the concept of remote control of chirality by helical peptides with self-assembly supramolecular complexes (figure 4-2).¹⁴ One of the interesting results was the transformation between the 3_{10} helicates and α -helix by the addition of fluoroalcohols. The α -helix will freeze the rotation of the peptide because of the increased helix width. As a result, the complex containing higher 3_{10} helicates content will have a stronger CD intensity compared with that of α -helix.



Figure 4-2. Remote control of chirality by helical peptide supramolecular Ni complex.¹⁴

4.1.3 Scope of study

In this study, the concept of remote control of chirality was applied to lanthanide complexes. Chiral ligands consist of short amino acid sequences were prepared. Position, length and types of amino acid was varied to investigate the effect of peptide towards the supramolecular formation. It was expected that the chiral information could be transmitted by chiral domino effect induced by the helical twisting of bulky disubstituted amino acid units.

4.2 Result and Discussion



4.2.1 Scheme of this study

Figure 4-3. Structures of L5-10.

In this chapter, six ligands were synthesized and their structures are shown in figure 4-3. **L5** was designed based on literature and it was expected to form chiral lanthanide supramolecular tetrahedron through chiral domino effect. However, no desired supramolecular product was observed in both ¹HNMR and ESI-HRMS. Therefore, five different ligands were synthesized and the supramolecular formation using peptide-consisted ligands was investigated. The scheme of this study was shown in figure 4-4.



Figure 4-4. Scheme showing the investigation of supramolecular formation using peptide ligands.

4.2.2 Ligand design of L5

L5 was designed based on the reported literature. The ligand consists of four parts, which are the chiral group, peptide, metal chelator and bridging unit. The chiral group and metal chelator are chosen to be 1-phenylethan-1-amine and pyridinedicarboxylamide (pcam). The designed bridging 2,6unit is diaminoanthracene-9,10-dione. The incorporation of the above three parts with europium shows bright red luminescence. The peptide part would be composed of four Aib residues, which was found to have a strong helix-forming ability due to its steric hinderance. It was expected that the chirality can be transmitted from the chiral group to the metal center through covalent chiral effect.



Figure 4-5. The component of the proposed L5.

4.2.3 Synthesis of Aib intermediates

<u>Step 1:</u>



Step 1 involved the esterification of Boc-Aib-OH using iodomethane. The desired Boc-Aib-OMe was successfully synthesized and the highest yield that obtained was 89.8%.

Step 2:



In the next step, TFA was used to deprotect the Boc group in the amino acid. The highest yield that obtained was 88.15%.

Step 3:



Boc-Aib-Aib-OMe was prepared by coupling H₃N⁺-Aib-OMe and Boc-Aib-OH using PyBOP as the coupling reagent. The amide coupled product can be purified by simply washing with acid and base. Result showed that 2 M HCl will deprotect the Boc group. Therefore, the concentration of acid was set to 0.5 M. Different coupling reagents were tested in this step including thinoyl chloride, oxalyl chloride, HOBt and EEDQ. However, PyBOP was found to be the most efficient coupling reagent since the
yield was high (up to 75.0%) and the reaction time was relatively short (16 hrs) compared to other coupling reagent.

Step 4:



Boc-Aib-Aib-Aib-OMe was prepared by in-situ deprotection of Boc-Aib-Aib-OMe followed by amide coupling reaction with Boc-Aib-OH. Different from general procedure, which suggested that deprotection of Boc group can be completed within an hour with the use of TFA, 3 hrs were required for deprotection in this step. The crude Boc-Aib-Aib-Aib-OMe can be purified simply by washing with acid, base and diethyl ether.

<u>Step 5:</u>

Boc-(Aib)₄-OMe was synthesized by in-situ deprotection of Boc-Aib-Aib-Aib-OMe followed by coupling with Boc-Aib-OH. The highest yield was found to be 54.3%.

<u>Step 6:</u>



Boc-(Aib)₄-OH was prepared by alkaline hydrolysis to Boc-(Aib)₄-OMe to deprotect the methyl ester group. The highest yield was found to be 85.1%.

<u>Step 7:</u>

Boc-(Aib)₄-spy was synthesized by classical HOBt, EDC coupling and purified by washing with acid and base. The product was only slightly soluble in ethyl acetate and diethyl ether, and highly soluble in DCM.

<u>Step 8:</u>



Boc-Aib-Aib-OH was successfully coupled with chiral group following the procedure of Boc-(Aib)₄-spy. The highest yield was 92.3%.

4.2.4 Synthesis of central bridging linker

Step 1:



Di-tert-butyl 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6diyl)bis(azanediyl))bis(carbonyl))dipicolinate was prepared by coupling two equivalents of 6-(tert-butoxycarbonyl)picolinic acid with one equivalent of 2,6diaminoanthracene-9,10-dione for 32 hrs. The pure product could be obtained by simply washing the crude product with water and MeCN. The yield was found to be 92.4%.

<u>Step 2:</u>



The central bridging linker of 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid was prepared by deprotection of tbutyl group in DCM. After 16 hrs, the solvent was evaporated, and the crude product was washed with diethyl ether. The highest yield that obtained for this step is 99.3% yield.



4.2.5 Synthesis and complexation of L5



L5 was synthesized by in-situ deprotection of Boc-(Aib)₄-spy followed by coupling with the central bridging linker 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid using HATU as the coupling reagent. The ligand was purified by simply washing with MeCN to obtain 49.6% yield.

Three equivalents of L5 was allowed to coordinate with two equivalents of Eu(OTf)₃ in a mixture of MeCN and DCM at 60°C for 3 hrs. ¹HNMR revealed the disappearance of ligand peaks, indicating the successful coordination of ligand with Eu. Compared with free ligand, the NMR signals arising from the complex are shifted due to the coordination to paramagnetic europium(III) ions. Diamagnetic La complex was also synthesized by replacing Eu with La and the ¹HNMR was done to confirm the complexation. However, compared the NMR spectrum with other lanthanide complexes from chapter 2 and 3, the NMR of Eu and La complexes with L5 were weak (figure 4-6). ESI-HRMS was also performed for both Eu and La complexes. In the ESI-HRMS specta, mononuclear species LnL5 was found to be the only species in solution state and no other higher order supramolecular complex can be observed (figure 4-7). For the structure of LnL5, it was hypothesized that one Ln ion coordinate with one L5 and the coordination sphere of Ln was saturated by both ligand and solvent molecules. However, due to the lack of X-ray crystal structure, exact determination of structure of LnL5 was not possible. Result showed that the formation of desired tetrahedron was unsuccessful using L5.

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Figure 4-6. ¹HNMR of (a) free ligand L5 in (CD₃)₂SO, (b) Eu complex of L5 in CD₃CN and (c) La complex

of **L5** in CD₃CN.



Figure 4-7. ESI-HRMS of (a) Eu complex of L5 (b) La complex of L5.

4.2.6 Synthesis and complexation of L6



Based on the result from L5, it showed that L5 can successfully coordinate with Eu and La. However, there were some factors that affect the supramolecular formation. Thus, L6 was prepared to compare the result with L5. Different from L5, L6 only consists of two Aib units. It was expected to form 3₁₀ helix so that the chiral information can be transferred from the chiral group to the metal centre. It was hypothesized that large number of carbonyl oxygen may affect the lanthanide supramolecular formation since carbonyl oxygen may act as a donor to coordinate with lanthanide ions. Therefore, the number of carbonyl oxygen was reduced in L6. Similar to **L5**, **L6** was synthesized by in-situ deprotection of Boc-Aib-Aib-spy followed by coupling with the central bridging linker 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid using HATU as the coupling reagent. The product can be simply washed with chloroform and the yield was found to be 53.6%.

Three equivalents of **L6** was allowed to coordinate with two equivalents of Eu(OTf)₃ in a mixture of MeCN and DCM at 60°C for 3 hrs. In the ¹HNMR spectrum of the Eu complex, similar result was observed compared with that of **L5**. All signals corresponding to the ligand were disappeared and broad peaks were observed due to the paramagnetic europium(III) ions (figure 4-8).



Figure 4-8. ¹*HNMR of (a) free ligand L6 in (CD₃)*₂*SO and (b) Eu complex of L5 in CD*₃*CN.*

The Eu complex of **L6** was further characterized by ESI-HMRS (figure 4-9). In the ESI-HRMS spectrum, the resolved peaks are either in +2 and +3 charge and the major species was found to be Eu_2L6_2 . No desired tetrahedron or helicate can be observed.



Figure 4-9. ESI-HRMS of Eu complex of L6.

4.2.7 Synthesis of Ac₆c intermediates



Two more ligands, L7 and L8, were designed and synthesized. Different from L5 and L6, both L7 and L7 consist of another type of bulky amino acid units. Ac₆c amino acid was used for chiral domino effect. Complexation was done for L7 and L8 to observe the effect of changing another type of amino acid towards supramolecular formation. In order to avoid the coordination between metal ion and carbonyl oxygen, the number of amino acids was greatly reduced (one for L7, and two for L8).



The first step is to prepare the H_2N-Ac_6c -OMe. According to the literature, this step can be achieved by using thionyl chloride and methanol and the carboxylic acid group can be protected by a methyl group. The following table shows five trials for this reaction.

Trial	Reaction time	Reagent	Yield/Result
1	16 hrs	Thionyl chloride	30.2%
2	16 hrs	Oxalyl chloride	No product can be obtained
3	96 hrs		35.9%
4	7 days	Thionyl chloride	46.1%
5	10 days		No product can be obtained

*Table 4-1. Trials for synthesizing H-Ac*₆*c-OMe.*

Although this method can synthesize the product successfully, the yield is relatively low. The unreacted thionyl chloride is difficult to be removed and some of the product would be precipitated out in a form of acidic salt. The acidic product may affect the next step reaction since the amide coupling reaction requires a basic environment.

An alternative synthetic procedure to prepare H_2N-Ac_6c -OMe. Although the H_3N-Ac_6c -OMe could be synthesized by the Boc group protection of the $H-Ac_6c$ -OH followed by the methylation by iodomethane and TFA deprotection in good yield.

However, three steps are required, and the reaction time is long in order to achieve high yield. Therefore, an alternative method is needed to increase the efficiency.

The new method is based on the findings by Sha's group.¹⁵ In 2008, Sha's group reported a convenient synthesis of amino acid methyl esters by employing trimethylchlorosilane (TMSCI) as the methylation reagent. The result shows that the use of TMSCI could obtain a high yield, which is comparable or even higher than those obtained with the use of thionyl chloride. TMSCI has a relatively lower boiling point compared with thionyl chloride which is easier for purification. Several attempts were done, and the results are summarized in the following table.

Trial	Reaction Time (hrs)	Temperature (°C)	Yield
1	16	rtp	22.8%
2	24	rtp	28.3%
3	16	50	51.5%
4	24	50	52.9%

Table 4-2. Trials for synthesizing H₂N -Ac₆c-OMe by TMSCI.

Based on the result, the condition in trial 3 is the most efficient since the reaction time is shorter and it provides a reasonable yield.

<u>Step 2:</u>

$$H_2N$$
 H_2N H_2O H_2O

The second step is to prepare the Boc-Ac₆c-OH. Several trials were tested to prepare it since the Boc-protection reaction is simple. The following table summarized the details of different trials.

Trial	Base	Solvent	Reaction time	Temperature	Yield//Result
1	КОН	THF/Water	16 hrs	0 °C	5.21%
2	TEA	DCM	16 hrs	rtp	No product can be found in NMR
3	NaOH	MeOH/Water	16 hrs	0 °C	No product can be found in NMR
4			16 hrs	0 °C	27.0%
5	NaOH	Dioxane	32 hrs	0 °C	46.4%
6			16 hrs	rtp	64.5%

 Table 4-3.
 Trials for synthesizing Boc-Ac₆c-OH.

Based on the theory of Boc group protection, the reaction should be done in anhydrous solvent since the reactant, Boc₂O, would react with water to generate carbon dioxide. Therefore, the first five trials were done in either rtp in dry solvent or 0°C in water in order to suppress the reaction between Boc₂O and water. Based on the result of trial 6, it shows that this reaction can also be done in water containing solvent and produce an acceptable yield.

<u>Step 3:</u>



Reaction 3 is the preparation of dipeptide Boc-Ac₆c-Ac₆c-OMe. It involves the coupling between Boc-Ac₆c-OH and H₂N-Ac₆c-OMe. Based on the experience in synthesizing the Boc-Aib-Aib-OMe, the dipeptide was the most challenging reaction. In the case of Boc-Aib-Aib-OMe, it was just a very short peptide and the hydrophobic character is not "obvious". For example, the Boc-Aib-Aib-OMe is slightly soluble in both hexane and diethyl ether, while the tripeptide (Boc-Aib-Aib-OMe) and tetrapeptide (Boc-Aib-Aib-Aib-Aib-OMe) are completely insoluble in hexane and diethyl ether. The solubility issue created a challenge in obtaining the pure product in high yield.

When preparing Boc-Ac₆c-Ac₆c-OMe, similar result was observed. The crude compound contained three UV-active impurities. The difference of the R_f value of the impurities and the desired product was quite large, but they cannot be completely separated by flash column chromatography. One of the attempts was the direct deprotection of methyl ester by lithium hydroxide monohydrate without further purification. Result shows that the UV-active impurities will not react with lithium hydroxide monohydrate and can be washed away by dichloromethane. The Boc-Ac₆c-Ac₆c-OH can be protonated and extracted by dichloromethane afterwards.

This reaction was repeated to prepare more intermediate compound and the reaction was summarized in the following table.

Trial	Yield
1	66.7%
2	73.0%

*Table 4-4. Trials for synthesizing Boc-Ac*₆*c-Ac*₆*c-OH.*



Boc-Ac₆c-Ac₆c-rpy was synthesized following the procedures of Boc-Aib-Aibspy. The reaction and workup were relatively easier, and the product can be purified by column chromatography to yield the desired white solid in 50.9% yield.

<u>Step 5:</u>



Boc-Ac₆c-OH were allowed to react with (R)-1-phenylethan-1-amine. The mixture was reacted for 48 hrs with the use of HATU as the coupling reagent. The workup was simple, and the pure product could be obtained by simply washing with acid and base. The yield was found to be 90.9% for Boc-Ac₆c-rpy.

4.2.8 Synthesis and complexation of L7



L7 was allowed to react with Eu(OTf)₃ in a ratio of 3 to 2. The solution turned to red luminescence which confirmed the complexation with Eu³⁺. However, the proton NMR signal was very weak (figure 4-10). Diamagnetic La complex of L7 was also prepared. However, unresolved broad peaks were observed in proton NMR and no conclusion can be drawn. ESI-HRMS was also performed for characterization of both Eu and La complexes. For Eu complex, undesired supramolecular Eu₂L7₃ complex was observed in the ESI-HRMS spectrum, and it was hypothesized that Eu₂L7₃ exhibited a helicate stucture. Due to the lack of X-ray crystal structure, confirmation of exact topology was not possible. Although L7 can form Eu₂L7₃ complex with Eu, La complex showed undesired La₂L7₂ lanthanide cluster. For Eu₂L7₃ complex, the stoichiometric ratio was confirmed by ESI-HRMS (figure 4-11). However, no structural information can be obtained from the NMR spectrum and further investigation is needed.



Figure 4-10. ¹HNMR of (a) free ligand **L7** in (CD₃)₂SO, (b) Eu complex of **L7** in CD₃CN and (c) La complex of **L7** in CD₃CN.



Figure 4-11. ESI-HRMS of (a) Eu complex of L7 (b) La complex of L7.

4.2.9 Synthesis and complexation of L8



Ligand with 2 Ac₆c units (**L8**) was allowed to react with La(OTf)₃ and Eu La(OTf)₃. Similar with the result of **L7**, red luminescence was also observed for Eu³⁺, indicating the successful coordination between **L8** and Eu³⁺. However, the proton NMR signal was very weak (figure 4-12). Similar result was obtained for La complex. ESI-HRMS result revealed both Eu and La complexes exhibited as Ln₂**L8**₂ lanthanide cluster in solution state no desired higher ordered supramolecular complex can be observed (figure 4-13).



Figure 4-12. ¹HNMR of (a) free ligand **L8** in (CD₃)₂SO, (b) Eu complex of **L8** in CD₃CN and (c) La complex of **L8** in CD₃CN.



Figure 4-13. ESI-HRMS of (a) Eu complex of L8 (b) La complex of L8.





Since **L5-8** cannot form desired supramolecular tetrahedron with lanthanide ions, it was hypothesized that the short peptide may create a large steric hindrance which prevent the formation of stable nine coordinated lanthanide centre. Therefore, **L9** was designed based on two pyridine-2,6-dicarboxamide (pcam) moieties, which tends to form stable nine-coordinated lanthanide complexes. Two pcam metal chelating pcam moieties are connected with a short Aib peptide to remove the bulkiness at the terminal site of the ligand. It is expected to form a stable bimetallic helicate since a monophenyl linker was used to connect two metal chelating units.

<u>Step 1:</u>

The first step involved the amide coupling of aniline with pyridine dicarboxylic acid. In the first trial, aniline was successfully coupled with pyridine dicarboxylic acid but the yield was low (30%). Possible reason may due to the fact that slightly excess amount of base (DIPEA) was added. More pyridine dicarboxylic acid was deprotected and react with the coupling reagent HATU. Therefore, more decoupling product was expected to be formed rather than the desired monocoupling product. However, in the second trial, the yield was low even limited amount of base was added. Possible reason may be due to the resonance structure of aniline, in which the nucleophilicity of the NH₂ group is stabilized by the resonance structure (figure 4-14). Therefore, the yield is low even if the reaction was performed under reflux



Figure 4-14. Resonance structure of aniline.

<u>Step 2:</u>



The second step involved the amide coupling of Boc-Aib-OH with the central bridging linker benzene-1,4-diamine using HATU as the coupling reagent. Pure product can be obtained easily by simply washing with MeCN. The highest yield was found to be 94%.

<u>Step 3:</u>



The third step involve the amide coupling Boc-Aib-OH with *Di-tert*-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.

EDCI/HOBt was used as the coupling reagent. Coupling α , α -disubstituted amino acids was difficult due to the steric hinderance and the highest yield was found to be 32.5%.

Step 4:



Following the previous step, desired *Di-tert*-butyl ((((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate can be prepared in 19% yield using EDCI/HOBt as the coupling reagent. Characterization by ¹³CNMR was not possible due to poor solubility.

4.2.11 Synthesis and complexation of L9



L9 was synthesized by in-situ deprotection of *di-tert*-butyl (((((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate followed by coupling with two equivalents of 6-(phenylcarbamoyl)picolinic acid. The yield was 41.0%.

Then **L9** was allowed to coordinate with Eu(OTf)₃ and La(OTf)₃ in a ratio of 3:2. The turbid ligand solution turned to be clear upon addition of lanthanide and the corresponding lanthanide complexes were characterized by NMR. The ¹HNMR was expected to be more complicated since no chiral group is introduced in the ligand and thus different isomers can be formed. Similar with **L8**, the ¹HNMR signal for both Eu and La complexes were very weak (figure 4-14). The ESI-HRMS showed a lot of fragments of Ln complexes and the major species was found to be mononuclear Ln**L9** for both Eu and La complexes (figure 4-15).



Figure 4-14. ¹*HNMR of (a) free ligand L9 in (CD₃)₂SO, (b) Eu complex of L9 in CD₃CN and (c) La complex*

of **L9** in CD₃CN.



Figure 4-15. ESI-HRMS of (a) Eu complex of L9 (b) La complex of L9. 186

4.2.12 Synthesis of intermediates for L10



Another ligand **L10** was synthesis for preventing the formation of intramolecular hydrogen bond. Intramolecular hydrogen bond can be formed between the carbonyl oxygen and amide NH group. Aib amino acid unit was found to form stable 3₁₀ helix, which means a helix is formed by 3 amino acid unit consisting of 10 atoms. The carbonyl oxygen in the pcam chelating moiety can be also participate in the helical formation by forming intramolecular hydrogen bond with the Aib peptide. Thus, the coordination power of the pcam moiety will be weakened. Moreover, the flexibility of the chelating group will be locked because of the intramolecular hydrogen bond of the peptide, which hindered the fine tuning of the whole supramolecular complex.

In **L10**, two pcam metal chelating moieties are first connected with a phenyl ring, which aims at separating the pcam chelating moiety with the peptide. It is

expected that the formation of intramolecular hydrogen bond between the pcam unit and the peptide will be avoided. Chiral amino acid unit Leu will be coupled into the peptide to introduced point chirality. Peptide chain Aib-Leu-Aib is expected to form a stable 3₁₀ helix through intramolecular hydrogen bond which can transmit the chiral information from Leu to the metal chelating unit.

Following the previous protocol of synthesizing L9, all the intermediates for L10 can be prepared in 4 steps and the synthetic route was shown in figure 4-16.



Figure 4-16. Synthetic route for preparation of L10 intermediates.

4.2.13 Synthesis and complexation of L10

L10 was synthesized by in-situ deprotection of di-tert-butyl ((((2S,2'S)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-

methyl-1-oxopentane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))dicarbamate followed by coupling with two equivalents of 4-(6-(isopropylcarbamoyl)picolinamido)benzoic acid. The yield was found to be 26.8%.



Upon coordination of **L10** with Eu(OTf)₃ in a ratio of 3:2. The ¹HNMR of the corresponding Eu complex showed no signal. For La complex, very weak signal was observed (figure 4-17). In the ESI-HRMS spectrum, the major species for Eu complex was found to be Eu₂L10₂ and EuL10₂. For La complex, the major species was found to be La₂L10₂. No desired supramolecular tetrahedron can be observed (figure 4-18).



Figure 4-17. ¹HNMR of (a) free ligand **L10** in (CD₃)₂SO, (b) Eu complex of **L10** in CD₃CN and (c) La

complex of **L10** in CD₃CN.



Figure 4-18. ESI-HRMS of (a) Eu complex of L10 (b) La complex of L10.

4.2.14 Review of ligand L5-10



Figure 4-19. Structure and complexation result of peptide ligands L5-L10.

Figure 4-19 showed the peptide ligands that have synthesized. The original ligand design principle depends on two factors, which were the choice of amino acid and the length of the peptide units. It was believed that the peptide sequence will not form helicate when the length is short. As a result, no intramolecular hydrogen bond between the carbonyl oxygen and amide NH group will be formed and the coordination site will not be affected. However, based on the result, we believed that the presence of two amide group will strongly affect the coordination. Although L7 can form Eu₂L7₃ complex, the structure cannot be confirmed by ¹HNMR. For L10, it was proposed that the coordination site will not be affected from the chelating group by a phenyl group. However, no supramolecular complexes can be observed upon coordination with lanthanide ions.

4.3 Conclusion and future perspectives

Various peptide ligand derivatives were synthesized in order to study the supramolecular formation. Although lanthanide ions can successfully coordinate with the metal chelating moiety, major product was found to be M₂L₂ and ML₂. Result showed that the steric bulkiness and the choice of the amino acid unit does not have an effect towards the coordination with lanthanide ion. Moreover, the presence of two amide bonds may hinder the formation of lanthanide supramolecular complexes.

One of the possible solutions is to change the metal chelating unit. Although pcam was found to be well known for coordinating with lanthanide ions to form ninecoordinated complexes, the complexes were unstable due to the neutral nature of the chelating group. It is believed that using other charged chelating group or macrocyclic ligand can help to improve the coordinating power.

4.4 Experimental

Methyl 2-((tert-butoxycarbonyl)amino)-2-methylpropanoate, Boc-Aib-OMe



lodomethane (1 ml, 12.3 mmol, 5 eq.) was added to a pressure vessel containing Boc-Aib-OH (0.5 g, 2.46 mmol, 1 eq.) and potassium carbonate (0.68 g, 4.92 mmol, 2 eq.). The reaction mixture was allowed to stir for 16 hrs. Dichloromethane (15 ml) was then added and the insoluble white solid was filtered. The crude product was purified by column chromatography (DCM/MeOH, 40:1, v/v) to give the desired white solid. **Boc-Aib-OMe**: (0.347g, 1.60 mmol, 64.9% yield), ¹H NMR (400 MHz, CDCl₃, 298K, δ): 1.43 (s, 9H), 1.49 (s, 6H), 3.73 (s, 3H), 4.99 (s, 1H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 175.26, 154.56, 56.10, 52.32, 28.25, 23.35. HRMS (ESI) calcd for C₁₀H₁₉NO₄Na [M+Na]⁺:240.26, found 240.12.

1-methoxy-2-methyl-1-oxopropan-2-aminium 2,2,2-trifluoroacetate, H₃N⁺-Aib-OMe



Boc-Aib-OMe (0.15 g, 0.69 mmol) was added to a mixture of DCM/TFA (4:1, v/v). The solution was allowed to stir for 2 hours. The solvent was then evaporated, and diethyl ether was added to precipitate out the desired white solid. H_3N^+ -Aib-OMe: (0.0279g,

0.0435 mmol, 71.7% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.25 (d, J = 7.0 Hz, 3H), 7.02 - 7.41 (m, 5H), 7.84 (s, 2H), 8.09 - 8.38 (m, 3H), 9.46 (t, J = 6.1 Hz, 1H), 10.77 (s, 1H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 172.45, 56.28, 53.61, 23.87. HRMS (ESI) calcd for C₅H₁₂NO₂ [M+H]⁺:118.15, found 118.10.

Methyl 2-(2-((*tert*-butoxycarbonyl)amino)-2-methylpropanamido)-2methylpropanoate, Boc-Aib-Aib-OMe



To a stirred solution of Boc-Aib-OH (2 g, 9.84 mmol, 1.0 eq) in anhydrous DCM (30 ml), DIPEA (5.14 ml, 29.52 mmol, 3.0 eq) was added. The solution was stirred for 1 hr. The PyBOP (5.12 g, 9.84 mmol, 1.0 eq) was added. Then the H₃N⁺-Aib-OMe (2.5 g, 10.82 mmol, 1.1 eq) was added and the mixture was reacted for 16 hrs at rtp. The DCM was evaporated and EA (150 ml) was added. The organic layer was washed by 5% HCl (3 x 50 ml), 5% NaHCO3 (3 x 50 ml), brine (3 x 50 ml) and then dried with anhydrous MgSO4. The crude oil was then purified by column chromatography (hex/EA: 1/1) to obtain a white solid. **Boc-Aib-Aib-OMe**: (2.23 g, 7.38 mmol, 75.0% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.14-1.43 (m, 21H), 3.52 (s, 1H), 6.71 (s, 1H), 7.58 (s, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 175.01, 174.44, 154.61, 55.94, 55.52, 52.23, 28.57, 25.17. HRMS (ESI) calcd for C₁₄H₂₆N₂O₅ [M+Na]⁺:325.37, found 325.13.

2-(2-((tert-butoxycarbonyl)amino)-2-methylpropanamido)-2-methylpropanoic acid, Boc-Aib-Aib-OH



To a stirred solution of Boc-Aib-Aib-OMe (1 g, 3.31 mmol, 1.0 eq) in THF (26 ml), lithium hydroxide monohydrate (0.42 g, 9.92 mmol, 3.0 eq) in water (13.2 ml) was added. The reaction was stirred for 16 hrs at rtp. The THF was evaporated and the water phase was concentrated to 8 ml. 1% HCl was added dropwise to acidify the water phase to pH 3. Then DCM (5 x 30 ml) was added and the organic phase was collected, dried to obtain the desired white solid. **Boc-Aib-Aib-OH**: (0.86 g, 2.99 mmol, 90.5%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.26 (s, 6H), 1.35 (s, 15H), 6.85 (s, 1H), 7.43 (s, 1H). HRMS (ESI) calcd. for C₁₃H₂₄N₂O₅ [M-H]⁻:287.34, found 287.60. ¹³C NMR (125 MHz, (CD₃)₂SO, 298K, δ): 176.45, 174.23, 154.73, 56.23, 55.55, 28.64, 25.42, 24.87.

Methyl 2,2,6,6,9,9,12,12-octamethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13oate, Boc-Aib-Aib-Aib-OMe

To a stirred solution of Boc-Aib-Aib-OH (0.2 g, 0.69 mmol, 1.0 eq) in anhydrous DMF (3 ml), DIPEA (0.26ml, 1.53 mmol, 2.2 eq) was added. The solution was stirred for 1 hr. HATU (0.26 g, 0.69 mmol, 1.0 eq) was then added. Then the H₃N⁺-Aib-OMe (0.16 g, 0.69 mmol, 1.0 eq) was added and the mixture was reacted for 12 hrs at rtp. DCM was evaporated and EA (30 ml) was added. The organic layer was washed with 1 M LiCl solution (10 ml), 5% HCl (3 x 10 ml), brine (10 ml), 0.5M K₂CO₃ (3 x 10 ml), brine (10

ml) and then dried with anhydrous MgSO₄. The crude oil was then purified by column chromatography (hex/EA: 1/1) to obtain a white solid. **Boc-Aib-Aib-Aib-OMe**: (17.3 mg, 0.047 mmol, 6.4% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.11-1.51 (m, 27H), 3.51 (s, 3H), 7.20 (s, 1H), 7.57 (s, 2H); ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 174.88, 174.31, 173.66, 155.62, 79.14, 56.28, 55.87, 55.36, 52.10, 28.57, 25.28, 25.07. HRMS (ESI) calcd. for C₁₈H₃₃N₃O₆ [M+Na]⁺: 410.48, found 410.17.

Methyl 2,2,6,6,9,9,12,12,15,15-decamethyl-4,7,10,13-tetraoxo-3-oxa-5,8,11,14tetraazahexadecan-16-oate, Boc-(Aib)₄-OMe



Boc-Aib-Aib-Aib-OMe (1 g, 2.68 mmol, 1.0 eq.) was dissolved in 10 ml of DCM. 10 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 3 hrs. The solvent was then evaporated to obtain a viscous oil. The residue was dissolved in 10 ml of DMF and to this solution is added Boc-Aib-OH (0.54 g, 2.68 mmol, 1.0 eq.), HOBt (0.36 g, 2.68 mmol, 1.0 eq.), EDC (0.51 g, 2.68 mmol, 1.0 eq.) and DIPEA (1.40 ml, 8.03 mmol, 3.0 eq.). The reaction was stirred at 40°C for 48 hrs. The solvent was removed and the residue was dissolved in ethyl acetate (100 ml). The organic layer was washed with 1 M LiCl solution (20 ml),5% HCl (3 x 20 ml), brine (20 ml), 0.5M K₂CO₃ (3 x 20 ml), brine (20 ml) and then dried with anhydrous MgSO₄. The solvent was then removed to obtain a yellowish gummy solid. The gummy solid was then washed with diethyl ether (3 x 10 ml) to obtain a white solid. **Boc-(Aib)**₄-**OMe**: (0.69 g, 1.45 mmol, 54.3%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.18-1.52 (m, 33H), 3.53 (s, 1H), 7.53 (d, J = 5.3 Hz, 2H), 8.15 (s, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO,

298K, δ): 174.88, 174.31, 173.65, 155.61, 79.14, 56.28, 55.86, 55.35, 52.10, 28.57, 25.29, 25.07. HRMS (ESI) calcd for C₂₂H₄₁N₄O₇ [M+H]⁺: 473.2970, found 473.2981.

2,2,6,6,9,9,12,12,15,15-decamethyl-4,7,10,13-tetraoxo-3-oxa-5,8,11,14tetraazahexadecan-16-oic acid, Boc-(Aib)₄-OH



To a stirred solution of Boc-(Aib)₄-OMe (0.3 g, 0.64 mmol, 1.0 eq) in THF (4 ml), lithium hydroxide monohydrate (0.080 g, 1.90 mmol, 3.0 eq) in minimum amount of water was added. The reaction was stirred for 16 hrs at rtp. The THF was evaporated. Water (10 ml) was then added. 1% HCl was added dropwise to acidify the water phase to pH 2. Then DCM (5 x 10 ml) was added and the organic phase was collected, dried to obtain the desired white solid. **Boc-(Aib)**₄-OH: (0.25 g, 0.54 mmol, 85.1%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.29 (dd, J = 19.7, 8.3 Hz, 24H), 1.42 (s, 9H), 7.34 (s, 1H), 7.44 (s, 1H), 7.53 (s, 1H), 8.11 (s, 1H), 11.83 (s, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 176.07, 175.50, 173.95, 173.77, 155.80, 79.41, 56.39, 56.17, 55.06, 28.57, 25.64, 25.14. HRMS (ESI) calcd for C₂₁H₃₇N₄O₇ [M-H]⁻: 457.2668, found 457.2674.

tert-butyl (S)-(2-methyl-1-((2-methyl-1-oxo-1-((1-phenylethyl)amino)propan-2yl)amino)-1-oxopropan-2-yl)carbamate, Boc-Aib-Aib-spy

To a stirred solution of Boc-Aib-Aib-OH (5 g, 17.34 mmol, 1.0 eq) in anhydrous DMF (75 ml), DIPEA (6.63 ml, 52.02 mmol, 3.0 eq) was added. The solution was stirred for 196

20 mins. HATU (7.25 g, 19.07 mmol, 1.1 eq) was then added. Then the (S)-1phenylethan-1-amine (2.21 ml, 17.34 mmol, 1.0 eq) was added and the mixture was reacted for 3 days at rtp. The reaction mixture was diluted with ethyl acetate (200 ml). The organic layer was washed by 0.5 M LiCl (3 x 50 ml),5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 50 ml) and then dried with anhydrous MgSO₄. to obtain the desired pure white solid. **Boc-Aib-Aib-spy**: (6.27 g, 16.0 mmol, 92.3%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 7.80 (d, *J* = 7.9 Hz, 1H), 7.58 (s, 1H), 7.23 (ddd, *J* = 33.1, 17.7, 7.3 Hz, 6H), 4.83 (p, *J* = 7.2 Hz, 1H), 1.52 – 1.11 (m, 24H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 173.93, 155.69, 145.44, 128.41, 126.71, 126.37, 79.19, 56.37, 56.20, 48.61, 28.56, 25.40, 23.06. HRMS (ESI) calcd for C₂₁H₃₃N₃O₄Na [M+Na]⁺:414.2363, found 414.2375.

Tert-butyl (S)-(5,5,8,8,11,11,14-heptamethyl-4,7,10,13-tetraoxo-2-phenyl-3,6,9,12tetraazapentadecan-14-yl)carbamate, Boc-(Aib)₄-spy



To a stirred solution of Boc-(Aib)₄-OH (0.1 g, 0.22 mmol, 1.0 eq) in anhydrous DMF (2 ml), DIPEA (0.11 ml, 0.085 mmol, 3.0 eq) was added. The solution was stirred for 1 hr. HATU (0.083 g, 0.22 mmol, 1.0 eq) was then added. Then the (S)-1-phenylethan-1amine (0.028 ml 0.22 mmol, 1.0 eq) was added and the mixture was reacted for 12 hrs at rtp. DCM was evaporated and EA (30 ml) was added. The organic layer was washed with 1 M LiCl solution (10 ml),5% HCl (3 x 10 ml), brine (10 ml), 0.5M K₂CO₃ (3 x 10 ml), brine (10 ml) and then dried with anhydrous MgSO₄. to obtain a white solid. **Boc-**(Aib)₄-spy: (24.7 mg, 0.044 mmol, 20.2% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.19-1.55 (m, 33H, 4.86 (t, J = 7.3 Hz, 1H), 7.09-7.44 (m, 6H), 7.57 (s, 1H), 7.72 (d, J = 8.2 Hz, 1H), 7.91 (s, 1H), 8.27 (s, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 175.73, 173.82, 155.95, 145.77, 128.36, 126.62, 126.35, 79.52, 56.60, 56.55, 56.42, 56.18, 48.56, 28.59, 25.83, 25.52, 25.04, 23.58. HRMS (ESI) calcd. for C₂₉H₄₇N₅O₆Na [M+Na]⁺: 584.3419, found 584.3429.

Di-tert-butyl 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6diyl)bis(azanediyl))bis(carbonyl))dipicolinate



To a stirred solution of 6-(tert-butoxycarbonyl)picolinic acid (0.3 g, 1.34 mmol, 3.0 eq.) in DMF (5ml), DIPEA (0.97 ml, 5.60 mmol, 12.5 eq.) and HATU (1.02 g, 2.69 mmol, 6.0 eq.) was added. The solution was allowed to stir for 20 mins before the addition of 2,6-diaminoanthracene-9,10-dione (0.11g, 0.45 mmol, 1.0 eq.). The reaction mixture was allowed to stir at room temperature and pressure for 16 hrs. The mixture was washed with water (3 x 10 ml) and MeCN (3 x 10 ml) to obtain the desired yellow product. **Di-tert-butyl 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6diyl)bis(azanediyl))bis(carbonyl))dipicolinate**: (0.27 g, 0.45 mmol, 93.5%), ¹H NMR (400 MHz, CDCl₃, 298K, δ): 1.73 (s, 9H), 8.10 (t, *J* = 7.8 Hz, 1H), 8.27 (d, *J* = 7.6 Hz, 1H), 8.41 (d, *J* = 8.5 Hz, 1H), 8.46 – 8.58 (m, 3H), 10.67 (s, 1H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 181.80, 163.11, 161.92, 149.34, 147.94, 143.17, 138.93, 134.98, 129.60, 129.23, 127.65, 125.42, 124.30, 117.42, 83.22, 29.71, 28.13, 1.02, -0.01. HRMS (ESI) calcd. for C₃₆H₃₂N₄O₈ [M+Na]⁺: 671.67, found 671.00.

6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-

diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid



TFA (5 ml) was added to a solution containing di-tert-butyl 6,6'-(((9,10-dioxo-9,10dihydroanthracene-2,6-diyl)bis(azanediyl))bis(carbonyl))dipicolinate (0.2 g, 0.31 mmol) in DCM (5 ml). The mixture was allowed to stir for 2 hrs. The solvent was then evaporated and washed with diethyl ether (3 x 10 ml) to obtain the desired yellow product. **6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid**: (0.15 g, 0.27 mmol, 89.0%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 8.23 – 8.57 (m, 6H), 8.71 (d, *J* = 2.2 Hz, 1H), 11.36 (s, 1H). N², N²'-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-((S)-5,5,8,8,11,11,14heptamethyl-4,7,10,13-tetraoxo-2-phenyl-3,6,9,12-tetraazapentadecan-14yl)pyridine-2,6-dicarboxamide), L5



Tert-butyl (S)-(5,5,8,8,11,11,14-heptamethyl-4,7,10,13-tetraoxo-2-phenyl-3,6,9,12 tetraazapentadecan-14-yl)carbamate (0.31 g, 0.56 mmol, 3.0 eq.) was dissolved in 4 ml of DCM. 2 ml of trifluoroacetic acid was added and the reaction was maintained in

room temperature for 3 hrs. The solvent was then evaporated to obtain a viscous oil. The residue was dissolved in 6 ml of DMF and to this solution is added 6,6'-(((9,10 dioxo-9,10-dihydroanthracene-2,6-diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid (0.10 g, 0.19 mmol, 1.0 eq.), HATU (0.43 g, 1.12 mmol, 6.0 eq.) and DIPEA (0.41 ml, 2.33 mmol, 12.5 eq.). The reaction was stirred for 16 hrs at room temperature and pressure. The solvent was then washed with water (2 x 20 ml) and MeCN (2 x 10 ml) to obtain the desired yellow product. **L5**: (0.13 g, 0.09 mmol, 49.6%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.28 – 1.42(m, 15H), 1.49 (d, *J* = 9.2 Hz, 6H), 1.66 (d, *J* = 3.1 Hz, 6H), 4.86 (q, *J* = 7.4 Hz, 1H), 7.21 (dt, *J* = 31.4, 7.2 Hz, 3H), 7.39 (d, *J* = 7.5 Hz, 2H), 7.66 – 7.91 (m, 2H), 8.27 – 8.43 (m, 4H), 8.43 – 8.61 (m, 2H), 8.88 (d, *J* = 2.1 Hz, 1H), 9.53 (s, 1H), 11.53 (s, 1H). ¹³C NMR (125 MHz, (CD₃)₂SO, 298K, δ): 182.05, 175.73, 174.94, 174.20, 173.89, 163.96, 163.13, 149.15, 148.82, 145.89, 144.24, 134.90, 129.50, 128.54, 126.47, 57.07, 56.76, 56.72, 56.57, 25.19, 23.75. HRMS (ESI) calcd for C₇₆H₉₀N₁₄O₁₄Na [M+Na]⁺: 1445.6653, found 1445.6676.

N²,N²'-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-(2-methyl-1-((2-methyl-1-oxo-1-(((S)-1-phenylethyl)amino)propan-2-yl)amino)-1-oxopropan-2-yl)pyridine-2,6-dicarboxamide), L6



Tert-butyl (S)-(2-methyl-1-((2-methyl-1-oxo-1-((1-phenylethyl)amino)propan-2yl)amino)-1-oxopropan-2-yl)carbamate (0.11 g, 0.28 mmol, 3.0 eq.) was dissolved in 2 ml of DCM. 1 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 3 hrs. The solvent was then evaporated to obtain a viscous oil. The residue was dissolved in 5 ml of DMF and to this solution is added 6,6'-(((9,10 dioxo-9,10-dihydroanthracene-2,6-diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid (50.0 mg, 0.093 mmol, 1.0 eq.), HATU (0.21 g, 0.56 mmol, 6.0 eq.) and DIPEA (0.20 ml, 1.17 mmol, 12.5 eq.). The reaction was stirred for 16 hrs at room temperature and pressure. The solvent was then washed with water (2 x 10 ml) and MeCN (2 x 10 ml) to obtain the desired yellow product. L6: (54.10 mg, 0.05 mmol, 53.6%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.34 (d, J = 16.7 Hz, 6H), 1.46 (d, J = 7.0 Hz, 3H), 1.64 (d, J = 3.7 Hz, 6H), 4.90 (p, J = 7.1 Hz, 1H), 7.19 (t, J = 7.1 Hz, 1H), 7.28 (t, J = 7.5 Hz, 2H), 7.39 (d, J = 7.3 Hz, 2H), 7.77 (s, 1H), 7.88 (d, J = 8.1 Hz, 1H), 8.22 (dd, J = 7.5 Hz, 2H), 8.30 (t, J = 7.7 Hz, 1H), 8.36 (d, J = 8.36 Hz, 1H), 8.40 – 8.53 (m, 2H), 8.89 (d, J = 2.2 Hz, 1H), 9.44 (s, 1H), 11.52 (s, 1H). ¹³C NMR (125 MHz, (CD₃)₂SO, 298K, δ): 182.02, 173.93, 173.28, 163.74, 163.15, 149.24, 148.81, 145.64, 144.26, 140.48, 134.87, 129.45, 129.10, 128.55, 126.81, 126.54, 126.12, 125.85, 125.67, 118.07, 57.29, 56.76, 48.72, 25.75, 25.57, 25.40, 23.34. HRMS (ESI) calcd for C₆₀H₆₂N₁₀O₁₀Na [M+Na]⁺: 1105.4543, found 1105.4554.

Methyl 1-aminocyclohexane-1-carboxylate, H₂N- Ac₆c-OMe



To a stirred solution of 1-aminocyclohexane-1-carboxylic acid (5 g, 34.92 mmol, 1.0 eq.) in methanol (50 ml), trimethylchlorosilane (8.86 ml, 69.84 mmol, 2.0 eq.) was added dropwise at 0°C. The mixture was allowed to react at 50°C for 16 hrs. The
solvent was evaporated and redissolved in water (50 ml). The crude product was washed with dichloromethane (3 x 20 ml) and NaHCO₃ was added to the aqueous layer with pH ~ 8. Dichloromethane (3 x 50 ml) was added and the organic layer was concentrated on a rotary evaporator to give the desired product as a yellow oil. H₂N-Ac₆c-OMe: (2.83 g, 18.00 mmol, 51.5%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.21 – 1.40 (m, 6H), 1.60 (ddd, *J* = 11.6, 9.1 6.1 Hz, 2H), 1.68 – 1.81 (m, 4H), 3.59 (s, 3H). ¹³C NMR (101 MHz, MeOD, 298K, δ): 176.80, 57.04, 51.16, 34.96, 24.99, 21.71. HRMS (ESI) calcd for C₈H₁₆NO₂ [M+H]⁺: 158.1176, found 158.1181.

1-((tert-butoxycarbonyl)amino)cyclohexane-1-carboxylic acid, Boc-Ac₆c-OH



To a stirred solution of 1-aminocyclohexane-1-carboxylic acid (10.52 g, 73.47 mmol, 1.0 eq.) in 2 N NaOH (110 ml), Boc₂O (19.24 g, 88.17 mmol, 1.2 eq.) in dioxane (110 ml) was added dropwise. The reaction was stirred for 16 hrs at rtp. The dioxane was then evaporated and water (200 ml) was added. The aqueous phase was washed with diethyl ether (3 x 60 ml) and then acidified to pH ~ 4 by 5% HCl. The product was then collected by filtration and washed with brine (3 x 60 ml) to obtain the desired white product. **Boc-Ac₆c-OH**: (8.77 g, 36.06 mmol, 49.1%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.22 (d, *J* = 10.9 Hz, 2H), 1.28 – 1.53 (m, 14H), 1.61 (dt, *J* = 14.1, 7.4 Hz, 2H), 1.90 (s, 2H), 6.87 (s, 1H), 12.09 (s, 1H). ¹³C NMR (101 MHz, MeOD, 298K, δ): 177.41, 155.95, 32.22, 27.39, 26.20, 25.14, 21.07. HRMS (ESI) calcd for C₁₂H₂₀NO₄ [M-H]⁻: 242.1398, found 242.1400.

1-(1-((tert-butoxycarbonyl)amino)cyclohexane-1-carboxamido)cyclohexane-1-

carboxylic acid, Boc-Ac₆c-Ac₆c-OH



To a stirred solution of Boc-Ac₆c-OH (5 g, 20.55 mmol, 1.0 eq.) in anhydrous DMF (50 ml), DIPEA (10.74 ml, 61.65 mmol, 3.0 eq.) was added. The solution was stirred for 20 mins. The PyBOP (11.76 g, 22.61 mmol, 1.1 eq.) was added. Then the H-Ac₆c-OMe (3.23 g, 20.55 mmol, 1.0 eq.) was added and the mixture was reacted for 16 hrs at rtp. The reaction mixture was diluted with ethyl acetate (150 ml). The organic layer was washed by 0.5 M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 50 ml) and then dried with anhydrous MgSO₄. The organic layer was concentrated. The residue was redissoved in THF (100 ml), lithium hydroxide monohydrate (2.59 g, 61.65 mmol, 3.0 eq.) in water (100 ml) was added. The reaction was stirred for 2 days at rtp. The solvent was evaporated. 5% HCl was added dropwise to acidify the aqueous phase to pH 4. Then DCM (5 x 50 ml) was added and the organic phase was collected, dried to obtain the desired white solid. Boc-Ac₆c-Ac₆c-OH: (5.53g, 15.00 mmol, 73.0%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.17 (d, J = 13.4 Hz, 2H), 1.41 (d, J = 25.2 Hz, 23H), 1.97 (d, J = 13.1Hz, 4H), 6.64 (s, 1H), 7.11 (s, 1H), 11.98 (s, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 21.14, 21.35, 25.40, 25.49, 28.58, 31.75, 31.98, 57.79, 58.87, 78.57, 155.04, 174.62, 176.04. HRMS (ESI) calcd for $C_{19}H_{31}N_2O_5$ [M-H]⁻: 367.2238, found 367.2285.

tert-butyl (S)-(1-((1-phenylethyl)carbamoyl)cyclohexyl)carbamate, Boc-Ac₆c-rpy



To a stirred solution of Boc-Ac₆c-OH (1 g, 4.11 mmol, 1.0 eq) in anhydrous DMF (40 ml), DIPEA (2.15 ml, 12.33 mmol, 3.0 eq) was added. The solution was stirred for 20 mins. HATU (1.72 g, 4.52 mmol, 1.1 eq) was then added. Then the (R)-1-phenylethan-1-amine (0.52 ml, 4.11 mmol, 1.0 eq) was added and the mixture was reacted for 3 days at rtp. The reaction mixture was diluted with ethyl acetate (100 ml). The organic layer was washed by 0.5 M LiCl (3 x 30 ml),5% HCl (3 x 30 ml), 0.5 M K₂CO₃ (3 x 30 ml), brine (3 x 30 ml) and then dried with anhydrous MgSO₄. The crude product was then purified by column chromatography (DCM/EtOH, 100/1, v/v) to obtain the desired pure white solid. **Boc-Ac₆c-rpy**: (0.13 g, 0.28 mmol, 50.9%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 7.59 (d, *J* = 8.1 Hz, 1H), 7.29 (d, *J* = 5.8 Hz, 3H), 7.27 – 7.14 (m, 1H), 6.51 (s, 1H), 4.89 (p, *J* = 7.1 Hz, 1H), 1.95 (s, 2H), 1.73 – 1.07 (m, 19H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 174.19, 154.83, 145.08, 128.52, 126.90, 126.33, 58.87, 48.05, 28.64, 25.51, 22.74, 21.48. HRMS (ESI) calcd for C₂₀H₃₀N₂O₃Na [M+Na]⁺: 369.2149, found 369.2154.

Tert-butyl

(S)-(1-((1-((1-

phenylethyl)carbamoyl)cyclohexyl)carbamoyl)cyclohexyl)carbamate, Boc-Ac₆c-Ac₆c-rpy

To a stirred solution of Boc-Ac₆c-Ac₆c-OH (0.2 g, 0.54 mmol, 1.0 eq) in anhydrous DMF (5 ml), DIPEA (0.28 ml, 1.63 mmol, 3.0 eq) was added. The solution was stirred for 20 mins. HATU (0.25 g, 0.65 mmol, 1.2 eq) was then added. Then the (S)-1-phenylethan-1-amine (0.069 ml 0.54 mmol, 1.0 eq) was added and the mixture was reacted for 3 days at rtp. The reaction mixture was diluted with ethyl acetate (10 ml). The organic layer was washed by 0.5 M LiCl (3 x 10 ml),5% HCl (3 x 10 ml), 0.5 M K₂CO₃ (3 x 10 ml), brine (3 x 10 ml) and then dried with anhydrous MgSO₄. The crude product was then purified by column chromatography (DCM/MeOH, 20/1, v/v) to obtain the desired pure white solid. **Boc-Ac₆c-Ac₆c-rpy**: (0.13 g, 0.28 mmol, 50.9%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 1.04-1.80 (m, 28H), 1.95 (d, *J* = 14.1 Hz, 4H), 4.85 (p, *J* = 7.3 Hz, 1H), 6.79 (s, 1H), 7.03 (s, 1H), 7.11-7.38 (m, 4H), 7.50 (d, *J* = 7.9 Hz, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 174.37, 173.58, 155.70, 145.16, 128.45, 126.78, 126.32, 79.35, 59.37, 59.10, 48.46, 31.93, 28.58, 25.41, 22.98, 21.34, 21.21. HRMS (ESI) calcd for C₂₇H₄₁N₃O₄Na [M+Na]⁺: 494.2989, found 494.3004.

N², N²'-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-(1-(((R)-1-phenylethyl)carbamoyl)cyclohexyl)pyridine-2,6-dicarboxamide), L7



Tert-butyl (R)-(1-((1-phenylethyl)carbamoyl)cyclohexyl)carbamate (1 g, 2.89 mmol, 3.0 eq.) was dissolved in 20 ml of DCM. 5 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 3 hrs. The solvent was then evaporated to obtain a viscous oil. The residue was dissolved in 10 ml of DMF and to

this solution added 6,6'-(((9,10 dioxo-9,10-dihydroanthracene-2,6is diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid (0.52 g, 0.96 mmol, 1.0 eq.), HATU (0.80 g, 2.12 mmol, 2.2 eq.) and DIPEA (1.17 ml, 6.73 mmol, 7.0 eq.). The reaction was stirred for 16 hrs at room temperature and pressure. The reaction mixture was diluted with ethyl acetate (30 ml). The organic layer was washed by 0.5 M LiCl (3 x 10 ml), 5% HCl ($3 \times 10 \text{ ml}$), 0.5 M K₂CO₃ ($3 \times 10 \text{ ml}$), brine ($3 \times 10 \text{ ml}$) and then dried with anhydrous MgSO₄. The organic layer was concentrated and then purified by HPLC with Sunfire Silica column (DCM/MeOH: 95/5; flow rate 8 ml/min). L7: (0.18 g, 0.18 mmol, 18.8%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 11.53 (s, 1H), 8.96 (s, 1H), 8.84 (d, J = 2.2 Hz, 1H), 8.42 (ddd, J = 7.3, 5.8, 1.9 Hz, 2H), 8.39 – 8.26 (m, 3H), 7.98 (d, J = 8.2 Hz, 1H), 7.35 (d, J = 7.6 Hz, 2H), 7.29 (t, J = 7.5 Hz, 2H), 7.20 (t, J = 7.1 Hz, 1H), 5.02 (q, J = 7.2 Hz, 1H), 2.44 (d, J = 16.3 Hz, 1H), 2.36 (d, J = 14.4 Hz, 1H), 1.96 (t, J = 12.0 Hz, 2H), 1.62 (d, J = 20.9 Hz, 5H), 1.38 (d, J = 7.0 Hz, 4H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 181.78, 173.16, 163.20, 149.87, 148.62, 145.14, 144.24, 140.24, 134.81, 129.26, 128.98, 128.51, 126.84, 125.70, 125.06, 117.63, 60.35, 48.29, 32.41, 31.88, 25.53, 22.57, 21.79. HRMS (ESI) calcd for C₅₈H₅₆N₈O₈Na [M+Na]⁺: 1015.4113, found 1015.4127.

N²,N^{2'}-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N6-(1-((1-(((R)-1-phenylethyl)carbamoyl)cyclohexyl)carbamoyl)cyclohexyl)pyridine-2,6-dicarboxamide), L8

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phenylethyl)carbamoyl)cyclohexyl)carbamoyl)cyclohexyl)carbamatete (1 g, 2.12 mmol, 3.0 eq.) was dissolved in 20 ml of DCM. 5 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 3 hrs. The solvent was then evaporated to obtain a viscous oil. The residue was dissolved in 10 ml of DMF and to this solution added 6,6'-(((9,10 dioxo-9,10-dihydroanthracene-2,6is diyl)bis(azanediyl))bis(carbonyl))dipicolinic acid (0.38 g, 0.71 mmol, 1.0 eq.), HATU (0.59 g, 2.1.55 mmol, 2.2 eq.) and DIPEA (0.86 ml, 4.95 mmol, 7.0 eq.). The reaction was stirred for 16 hrs at room temperature and pressure. The reaction mixture was diluted with ethyl acetate (30 ml). The organic layer was washed by 0.5 M LiCl (3 x 10 ml), 5% HCl (3 x 10 ml), 0.5 M K_2CO_3 (3 x 10 ml), brine (3 x 10 ml) and then dried with anhydrous MgSO₄. The organic layer was concentrated and then purified by HPLC with Sunfire Silica column (DCM/MeOH: 95/5; flow rate 8 ml/min). L8: (0.22 g, 0.18 mmol, 25.0%), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 11.62 (s, 1H), 9.36 (s, 1H), 8.87 (s, 1H), 8.48 (d, J = 8.7 Hz, 1H), 8.42 (d, J = 7.7 Hz, 1H), 8.36 (d, J = 8.5 Hz, 1H), 8.27 (t, J = 7.8 Hz, 1H), 8.15 (d, J = 7.7 Hz, 1H), 7.82 (d, J = 8.2 Hz, 1H), 7.36 (d, J = 7.6 Hz, 2H), 7.32 -7.21 (m, 3H), 7.17 (t, J = 7.2 Hz, 1H), 4.93 (q, J = 7.3 Hz, 1H), 2.38 – 2.12 (m, 4H), 2.03 (s, 2H), 1.80 – 1.10 (m, 17H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 181.64, 173.69, 164.01, 163.01, 149.21, 148.71, 145.48, 144.19, 140.11, 134.67, 129.17, 128.89, 128.44, 126.72, 126.41, 125.78, 124.91, 117.68, 60.18, 48.45, 33.01, 32.43, 31.71, 25.58, 23.22, 22.04, 21.58. HRMS (ESI) calcd for C₇₂H₇₈N₁₀O₁₀Na [M+Na]⁺: 1265.5795, found 1265.5807.

((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-

Di-tert-butyl

diyl))dicarbamate

To a stirred solution of 2-((*tert*-butoxycarbonyl)amino)-2-methylpropanoic acid (3 g, 14.76 mmol, 2.2 eq) in DCM (10 ml), DIPEA (7.02 ml, 40.26 mmol, 6 eq) was added and the solution was allowed to stirred for 10 mins. HATU (5.61 g, 14.76 mmol, 2.2 eq) was then added and the solution was allowed to stir for another 10 mins. Benzene-1,4-diamine (0.73 g, 6.71 mmol, 1 eq) was added and the reaction mixture was allowed to stir for 48 hrs at rtp. Solvent was then evaporated and the crude product was washed with water (3 x 50 ml) and MeCN (3 x 50 ml) to obtained the desired white product.

Di-tert-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))dicarbamate: (3.02 g, 6.31 mmol, 94.0% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 9.29 (s, 2H), 7.48 (s, 4H), 6.88 (s, 2H), 1.37 (s, 15H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 173.49, 154.70, 135.10, 120.50, 78.61, 56.81, 28.57, 25.49. HRMS (ESI) calcd for C₂₄H₃₈N₄O₆Na [M+Na]⁺: 501.2684, found 501.2692.

Di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate

Di-tert-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))dicarbamate (2 g, 4.18 mmol, 1.0 eq.) was dissolved in 80 ml of DCM. 20 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 10 ml of DMF and to this solution is added 2-((*tert*butoxycarbonyl)amino)-2-methylpropanoic acid (1.87 g, 9.19 mmol, 2.2 eq.), HOBt (1.98 g, 14.63 mmol, 3.5 eq.), EDCI (2.27 g, 14.63 mmol, 3.5 eq.) and DIPEA (4.37 ml, 25.07 mmol, 6 eq.). Solvent was then evaporated and the crude product was washed with water (3 x 30 ml) and MeCN (3 x 30 ml) to obtained the desired white product.

Di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate: (0.88 g, 1.36 mmol, 32.5% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 9.17 (s, 2H), 7.84 (s, 2H), 7.61 (s, 4H), 7.35 (s, 2H), 1.43 (s, 18H), 1.37 (s, 12H), 1.28 (s, 12H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 178.40, 177.15, 159.76, 138.59, 124.34, 84.68, 61.19, 60.64, 32.06, 29.30, 28.71. HRMS (ESI) calcd for C₃₂H₅₂N₆O₈Na [M+Na]⁺: 671.3739, found 671.3751.

Di-tert-butyl ((((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2methyl-1-oxopropane-1,2-diyl))dicarbamate

Di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate (0.5 g, 0.77 mmol, 1.0 eq.) was dissolved in 30 ml of DCM. 10 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 5 ml of DMF and to this solution is added 2-((*tert*-butoxycarbonyl)amino)-2-methylpropanoic acid (0.35 g, 1.70 mmol, 2.2 eq.), HOBt (0.36 g, 2.80 mmol, 3.5 eq.), EDCI (0.42 g, 2.80 mmol, 3.5 eq.) and DIPEA (0.81 ml, 4.62 mmol, 6 eq.). Solvent was then evaporated and the crude product was washed with water (3 x 10 ml) and MeCN (3 x 10 ml) to obtained the desired white product. *Di-tert*-butyl ((((((1,4-phenylenebis(azanediyl)))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-

diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate: (0.12 g, 0.15 mmol, 19.0% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 9.15 (s, 2H), 8.29 (s, 2H), 7.68 (s, 4H), 7.41 (s, 2H), 1.42 (d, J = 2.5 Hz, 32H), 1.34 (s, 12H), 1.29 (s, 12H). HRMS (ESI) calcd for C₄₀H₆₆N₈O₁₀Na [M+Na]⁺: 841.4794, found 841.4801.

6-(phenylcarbamoyl)picolinic acid



To a stirred solution of 2,6-pyridinedicarboxylic acid (5 g, 29.9 mmol, 2.5 eq.) in anhydrous DMF (75 ml), DIPEA (4.59 ml, 26.3 mmol, 2.2 eq.) was added. The solution was stirred for 20 mins. HATU (4.55 g, 12.0 mmol, 1.0 eq.) was then added. Aniline (1.11 g, 12.0 mmol, 1.0 eq.) was added after 20 mins. The reaction was allowed to stir at rtp for 16 hrs. The reaction mixture was diluted with ethyl acetate (200 ml). The organic layer was washed by 0.5 M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml) and brine (3 x

50 ml) and then dried with anhydrous MgSO₄. The organic layer was was collected, dried and purified by column chromatography. **6-(phenylcarbamoyl)picolinic acid**: (0.88 g, 3.63 mmol, 30.4% yield), ¹H NMR (400 MHz, MeOD, 298K, δ): 8.44 (dd, *J* = 7.8, 1.1 Hz, 1H), 8.36 (dd, *J* = 7.8, 1.2 Hz, 1H), 8.20 (t, *J* = 7.8 Hz, 1H), 7.90 – 7.80 (m, 2H), 7.45 – 7.31 (m, 2H), 7.16 (ddt, *J* = 8.7, 7.4, 1.2 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 170.22, 166.09, 154.06, 150.50, 143.23, 141.84, 132.43, 131.30, 129.54, 128.29, 124.26. HRMS (ESI) calcd for C₁₃H₉N₂O₃ [M-H]⁻: 241.0619, found 241.0582.

4-(6-(phenylcarbamoyl)picolinamido)benzoic acid



To a stirred solution of 6-(phenylcarbamoyl)picolinic acid (1 g, 4.13 mmol, 1.0 eq.) in anhydrous DMF (20 ml), DIPEA (1.59 ml, 9.1 mmol, 2.2 eq.) was added. The solution was stirred for 20 mins. HATU (1.57 g, 4.13 mmol, 1.0 eq.) was then added. Then methyl 4-aminobenzoate (0.62 g, 4.13 mmol, 1.0 eq.) was added and the mixture was reacted for 16 hrs at rtp. The reaction mixture was diluted with ethyl acetate (100 ml). The organic layer was washed by 0.5 M LiCl (3 x 30 ml), 5% HCl (3 x 30 ml), 0.5 M K₂CO₃ (3 x 30 ml), brine (3 x 30 ml) and then dried with anhydrous MgSO₄. The organic layer was was collected, dried to obtain the desired white solid. The white solid was then dissolved in THF (100 ml), lithium hydroxide monohydrate (0.34 g, 7.99 mmol, 3.0 eq) in minimum amount of water was added. The reaction was stirred for 16 hrs at rtp. THF was evaporated. Water (50 ml) was then added. 1% HCl was added dropwise to acidify the water phase to pH 3. Then DCM (5 x 20 ml) was added and the organic phase was collected, dried to obtain the desired white solid. **4-(6-(phenylcarbamoyl)picolinamido)benzoic acid**: (0.84 g, 2.32 mmol, 87.3% yield), ¹H NMR (400 MHz, CDCl₃, 298K, δ): 11.24 (s, 1H), 11.05 (s, 1H), 8.43 (d, *J* = 7.7 Hz, 2H), 8.33 (dd, *J* = 8.5, 7.0 Hz, 1H), 8.09 (d, *J* = 8.8 Hz, 2H), 8.03 (d, *J* = 8.7 Hz, 2H), 7.98 – 7.89 (m, 2H), 7.46 (t, *J* = 7.9 Hz, 2H), 7.21 (t, *J* = 7.4 Hz, 1H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 167.38, 162.50, 162.04, 149.38, 148.97, 142.58, 140.53, 138.48, 130.80, 129.27, 126.67, 125.98, 124.88, 121.52, 120.66. HRMS (ESI) calcd for C₂₀H₁₅N₃O₄Na [M+Na]⁺: 384.0955, found 384.0964.

*N*²,*N*²'-(((((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2methyl-1-oxopropane-1,2-diyl))bis(*N*⁶-phenylpyridine-2,6-dicarboxamide), L9

Di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate (0.1 g, 0.12 mmol, 1.0 eq.) was dissolved in 4 ml of DCM. 1 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 3 ml of DMF and to this solution is added 6-(phenylcarbamoyl)picolinic acid (0.065 g, 0.27 mmol, 2.2 eq.), HATU (0.10 g, 0.27 mmol, 2.2 eq.), and DIPEA (0.13 ml, 0.73 mmol, 6 eq.). Solvent was then evaporated and the crude product was washed with water (3 x 10 ml) and MeCN (3 x 10 ml) to obtained the desired white product. **L9**: (53.3 mg, 0.050 mmol, 41.0% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 10.97 (s, 2H), 9.44 (s, 2H), 9.12 (s, 2H), 8.42 – 8.30 (m, 4H), 8.24 (d, J = 4.5 Hz, 4H), 7.89 (d, J = 7.9 Hz, 4H), 7.63 (d, J = 13.0 Hz, 6H), 7.46 (t, J = 7.8 Hz, 2H), 7.21 (t, J = 7.4 Hz, 2H), 1.63 (s, 12H), 1.54 (s, 12H), 1.29 (s, 12H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 175.20, 173.91, 173.61, 163.95, 162.14, 149.30, 148.87, 140.31, 138.37, 135.31, 129.30, 125.77, 125.53, 124.99, 121.70, 119.71, 57.04, 56.80, 25.74, 25.24. HRMS (ESI) calcd for C₅₆H₆₆N₁₂O₁₀Na [M+Na]⁺: 1089.4917, found 1089.4940.

Di-*tert*-butyl ((2*S*,2'*S*)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))dicarbamate



Di-tert-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))dicarbamate (1 g, 2.09 mmol, 1.0 eq.) was dissolved in 40 ml of DCM. 10 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 15 ml of DMF and to this solution is added (Tert-butoxycarbonyl)-*L*-leucine (1.06 g, 4.60 mmol, 2.2 eq.), HOBt (0.99 g, 7.31 mmol, 3.5 eq.), EDCI (1.14 g, 7.31 mmol, 3.5 eq.) and DIPEA (2.19 ml, 12.54 mmol, 6 eq.) and allowed to react for 16 hrs. The reaction mixture was diluted with ethyl acetate and washed with 1M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 30 ml). The organic phase was then collected and evaporated to obtain the desired white product. **Di-tert-butyl** ((25,2'S)-(((1,4-phenylenebis(azanediyl)))bis(2-methyl-1-oxopropane-1,2-

diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))dicarbamate: (1.28 g, 1.82

mmol, 86.9% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 8.95 (s, 1H), 8.14 (s, 1H), 7.51 (s, 2H), 7.08 (d, J = 6.2 Hz, 1H), 3.89 (q, J = 6.9 Hz, 1H), 1.68 – 1.51 (m, 1H), 1.40 (d, J = 31.4 Hz, 17H), 0.87 (dd, J = 8.4, 6.6 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 173.05, 172.59, 156.59, 134.45, 120.74, 80.75, 57.84, 53.95, 40.51, 28.18, 25.62, 24.96, 24.72, 22.77, 21.79. HRMS (ESI) calcd for C₃₆H₆₀N₆O₈Na [M+Na]⁺:727.4365, found 727.4377.

Di-tert-butyl ((((2S,2'S)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))bis(azanediyl))bis(2methyl-1-oxopropane-1,2-diyl))dicarbamate



Di-*tert*-butyl ((2*S*,2'*S*)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))dicarbamate (1 g, 1.42 mmol, 1.0 eq.) was dissolved in 40 ml of DCM. 10 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 15 ml of DMF and to this solution is added 2-((*tert*-butoxycarbonyl)amino)-2-methylpropanoic acid (0.63 g, 3.12 mmol, 2.2 eq.), HOBt (0.67 g, 4.97 mmol, 3.5 eq.), EDCl (0.77 g, 4.97 mmol, 3.5 eq.) and DIPEA (1.48 ml, 8.51 mmol, 6 eq.) and allowed to react for 16 hrs. The reaction mixture was diluted with ethyl acetate and washed with 1M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 30 ml). The organic phase was then collected and evaporated to obtain the desired white product. **Di-tert-butyl** ((((2,2'S)-(((1,4-phenylenebis(azanediyl)))bis(2-methyl-1-oxopropane-1,2-

diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))bis(azanediyl))bis(2-

methyl-1-oxopropane-1,2-diyl))dicarbamate: (1 g, 1.14 mmol, 80.6%). ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 8.91 (s, 1H), 8.05 (s, 1H), 7.93 (s, 1H), 7.59 (s, 2H), 7.21 (s, 1H), 4.04 – 3.90 (m, 1H), 1.70 – 1.55 (m, 2H), 1.55 – 1.16 (m, 22H), 0.86 (dd, *J* = 15.6, 6.2 Hz, 6H). HRMS (ESI) calcd for C₄₄H₇₄N₈O₁₀Na [M+Na]⁺: 897.5420, found 897.5440.

6-(isopropylcarbamoyl)picolinic acid



To a stirred solution of 6-(*tert*-butoxycarbonyl)picolinic acid (3 g, 13.43 mmol, 1.1 eq.) in anhydrous DMF (50 ml), DIPEA (3.2 ml, 18.33 mmol, 1.5 eq.) was added. The solution was stirred for 20 mins. HOBt (1.82 g, 13.43 mmol, 1.1 eq.), EDCI (2.09 g, 13.43 mmol, 1.1 eq.) were then added Then isopropylamine (0.72 g, 12.22 mmol, 1.0 eq.) was added and the mixture was reacted for 16 hrs at rtp. The reaction mixture was diluted with ethyl acetate (150 ml). The organic layer was washed by 0.5 M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 50 ml) and then dried with anhydrous MgSO₄. The organic layer was was collected, dried to obtain the white solid. The white solid was then dissolved in 80 ml of DCM. 10 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 2 hr. The solvent was then evaporated to obtain white solid. **6-(isopropylcarbamoyl)picolinic acid**: (2.06 g, 9.89 mmol, 81.0%). ¹H NMR (400 MHz, CDCl₃, 298K, δ): 8.43 (dd, *J* = 7.9, 1.2 Hz, 1H), 8.33 (dd, *J* = 7.8, 1.2 Hz, 1H), 8.16 – 8.05 (m, 2H), 4.28 (dp, *J* = 8.2, 6.5 Hz, 1H), 1.28 (d, *J* = 6.6 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃, 298K, δ): 166.17, 163.15, 149.33, 145.45,

139.69, 127.55, 126.75, 42.47, 22.32. HRMS (ESI) calcd for $C_{10}H_{11}N_2O_3$ [M-H]⁻: 207.0775, found 207.0775.

4-(6-(isopropylcarbamoyl)picolinamido)benzoic acid



To a stirred solution of 6-(isopropylcarbamoyl)picolinic acid (2.5 g, 12.0 mmol, 1.0 eq.) in anhydrous DMF (50 ml), DIPEA (3.14 ml, 18 mmol, 1.5 eq.) was added. The solution was stirred for 20 mins. HATU (5.02 g, 13.2 mmol, 1.1 eq.) was then added. Then methyl 4-aminobenzoate (1.81 g, 12.0 mmol, 1.0 eq.) was added and the mixture was reacted for 16 hrs at rtp. The reaction mixture was diluted with ethyl acetate (100 ml). The organic layer was washed by 0.5 M LiCl (3 x 50 ml), 5% HCl (3 x 50 ml), 0.5 M K₂CO₃ (3 x 50 ml), brine (3 x 50 ml) and then dried with anhydrous MgSO₄. The organic layer was was collected, dried to obtain the desired white solid. The white solid was then dissolved in THF (300 ml), lithium hydroxide monohydrate (1.53 g, 36.0 mmol, 3.0 eq) in minimum amount of water was added. The reaction was stirred for 16 hrs at rtp. THF was evaporated. Water (100 ml) was then added. 1% HCl was added dropwise to acidify the water phase to pH 3. Then DCM (5 x 50 ml) was added and the organic phase was collected, dried to obtain the desired white solid. 4-(6-(isopropylcarbamoyl)picolinamido)benzoic acid: (1.9 g, 5.80 mmol, 48.3%). ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 12.78 (s, 1H), 10.97 (s, 1H), 9.05 (d, *J* = 8.0 Hz, 1H), 8.29 (dd, J = 7.4, 1.5 Hz, 1H), 8.25 – 8.15 (m, 2H), 7.96 (s, 4H), 4.14 (dp, J = 7.9, 6.5 Hz, 1H), 1.24 (d, J = 6.6 Hz, 6H). ¹³C NMR (125 MHz, (CD₃)₂SO, 298K, δ): 167.45, 162.75, 162.63,

149.86, 148.85, 142.51, 140.19, 130.84, 126.78, 125.71, 125.45, 120.91, 41.69, 22.83. HRMS (ESI) calcd for C₁₇H₁₆N₃O₄ [M-H]⁻: 326.1146, found 326.1176.

N², N²'-((((2S,2'S)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))bis(azanediyl))bis(2methyl-1-oxopropane-1,2-diyl))bis(N6-isopropylpyridine-2,6-dicarboxamide), L10

Di-tert-butyl ((((2S,2'S)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))bis(azanediyl))bis(2methyl-1-oxopropane-1,2-diyl))dicarbamate (0.34 g, 0.39 mmol, 1.0 eq.) was dissolved in 8 ml of DCM. 2 ml of trifluoroacetic acid was added and the reaction was maintained in room temperature for 1 hr. The solvent was then evaporated to obtain white solid. The residue was dissolved in 3 ml of DMF and to this solution is added 4-(6-(isopropylcarbamoyl)picolinamido)benzoic acid (0.28 g, 0.85 mmol, 2.2 eq.), HATU (0.32 g, 0.85 mmol, 2.2 eq.), and DIPEA (0.4 ml, 2.33 mmol, 6 eq.). Solvent was then evaporated and redissolved in EA (50 ml). The organic phase was washed with 1M LiCl (3 x 10 ml), 5% HCl (3 x 10 ml), 0.5 M K₂CO₃ (3 x 10 ml), brine (3 x 10 ml). The organic phase was then collected, evaporated and purified by column chromatography to obtained the desired white product. **L10**: (0.11 g, 0.10 mmol, 26.8% yield), ¹H NMR (400 MHz, (CD₃)₂SO, 298K, δ): 11.00 (s, 1H), 9.12 (d, *J* = 8.0 Hz, 1H), 8.91 (s, 1H), 8.61 (s, 1H), 8.35 (dd, J = 7.4, 1.5 Hz, 1H), 8.32 – 8.20 (m, 2H), 8.12 (d, J = 6.7 Hz, 1H), 8.02 (q, J = 8.9 Hz, 4H), 7.80 (s, 1H), 7.57 (s, 2H), 4.20 (dq, J = 13.7, 6.8 Hz, 1H), 4.05 – 3.93

(m, 1H), 1.70 - 1.36 (m, 13H), 1.36 - 1.21 (m, 7H), 0.85 (dd, J = 19.1, 6.1 Hz, 7H). ¹³C NMR (101 MHz, (CD₃)₂SO, 298K, δ): 175.81, 173.23, 172.25, 166.75, 162.70, 162.52, 149.79, 148.84, 141.30, 140.15, 135.10, 129.96, 129.10, 125.61, 125.36, 120.53, 120.18, 57.09, 56.92, 53.21, 31.42, 25.85, 25.82, 25.50, 25.36, 24.84, 23.58, 22.77, 22.53, 21.67, 14.43. HRMS (ESI) calcd for C₆₈H₈₈N₁₄O₁₂Na [M+Na]⁺: 1315.6598, found 1315.6675.

4.5 References

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General experimental procedures

Synthesis

Unless otherwise stated, all chemicals and solvents were obtained commercially and without further purification before used. All moisture-sensitive compounds were handled using standard Schlenk line techniques. All moisturesensitive reactions were conducted under a nitrogen atmosphere in glasswares that were oven-dried at 140 °C overnight prior to use. Anhydrous dimethylformamide (DMF) and N,N-Diisopropylethylamine (DIPEA) were purchased from Acros. Solvents were used as received. Grace silica gel 60 (40-63 mesh) was used for column chromatography. 1D NMR spectra were recorded on a Bruker Ultrashield Advance Pro 400 MHz instrument and Jeol ECZ500R 500 MHz NMR spectrometer and the chemical shifts were determined with tetramethylsilane (TMS) or solvents in parts per million (ppm). 2D NMR spectra were recorded on Jeol ECZ500R 500 MHz NMR spectrometer. ESI-HRMS were performed on a Waters Synapt G2-Si Quadrupole MS. Elemental analyses were performed on an Elementar Vario EL cube elemental analyzer. All lanthanide salts were purchased from Sigma-Aldrich with purity 99.9% and used without further purification.

Photophysical measurement

Single-photon luminescence spectra were recorded using an Edinburgh Instrument FLSP920 spectrophotometer that was equipped with Xe900 continuous xenon lamp, µF920 microsecond flashlamp and a single photon counting Photomultiplier Tube. The excitation and emission spectra recorded on the FLSP920 were corrected with the correction file from the F900 software. CD spectra were recorded with a Jasco J-801 spectropolarimeter with a 1 cm cell at 25 °C. and presented as $\Delta \epsilon$ in M⁻¹cm⁻¹. Hellma quartz cuvettes (1mm path length) were employed. All spectra were baselined subtracted with the blank solvent. UV-Visible absorption spectra were recorded with an Agilent Technologies Cary 8454 spectrophotometer. CPL spectra in chapter 2a were recorded on a Olis 17 UV/VIS/NIR/CD/CPL spectrophotometer On-Line Instrument Systems. CPL spectra in chapter 2b on a custom-build spectrometer consisting of a laserdriven light source (Energetig EQ-99 LDLS, spectral range 170–2100 nm) coupled to an Acton SP2150 monochromator (600 g/nm, 300 nm Blaze)CPL spectra in chapter 3 were recorded with a Jasco CPL-300 circularly polarized luminescence spectrophotometer with a 1 cm cell at 25 °C. All photophysical measurement were done in triplicate.

Single crystal X-ray diffraction

Measurements of crystal data were carried out on a Bruker Smart 1000 system equipped with an APEX II CCD device for $Eu_2L1^{SS}_3$, $Eu_2L2^{RR}_3$ and $L2^{RR}$ with graphite monochromated Mo-K α radiation at room temperature (HKPolyU). Multi-scan absorption correction was applied by SADABS program, and the SAINT program was utilized for integration of the diffraction profiles. The structures were solved by direct method and was refined by a full matrix least-squares treatment on *F*2 using the SHELXTL programme system.3 Structure Eu₂L2^{RR}₃ contains highly disordered side arms, and were removed from the refinement using the PLATON/SQUEEZE program. Although the side arms cannot be defined clearly, the helical structure was clearly observed. Crystallographic data for the structural analysis has been deposited with Cambridge Crystallographic Data Centre, CCDC No. for Eu₂L1^{SS}₃ is 996693; No. for Eu₂L2^{RR}₃ is 991735; No. for L2^{RR} is 1003776.

Tb₂L1^{SS}₃ was collected on a Bruker D8-Venture Diffractometer System with a micro-focus Mo-K α radiation (HKPolyU). Of a total of 210003 reflections collected, 31661 were unique ($R_{int} = 0.0893$). Multi-scan absorption correction was applied by SADABS program, and the SAINT program utilised for the integration of the diffraction profile. The structure was solved by direct method and was refined by a full-matrix least-squares treatment on F^2 using the SHELXLE programme system. The final R-values were: R1 = 0.0524 [I>2s(I)] and wR2 = 0.1159 (all data). Crystallographic data for the structural analysis has been deposited with Cambridge Crystallographic Data Centre, CCDC No. for Tb₂L1^{SS}₃ is 1009616.

Data of Eu₄L3^{ss}₆ were collected on a Bruker D8 Venture single crystal diffractometer with a Bruker Photon II detector and an Incoatec IµS 3 Cu micro-focus X-ray source (Newcastle University). These data were then processed within the APEX3 software suite and solved and refined using the OLEX2 interface to the SHELX suite of programs. The data processing was truncated to only include the frames from

the unit cell data collection along with the first 3 experimental 'runs'. Crystallographic data for the structural analysis has been deposited with Cambridge Crystallographic Data Centre, CCDC No. for $Eu_4L3^{ss}_{6}$ is 2014348.

Single crystals of Eu₂Gd₂L3^{SS}₆ C₂₂₅H₁₉₂Eu₂F₂₇Gd₂N₃₆O₅₁S₉ was selected and mounted on a standard Kapton micromount, using neat paratone oil to try and reduce solvent loss during mounting procedure on a Bruker D8 Vantage diffractometer (Newcastle University). The crystal was kept at 220.0 K during data collection. Using Olex2, the structure was solved with the XT structure solution program using Intrinsic Phasing and refined with the XL refinement package using Least Squares minimisation. Crystallographic data for the structural analysis has been deposited with Cambridge Crystallographic Data Centre, CCDC No. for Eu₂Gd₂L3^{SS}₆ is 2092789.

Appendix

Chapter 2a



Figure S2a-1. Excitation spectrum of [Eu₂(L1^{RR})₃](CF₃SO₃)₆ (powder, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-2. Emission spectrum of $[Eu_2(L1^{RR})_3](CF_3SO_3)_6$ (powder, lex = 369 nm, slits = 5-1, filter 455nm).



Figure S2a-3. Excited state decay curve and its mono exponential fit of $Eu_2(L1^{RR})_3](CF_3SO_3)_6$ (powder, lex = 369 nm, slits = 15-3, filter 455nm).



Figure S2a-4. Excitation spectrum of [Eu₂(L1^{SS})₃](CF₃SO₃)₆ (powder, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-5. Emission spectrum of $[Eu_2(L1^{SS})_3](CF_3SO_3)_6$ (powder, lex = 369 nm, slits = 5-1, filter 455nm).



Figure S2a-6. Excited state decay curve and its mono exponential fit of $[Eu_2(L1^{ss})_3](CF_3SO_3)_6$ (powder, lex = 369 nm, slits = 15-3, filter 455nm).



Figure S2a-7. Excitation spectrum of [Eu₂(L2^{RR})₃](CF₃SO₃)₆ (powder, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-8. Emission spectrum of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (powder, lex = 372 nm, slits = 5-1, filter 455nm).



Figure S2a-9. Excited state decay curve and its mono exponential fit of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (powder, lex = 372 nm, slits = 15-3, filter 455nm).



Figure S2a-10. Excitation spectrum of $[Eu_2(L2^{SS})_3](CF_3SO_3)_6$ (powder, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-11. Emission spectrum of $[Eu_2(L2^{ss})_3](CF_3SO_3)_6$ (powder, lex = 373 nm, slits = 5-1, filter 455nm).



Figure S2a-12. Excited state decay curve and its mono exponential fit of $[Eu_2(L2^{ss})_3](CF_3SO_3)_6$ (powder, lex = 373 nm, slits = 15-3, filter 455nm).



Figure S2a-13. Excitation spectrum of $[Eu_2(L1^{RR})_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M in MeCN, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-14. Emission spectrum of [Eu₂(L1^{RR})₃](CF₃SO₃)₆ (3.23 × 10⁻⁵ M in MeCN, lex = 363 nm, slits = 5-1, filter 455nm).



Figure S2a-16. Excited state decay curve and its mono exponential fit of $[Eu_2(L1^{RR})_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M in MeCN, lex = 363 nm, slits = 15-3, filter 455nm).



Figure S2a-16. Excitation spectrum of $[Eu_2(L1^{SS})_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M in MeCN, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-17. Emission spectrum of $[Eu_2(L1^{SS})_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M in MeCN, lex = 358 nm, slits = 5-1, filter 455nm).



Figure S2a-18. Excited state decay curve and its mono exponential fit of $[Eu_2(L1^{ss})_3](CF_3SO_3)_6$ (3.23 × 10⁻⁵ M in MeCN, lex = 358 nm, slits = 15-3, filter 455nm).



Figure S2a-19. Excitation spectrum of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (3.47 × 10⁻⁵ M in MeCN, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-20. Emission spectrum of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (3.47 × 10⁻⁵ M in MeCN, lex = 351 nm, slits = 5-1, filter 455nm).



Figure S2a-21. Excited state decay curve and its mono exponential fit of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (3.47 × 10⁻⁵ M in MeCN, lex = 351 nm, slits = 15-3, filter 455nm).



Figure S2a-22. Excitation spectrum of $[Eu_2(L2^{SS})_3](CF_3SO_3)_6$ (3.15 × 10⁻⁵ M in MeCN, lem = 616 nm, slits = 5-1, filter 455nm).



Figure S2a-23. Emission spectrum of $[Eu_2(L2^{SS})_3](CF_3SO_3)_6$ (3.15 × 10⁻⁵ M in MeCN, lex = 351 nm, slits = 5-1, filter 455nm).



Figure S2a-24. Excited state decay curve and its mono exponential fit of $[Eu_2(L2^{ss})_3](CF_3SO_3)_6 (3.15 \times 10^{-5} \text{ M in MeCN}, \text{lex} = 351 \text{ nm}, \text{slits} = 15-3, \text{ filter } 455 \text{ nm}).$



Figure S2a-25. Excitation spectrum of $[Gd_2(L1^{SS})_3](CF_3SO_3)_6$ (2.88 × 10⁻⁶ M in 1:4, v/v of MeOH/EtOH at 77K, lem = 450 nm, slits = 5-1).



Figure S2a-26. Emission spectrum of $[Gd_2(L1^{SS})_3](CF_3SO_3)_6$ (2.88 × 10⁻⁶ M in 1:4 of MeOH/EtOH at 77K, lex = 340 nm, slits = 5-1).


Figure S2a-27. Excited state decay curve and its mono exponential fit of $[Gd_2(L1^{SS})_3](CF_3SO_3)_6$ (2.88 × 10⁻⁶ M in 1:4 of MeOH/EtOH at 77K, lex = 340 nm, slits = 15-3).



Figure S2a-28. Emission spectrum of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (3.15 × 10⁻⁵ M in MeCN or d-MeCN, lex = 351 nm, slits = 5-1, filter 455nm).



Figure S2a-29. Excited state decay curve and its mono exponential fit of $[Eu_2(L2^{RR})_3](CF_3SO_3)_6$ (3.15 × 10⁻⁵ M in d-MeCN, lex = 351 nm, slits = 15-3, filter 455nm).



Figure S2a-30. Preliminary results of CPL for $[Eu_2(L1)_3](CF_3SO_3)_6(2.30 \times 10^{-3} \text{ M}, 10 \text{ mm} \text{ cuvette, MeCN}).^5$



Figure S2a-31. ¹H NMR spectrum of **(S)-6-(1-phenylethyl carbamoyl)picolinic acid** in CD₃OD. The insets show the expansion of the corresponding region as indicated.



Figure S2a-32. ¹³C NMR spectrum of **(S)-6-(1-phenylethyl carbamoyl)picolinic acid** in CD_3OD . The insets show the expansion of the corresponding region as indicated.



Figure S2a-33. ¹H NMR spectrum of $L1^{RR}$ in $(CD_3)_2SO$. The insets show the expansion of the corresponding region as indicated.



Figure S2a-34. ¹³C NMR spectrum of $L1^{RR}$ in $(CD_3)_2SO$. The insets show the expansion of the corresponding region as indicated.



Figure S2a-35. ¹³C NMR spectrum of $[La_2(L1^{SS})_3](CF_3SO_3)_6$ in CD₃CN. The insets show the expansion of the corresponding region as indicated. The chemical shift with an asteroid is not very clear. It was shown to correlate a chemical shift at 2.10 ppm (singlet) in the corresponding ¹H NMR spectrum in HSQC experiment.



Figure S2a-36. ¹H NMR spectrum of **(S)-6-(2-phenylpropyl carbamoyl)picolinic acid** in CD₃OD. The insets show the expansion of the corresponding region as indicated.



Figure S2a-37. ¹³C NMR spectrum of **(S)-6-(2-phenylpropyl carbamoyl)picolinic acid** in CD_3OD . The insets show the expansion of the corresponding region as indicated.



Figure S2a-38. ¹H NMR spectrum of **L2**^{RR} in CDCl₃. The insets show the expansion of the corresponding region as indicated.



Figure S2a-39. ¹³C NMR spectrum of L2^{RR} in CDCl₃. The insets show the expansion of the corresponding region as indicated.



Figure S2a-40. ¹³C NMR spectrum of $[La_2(L2^{RR})_3](CF_3SO_3)_6$ in CD₃CN. The insets show the expansion of the corresponding region as indicated.







Figure S2a-42. HPLC spectra of **or (S)-6-(2-phenylpropyl carbamoyl)picolinic acid** and (*R*)-6-(2-phenylpropylcarbamoyl)picolinic acid.

Chapter 2b



Figure S2b-1. ¹H NMR of L3^{SS} in (CD₃)₂SO.



Figure S2b-2. ¹³C NMR of L3^{SS} in (CD₃)₂SO.



Figure S2b-3. ¹H NMR of $L4^{RR}$ in $(CD_3)_2SO$.



Figure S2b-4. ¹³C NMR of L4^{RR} in (CD₃)₂SO.



Figure S2b-5. ¹H NMR of $[La_2(L3^{SS})_3]$ in CD₃CN.



Figure S2b-6. ¹³C NMR of [La₂(**L3**^{SS})₃] in CD₃CN.



Figure S2b-7. ¹H NMR of [Sm₂(L3^{RR})₃] in CD₃CN.



Figure S2b-8. ¹³C NMR of [Sm₂(L3^{RR})₃] in CD₃CN.



Figure S2b-9. ¹H NMR of [Eu₂(L3^{SS})₃] in CD₃CN.



Figure S2b-10. ¹³C NMR of [Eu₂(L3^{RR})₃] in CD₃CN.



Figure S2b-11. ¹H NMR of [Lu₂(L3^{RR})₃] in CD₃CN.



Figure S2b-12. ¹³C NMR of [Lu₂(L3^{SS})₃] in CD₃CN.



Figure S2b-13. ¹H NMR of $[Eu_4(L3^{SS})_6]$ in CD₃CN.



Figure S2b-14. ¹³C NMR of $[Eu_4(L3^{SS})_6]$ in CD₃CN.



Figure S2b-15. ¹H NMR of $[Sm_4(L3^{SS})_6]$ in CD₃CN.



Figure S2b-16. ¹³C NMR of $[Sm_4(L3^{RR})_6]$ in CD₃CN.



Figure S2b-17. ¹H NMR of $[Lu_4(L3^{ss})_6]$ in CD₃CN.



Figure S2b-18. ¹H NMR of [La₂(L4^{SS})₃] in CD₃CN.



Figure S2b-19. ¹³C NMR of $[La_2(L4^{RR})_3]$ in CD₃CN.



Figure S2b-20. ¹H NMR of [Sm₂(L4^{SS})₃] in CD₃CN.



Figure S2b-21. ¹³C NMR of [Sm₂(**L4**^{SS})₃] in CD₃CN.



Figure S2b-22. ¹H NMR of [Eu₂(L4^{SS})₃] in CD₃CN.



Figure S2b-23. ¹³C NMR of [Eu₂(L4^{RR})₃] in CD₃CN.



Figure S2b-24. ¹H NMR of [Lu₂(L4^{RR})₃] in CD₃CN.



Figure S2b-25. ¹³C NMR of [Lu₂(L4^{RR})₃] in CD₃CN.



Figure S2b-26. COSY NMR of L3 in (CD₃)₂SO.



Figure S2b-27. COSY NMR of L4 in (CD₃)₂SO.



Figure S2b-28. COSY NMR of [La₂(L3^{RR})₃] in CD₃CN.



Figure S2b-29. COSY NMR of [Eu₂(L3^{RR})₃] in CD₃CN.



Figure S2b-30. COSY NMR of $[Sm_2(L3^{RR})_3]$ in CD₃CN.



Figure S2b-31. COSY NMR of [Lu₂(L3^{RR})₃] in CD₃CN.



Figure S2b-32. COSY NMR of [Sm₄(L3^{RR})₆] in CD₃CN.



Figure S2b-33. COSY NMR of [Eu₄(L3^{RR})₆] in CD₃CN.



Figure S2b-34. COSY NMR of $[Lu_4(L3^{RR})_6]$ in CD₃CN.



Figure S2b-35. COSY NMR of [La₂(L4^{RR})₃] in CD₃CN.



Figure S2b-36. COSY NMR of $[Sm_2(L4^{RR})_3]$ in CD₃CN.



Figure S2b-37. COSY NMR of [Eu₂(L4^{RR})₃] in CD₃CN.



Figure S2b-38. COSY NMR of [Lu₂(L4^{RR})₃] in CD₃CN.



Figure S2b-39. ESI-HRMS of bimetallic helicate $[La_2L3_3]$. (A) The full spectrum. Simulated m/z for $[La_3L3_3 + 4 \text{ OTf}^-]^{2+}$ is 1355.6841(100%), Experimental found m/z is 1355.6792(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.

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Figure S2b-40. ESI-HRMS of bimetallic helicate $[Sm_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Sm_2L3_3 + 3 \text{ OTf}^-]^{3+}$ is 861.8131(100%), Experimental found m/z is 861.8192(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks



Figure S2b-41. ESI-HRMS of bimetallic helicate $[Eu_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Eu_3L3_3 + 4 \text{ OTf}]^{2+}$ is 1369.1985(100%), Experimental found m/z is 1369.1962(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



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Figure S2b-42. ESI-HRMS of bimetallic helicate $[Gd_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Gd_2L3_3 + 3 \text{ OTf}^-]^{3+}$ is 866.4836 (100%), Experimental found m/z is 866.4828(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-43. ESI-HRMS of bimetallic helicate $[Tb_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Tb_2L3_3 + OTf^-2H^+]^{3+}$ is 767.5113(100%), Experimental found m/z is 767.5154(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-44. ESI-HRMS of bimetallic helicate $[Lu_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Lu_2L3_3 + 30Tf]^{3+}$ is 878.1615(100%), Experimental found m/z is 878.1616(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-45. ESI-HRMS of tetrahedron $[Eu_4L3_6]$. (A) The full spectrum. Simulated m/z for $[Eu_4L3_6 + 9 \text{ OTf}^-]^{3+}$ is 1875.2490(100%), Experimental found m/z is 1875.2516(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-46. ESI-HRMS of tetrahedron $[Sm_4L3_6]$. (A) The full spectrum. Simulated m/z for $[Sm_4L3_6 + 6 \text{ OTf}^- - \text{H}^+]^{5+}$ is 1034.1745(100%), Experimental found m/z is 1034.1748(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-47. ESI-HRMS of tetrahedron [Gd₄L**3**₆]. (A) The full spectrum. Simulated m/z for [Gd₄L**3**₆ + 8 OTf⁻]⁴⁺ is 1374.4520 (100%), Experimental found m/z is 1374.4512(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.


Figure S2b-48. ESI-HRMS of tetrahedron [Tb₄L3₆]. (A) The full spectrum. Simulated m/z for [Tb₄L3₆ + 6 OTf⁻ - H⁺]⁵⁺ is 1040.9802 (100%), Experimental found m/z is 1040.9800(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.

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Figure S2b-49. ESI-HRMS of tetrahedron [Lu₄L**3**₆]. (A) The full spectrum. Simulated m/z for [Lu₄L**3**₆ + 9OTf⁻]³⁺ is 1905.6090(100%), Experimental found m/z is 1905.6060(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-50. ESI-HRMS of bimetallic helicate $[La_2L4_3]$. (A) The full spectrum. Simulated m/z for $[La_2L4_3 + 3OTf^-]^{3+}$ is 1006.2012(100%), Experimental found m/z is 1006.2037(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.

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Figure S2b-51. ESI-HRMS of bimetallic helicate $[Sm_2L4_3]$. (A) The full spectrum. Simulated m/z for $[Sm_2L4_3 + 3OTf^-]^{3+}$ is 1013.8759(100%), Experimental found m/z is 1013.8749(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-52. ESI-HRMS of bimetallic helicate $[Eu_2L4_3]$. (A) The full spectrum. Simulated m/z for $[Eu_2L4_3 + 3OTf^-]^{3+}$ is 1015.2111(100%), Experimental found m/z is 1015.2097(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-53. ESI-HRMS of bimetallic helicate $[Gd_2L4_3]$. (A) The full spectrum. Simulated m/z for $[Gd_2L4_3 + 3OTf^-]^{3+}$ is 1018.5452(100%), Experimental found m/z is 1018.5452(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-54. ESI-HRMS of bimetallic helicate $[Tb_2L4_3]$. (A) The full spectrum. Simulated m/z for $[Tb_2L4_3 + 3OTf^-]^{3+}$ is 1019.5472(100%), Experimental found m/z is 1019.5485(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-55. ESI-HRMS of bimetallic helicate $[Lu_2L4_3]$. (A) The full spectrum. Simulated m/z for $[Lu_2L4_3 + OTf^- - 2H^+]^{3+}$ is 930.2509(100%), Experimental found m/z is 930.2548(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S2b-56. Luminescent data of $[Eu_2(L3^{SS})_3](OTf)_6$ (1.58 x 10⁻⁶ M in CH₃CN).(A) Excitation spectrum, λ_{em} = 616 nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, λ_{ex} = 311 nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, λ_{em} = 616 nm, slits = 6-5, filter 380 nm.



Figure S2b-57. Luminescent data of $[Eu_2(L3^{RR})_3](OTf)_6$ (1.60 x 10⁻⁵ M in CH₃CN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 311$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 6-5, filter 380 nm.



Figure S2b-58. Luminescent data of $[Eu_4(L3^{SS})_6](OTf)_{12}$ (2.50 x 10⁻⁶ M in CH₃CN).(A) Excitation spectrum, λ_{em} = 616 nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, λ_{ex} = 330 nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, λ_{em} = 616 nm, slits = 5-3, filter 380 nm.



Figure S2b-59. Luminescent data of $[Eu_4(L3^{RR})_6](OTf)_{12}$ (2.41 x 10⁻⁶ M in CH₃CN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5-3, filter 380 nm.



Figure S2b-60. Luminescent data of $[Gd_2(L3^{RR})_3](OTf)_6$ (6.13 x 10⁻⁶ M in 1:4 of MeOH/EtOH at 77K).(A) Excitation spectrum, $\lambda_{em} = 462$ nm, slits = 1.0-0.5, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 315$ nm, slits = 1.0-0.5, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 462$ nm, filter 380 nm.



Figure S2b-61. Luminescent data of $[Gd_2(L4^{SS})_3](OTf)_{12}$ (2.55 x 10⁻⁵ M in 1:4 of MeOH/EtOH at 77K).(A) Excitation spectrum, $\lambda_{em} = 480$ nm, slits = 2.0-0.5, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 345$ nm, slits = 2.0-0.5, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 480$ nm, filter 380 nm.



Figure S2b-62. Luminescent data of $[Gd_4(L3^{RR})_6](OTf)_{12}$ (9.05 x 10⁻⁶ M in 1:4 of MeOH/EtOH at 77K).(A) Excitation spectrum, λ_{em} = 466 nm, slits = 2.0-0.5, filter 380 nm. (b) Emission spectrum, λ_{ex} = 330 nm, slits = 2.0-0.5, filter 380 nm. (c) Excited state decay curve, λ_{em} = 466 nm, filter 380 nm.



Figure S2b-63. CD spectra of $[La_2(L3^{SS})_3](OTf)_6$ and $[La_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-64. CD spectra of $[Sm_2(L3^{SS})_3](OTf)_6$ and $[Sm_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-65. CD spectra of $[Eu_2(L3^{SS})_3](OTf)_6$ and $[Eu_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-66. CD spectra of $[Gd_2(L3^{SS})_3](OTf)_6$ and $[Gd_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-67. CD spectra of $[Tb_2(L3^{SS})_3](OTf)_6$ and $[Tb_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-68. CD spectra of $[Lu_2(L3^{SS})_3](OTf)_6$ and $[Lu_2(L3^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-69. CD spectra of [Sm₄(L3^{SS})₆](OTf)₁₂ and [Sm₄(L3^{RR})₆](OTf)₁₂ in CH₃CN.



Figure S2b-70. CD spectra of [Eu₄(L3^{SS})₆](OTf)₁₂ and [Eu₄(L3^{RR})₆](OTf)₁₂ in CH₃CN.



Figure S2b-71. CD spectra of $[Gd_4(L3^{SS})_6](OTf)_{12}$ and $[Gd_4(L3^{RR})_6](OTf)_{12}$ in CH₃CN.



Figure S2b-72. CD spectra of [Tb₄(L3^{SS})₆](OTf)₁₂ and [Tb₄(L3^{RR})₆](OTf)₁₂ in CH₃CN.



Figure S2b-73. CD spectra of [Lu₄(L3^{SS})₆](OTf)₁₂ and [Lu₄(L3^{RR})₆](OTf)₁₂ in CH₃CN.



Figure S2b-74. CD spectra of $[La_2(L4^{SS})_3](OTf)_6$ and $[La_2(L4^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-75. CD spectra of $[Sm_2(L4^{SS})_3](OTf)_6$ and $[Sm_2(L4^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-76. CD spectra of $[Eu_2(L4^{SS})_3](OTf)_6$ and $[Eu_2(L4^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-77. CD spectra of $[Gd_2(L4^{SS})_3](OTf)_6$ and $[Gd_2(L4^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-78. CD spectra of $[Tb_2(L4^{SS})_3](OTf)_6$ and $[Tb_2(L4^{RR})_3](OTf)_6$ in CH₃CN.



Figure S2b-79. CD spectra of $[Lu_2(L4^{SS})_3](OTf)_6$ and $[Lu_2(L4^{RR})_3](OTf)_6$ in CH₃CN.







Figure S2b-80. Three sets of Arrhenius plot of Eu tetrahedron-to-helicate transformation.









Figure S2b-81. Four sets of Arrhenius plot of Sm tetrahedron-to-helicate transformation.

Chapter 3



Figure S3-1. ESI-HRMS of bimetallic helicate $[Dy_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Dy_2L3_3 + 40Tf^-]^{2+}$ is 1379.7060(100%), Experimental found m/z is 1379.7075(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-2. ESI-HRMS of bimetallic helicate $[Ho_2L3_3]$. (A) The full spectrum. Simulated m/z for $[Ho_2L3_3 + 40Tf^-]^{2+}$ is 1381.7072(100%), Experimental found m/z is 1381.7081(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-3. ESI-HRMS of tetrahedron $[Eu_nGd_{4-n}L3_6]$ (prepared by ratio Eu/Gd: 3/1). (A) The full spectrum. Simulated m/z for $[Eu_3GdL3_6 + 7 \text{ OTf}^-]^{5+}$ is 1066.5690(100%), Experimental found m/z is 1066.5699(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-4. ESI-HRMS of tetrahedron $[Eu_nGd_{4-n}L3_6]$ (prepared by ratio Eu/Gd: 1/1). (A) The full spectrum. Simulated m/z for $[Eu_2Gd_2L3_6 + 7 \text{ OTf}^-]^{5+}$ is 1067.5697(100%), Experimental found m/z is 1067.5664(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-5. ESI-HRMS of tetrahedron [Eu_nGd_{4-n}L**3**₆] (prepared by ratio Eu/Gd: 1/3). (A) The full spectrum. Simulated m/z for [EuGd₃L**3**₆ + 7 OTf⁻]⁵⁺ is 1068.5703(100%), Experimental found m/z is 1068.5665(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-6. ESI-HRMS of tetrahedron $[Dy_4L3_6]$. (A) The full spectrum. Simulated m/z for $[Dy_4L3_6 + 7 \text{ OTf}^-]^{5+}$ is 1073.9743(100%), Experimental found m/z is 1073.9764(100%). Inset showing the experimental (upper) and calculated(lower) isotopic patterns. (B) Expanded region of the mass spectrum to show the possible assignments of the corresponding prominent peaks.



Figure S3-7. CD spectra of $[Eu_4(L3^{SS})_6]$ and $[Eu_4(L3^{RR})_6]$ in CH₃CN.



Figure S3-8. CD spectra of $[Eu_nGd_{4-n}L3^{SS}_6]$ and $[Eu_nGd_{4-n}L3^{RR}_6]$ in MeCN (prepared by ratio Eu/Gd: 1/3).



Figure S3-9. CD spectra of $[Eu_nGd_{4-n}L3^{SS}_6]$ and $[Eu_nGd_{4-n}L3^{RR}_6]$ in MeCN (prepared by ratio Eu/Gd: 1/1).



Figure S3-10. CD spectra of $[Eu_nGd_{4-n}L3^{SS}_6]$ and $[Eu_nGd_{4-n}L3^{RR}_6]$ in MeCN (prepared by ratio Eu/Gd: 1/2).



Figure S3-11. CD spectra of $[Gd_4(L3^{SS})_6]$ and $[Gd_4(L3^{RR})_6]$ in MeCN.



Figure S3-12. CD spectra of $[Eu_nSm_{4-n}L3^{SS}_6]$ and $[Eu_nSm_{4-n}L3^{RR}_6]$ in MeCN (prepared by ratio Eu/Sm: 1/1).



Figure S3-13. Luminescent data of $[Eu_4(L3^{SS})_6]$ (2.50 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5-3, filter 380 nm.


Figure S3-14. Luminescent data of $[Eu_4(L3^{RR})_6]$ (2.41 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5-3, filter 380 nm.



Figure S3-15. Luminescent data of $[Eu_nGd_{4-n}L3^{ss}_6]$ (prepared by ratio Eu/Gd: 3/1), (2.29 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5.0-3.0, filter 380 nm.



Figure S3-16. Luminescent data of $[Eu_nGd_{4-n}L3^{RR}_6]$ (prepared by ratio Eu/Gd: 3/1), (2.33 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5.0-3.5, filter 380 nm.



Figure S3-17. Luminescent data of $[Eu_nGd_{4-n}L3^{ss}_6]$ (prepared by ratio Eu/Gd: 1/1), (2.60 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5.0-4.0, filter 380 nm.



Figure S3-18. Luminescent data of $[Eu_nGd_{4-n}L3^{RR}_6]$ (prepared by ratio Eu/Gd: 1/1), (2.60 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 1.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 1.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 5.0-4.0, filter 380 nm.



Figure S3-19. Luminescent data of $[Eu_nGd_{4-n}L3^{ss}_6]$ (prepared by ratio Eu/Gd: 1/3), (2.44 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 2.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 2.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 6.0-5.0, filter 380 nm.



Figure S3-20. Luminescent data of $[Eu_nGd_{4-n}L3^{RR}_6]$ (prepared by ratio Eu/Gd: 1/3), (2.59 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 2.5-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 2.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 6.0-5.0, filter 380 nm.



Figure S3-21. Luminescent data of $[Eu_nSm_{4-n}L3^{ss}_6]$ (prepared by ratio Eu/Sm: 1/1), (2.19 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 2.0-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 2.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 6.0-4.0, filter 380 nm.



Figure S3-22. Luminescent data of $[Eu_nSm_{4-n}L3^{RR}_6]$ (prepared by ratio Eu/Sm: 1/1), (2.48 x 10⁻⁶ M in MeCN).(A) Excitation spectrum, $\lambda_{em} = 616$ nm, slits = 2.0-1.0, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 2.5-1.0, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 616$ nm, slits = 6.0-4.0, filter 380 nm.



Figure S3-24. Luminescent data of $[Gd_4(L3^{RR})_6](OTf)_{12}$ (9.05 x 10⁻⁶ M in 1:4 of MeOH/EtOH at 77K).(A) Excitation spectrum, $\lambda_{em} = 466$ nm, slits = 2.0-0.5, filter 380 nm. (b) Emission spectrum, $\lambda_{ex} = 330$ nm, slits = 2.0-0.5, filter 380 nm. (c) Excited state decay curve, $\lambda_{em} = 466$ nm, slits = 2.0-1.0, filter 380 nm.



Figure S3-25. A stack of g_{lum} values of homometallic and heterometallic tetrahedra.

Chapter 4



Figure S4-2. ¹³CNMR of methyl 2-amino-2-methylpropanoate.



Figure S4-3. ¹HNMR of 1-methoxy-2-methyl-1-oxopropan-2-aminium 2,2,2-trifluoroacetate.



Figure S4-4. ¹³CNMR of 1-methoxy-2-methyl-1-oxopropan-2-aminium 2,2,2-trifluoroacetate.



Figure S4-5. ¹HNMR of Boc-Aib-Aib-OMe.



Figure S4-6. ¹³CNMR of Boc-Aib-Aib-OMe.



Figure S4-7. ¹HNMR of Boc-Aib-Aib-OH.



Figure S4-8. ¹³CNMR of Boc-Aib-Aib-OH.



Figure S4-9. ¹HNMR of Boc-Aib-Aib-Aib-OMe.



Figure S4-10. ¹³CNMR of Boc-Aib-Aib-Aib-OMe.



Figure S4-11. ¹HNMR of Boc-(Aib)₄-OMe.



Figure S4-12. ¹³CNMR of Boc-(Aib)₄-OMe.



Figure S4-13. ¹HNMR of Boc-(Aib)₄-OH.



Figure S4-14. ¹³CNMR of Boc-(Aib)₄-OH.



Figure S4-15. ¹HNMR of Boc-Aib-Aib-spy.



Figure S4-16. ¹³CNMR of Boc-Aib-Aib-spy.



Figure S4-17. ¹HNMR of Boc-(Aib)₄-spy.



Figure S4-18. ¹³CNMR of Boc-(Aib)₄-spy.



Figure S4-19. ¹HNMR of di-tert-butyl 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-diyl))bis(carbonyl))dipicolinate.



Figure S4-20. ¹HNMR of di-tert-butyl 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-diyl))bis(carbonyl))dipicolinate.



Figure S4-21. ¹HNMR of 6,6'-(((9,10-dioxo-9,10-dihydroanthracene-2,6-diyl))bis(carbonyl))dipicolinic acid.



Figure S4-22. ¹HNMR of N^2 , N^2 '-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N^6 -((S)-5,5,8,8,11,11,14-heptamethyl-4,7,10,13-tetraoxo-2-phenyl-3,6,9,12-tetraazapentadecan-14-yl)pyridine-2,6-dicarboxamide), **L5**.



Figure S4-23. ¹³CNMR of N²,N^{2'}-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-((S)-5,5,8,8,11,11,14-heptamethyl-4,7,10,13-tetraoxo-2-phenyl-3,6,9,12-tetraazapentadecan-14-yl)pyridine-2,6-dicarboxamide), **L5**.



Figure S4-24. ¹HNMR of N², N^{2'}-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-(2-methyl-1-(((2-methyl-1-oxo-1-(((S)-1-phenylethyl)amino)propan-2-yl)amino)-1-oxopropan-2-yl)pyridine-2,6-dicarboxamide), L6.



Figure S4-25. ¹³CNMR of N²,N^{2'}-(9,10-dioxo-9,10-dihydroanthracene-2,6-diyl)bis(N⁶-(2-methyl-1-((2-methyl-1-oxo-1-(((S)-1-phenylethyl)amino)propan-2-yl)amino)-1oxopropan-2-yl)pyridine-2,6-dicarboxamide), **L6**.



Figure S4-26. ¹HNMR spectrum of H₂N-Ac₆c-OH.



Figure S4-27. ¹³CNMR spectrum of H₂N-Ac₆c-OH.



Figure S4-28. ¹HNMR spectrum of Boc-Ac₆c-OH.



Figure S4-29. ¹³CNMR spectrum of Boc-Ac₆c-OH.



Figure S4-30. ¹HNMR spectrum of Boc-Ac₆c-Ac₆c-OH.



Figure S4-31. ¹³CNMR spectrum of Boc-Ac₆c-Ac₆c-OH.



Figure S4-32. ¹HNMR spectrum of Boc-Ac₆c-rpy.



Figure S4-33. ¹³CNMR spectrum of Boc-Ac₆c-rpy.



Figure S4-34. ¹³CNMR spectrum of Boc-Ac₆c-Ac₆c-rpy.



Figure S4-35. ¹³CNMR spectrum of Boc-Ac₆c-Ac₆c-rpy.



Figure S4-36. ¹HNMR of L7.



Figure S4-37. ¹³CNMR of L7.



Figure S4-38. ¹HNMR of L8.



Figure S4-40. ¹HNMR spectrum of *di-tert*-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.



Figure S4-41. ¹³CNMR spectrum of *di-tert*-butyl ((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.



Figure S4-42. ¹HNMR spectrum of di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.



Figure S4-43. ¹³CNMR spectrum of di-tert-butyl ((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(aza



Figure S4-44. ¹HNMR spectrum of *Di-tert*-butyl (((((((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.



Figure S4-45. ¹HNMR spectrum of 6-(phenylcarbamoyl)picolinic acid.



Figure S4-46. ¹³CNMR spectrum of 6-(phenylcarbamoyl)picolinic acid.



Figure S4-47. ¹HNMR spectrum of 4-(6-(phenylcarbamoyl)picolinamido)benzoic acid.



Figure S4-48. ¹HNMR spectrum of 4-(6-(phenylcarbamoyl)picolinamido)benzoic acid.



Figure S4-50. ¹³CNMR spectrum of L9.


Figure S4-51. ¹HNMR spectrum of Di-*tert*-butyl ((2*S*,2'*S*)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))dicarbamate.



Figure S4-52. ¹³CNMR spectrum of Di-*tert*-butyl ((2*S*,2'*S*)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopentane-1,2-diyl))dicarbamate.



Figure S4-53. ¹HNMR spectrum of Di-tert-butyl ((((2S,2'S)-(((1,4-phenylenebis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(4-methyl-1-oxopropane-1,2-diyl))bis(azanediyl))bis(2-methyl-1-oxopropane-1,2-diyl))dicarbamate.



Figure S4-54. ¹HNMR spectrum of 6-(isopropylcarbamoyl)picolinic acid.



Figure S4-55. ¹³CNMR spectrum of 6-(isopropylcarbamoyl)picolinic acid.



Figure S4-56. ¹HNMR spectrum of 4-(6-(isopropylcarbamoyl)picolinamido)benzoic

acid



Figure S4-57. ¹³CNMR spectrum of 4-(6-(isopropylcarbamoyl)picolinamido)benzoic acid



Figure S4-58. ¹HNMR spectrum of L10.



Figure S4-59. ¹³CNMR spectrum of L10.